

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization  
International Bureau



(43) International Publication Date  
21 November 2002 (21.11.2002)

PCT

(10) International Publication Number  
**WO 02/092801 A2**

(51) International Patent Classification<sup>7</sup>: **C12N 9/00**

**Thomas, Günter** [DE/CH]; Syngenta Crop Protection AG, Schwarzwallee 215, CH-4058 Basel (CH).

(21) International Application Number: PCT/EP02/05363

(22) International Filing Date: 15 May 2002 (15.05.2002)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:  
60/291,149 16 May 2001 (16.05.2001) US

(71) Applicant (for all designated States except US): **SYNGENTA PARTICIPATIONS AG** [CH/CH]; Schwarzwaldallee 215, CH-4058 Basel (CH).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **MOLNAR, Istvan** [HU/US]; Syngenta Biotechnology, Inc., 3054 Cornwallis Road, Research Triangle Park, NC 27709 (US). **LIGON, James, Madison** [US/US]; Syngenta Biotechnology, Inc., 3054 Cornwallis Road, Research Triangle Park, NC 27709 (US). **ZIRKLE, Ross, Eric** [US/US]; Syngenta Biotechnology, Inc., 3054 Cornwallis Road, Research Triangle Park, NC 27709 (US). **HAMMER, Philip, Eugene** [US/US]; 212A Waldo Street, Cary, NC 27511 (US). **HILL, Dwight, Steven** [US/US]; Syngenta Biotechnology, Inc., 3054 Cornwallis Road, Research Triangle Park, NC 27709 (US). **PACHLATKO, Johannes, Paul** [CH/CH]; Syngenta Crop Protection AG, Schwarzwaldallee 215, CH-4058 Basel (CH). **BUCKEL,**

(74) Agent: **BASTIAN, Werner**; c/o Syngenta Participations AG, Intellectual Property, P.O. Box, CH-4002 Basel (CH).

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZM, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

**Published:**

- without international search report and to be republished upon receipt of that report
- with (an) indication(s) in relation to deposited biological material furnished under Rule 13bis separately from the description

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: METHODS AND COMPOSITIONS FOR MAKING EMAMECTIN

(57) Abstract: Disclosed is a family of P450 monooxygenases, each member of which regioselectively oxidizes avermectin to 4"-keto-avermectin. The P450 monooxygenases find use in methods and formulations for making emamectin from avermectin. Also disclosed are methods for purifying the P450 monooxygenases of the invention, binding agents that specifically bind to the P450 monooxygenases of the invention, and genetically engineered cells that express the P450 monooxygenases of the invention. Also disclosed are ferredoxins and ferredoxin reductases that are active with the P450 monooxygenases of the invention.



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## METHODS AND COMPOSITIONS FOR MAKING EMAMECTIN

The invention relates to the field of agrochemicals, and in particular, to insecticides. More specifically, this invention relates to the derivatization of avermectin, particularly for the synthesis of emamectin.

Emamectin is a potent insecticide and controls many pests such as thrips, leafminers, and worm pests including alfalfa caterpillar, beet armyworm, cabbage looper, corn earworm, cutworms, diamondback moth, tobacco budworm, tomato fruitworm, and tomato pinworm. Emamectin (4"-deoxy-4"-epi-N-methylamino avermectin Bl<sub>a</sub>/Bl<sub>b</sub>) is described in U.S. Patent No. 4,874,749 and in Cvetovich, R.J. *et al.*, *J. Organic Chem.* **59**:7704-7708, 1994 (as MK-244).

U.S. Patent No. 5,288,710 describes salts of emamectin that are especially valuable agrochemically. These salts of emamectin are valuable pesticides, especially for combating insects and representatives of the order Acarina. Some pests for which emamectin is useful in combating are listed in European Patent Application EP-A 736,252.

One drawback to the use of emamectin is the difficulty of its synthesis from avermectin. This is due to the first step of the process, which is the most costly and time-consuming step of producing emamectin, in which the 4"-carbinol group of avermectin must be oxidized to a ketone. The oxidation of the 4"-carbinol group is problematic due to the presence of two other hydroxyl groups on the molecule that must be chemically protected before oxidation and deprotected after oxidation. Thus, this first step, significantly increases the overall cost and time of producing emamectin from avermectin.

Because of the efficacy and potency of emamectin as an insecticide, there is a need to develop a cost and time effective method and/or reagent for regioselectively oxidizing the 4"-carbinol group of avermectin to produce 4"-keto-avermectin, which is a necessary intermediate for producing emamectin from avermectin.

The invention provides a novel family of P450 monooxygenases, each member of which is able to regioselectively oxidize the 4"-carbinol group of unprotected avermectin, thereby resulting in a cheap, effective method to produce 4"-keto-avermectin, a necessary intermediate in the production of emamectin. The invention allows elimination of the costly, time-consuming steps of (1) chemically protecting the two other hydroxyl groups on the avermectin

molecule prior to oxidation of the 4''-carbinol group that must be chemically protected before oxidation; and (2) chemically deprotecting these two other hydroxyl groups after oxidation. The invention thus provides reagents and methods for significantly reducing the overall cost of producing emamectin from avermectin.

- Accordingly, in one aspect, the invention provides a purified nucleic acid molecule encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin.
- In a specific embodiment, the invention relates to an purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin, which polypeptide is substantially similar, and preferably has between at least 50%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:2, with each individual number within this range of between 50% and 99% also being part of the invention.
- In a further specific embodiment, the invention relates to an purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin, which polypeptide is immunologically reactive with antibodies raised against a polypeptide of SEQ ID NO:2.
- The invention further provides a purified nucleic acid molecule comprising a nucleotide sequence
  - a) as given in SEQ ID NO:1;
  - b) having substantial similarity to (a);
  - c) capable of hybridizing to (a) or the complement thereof;
  - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, or the complement thereof;
  - e) complementary to (a), (b) or (c);
  - f) which is the reverse complement of (a), (b) or (c), or

- g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin.
- In a specific embodiment, the invention relates to a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin, which polypeptide is substantially similar, and preferably has at least between 60%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95, with each individual number within this range of between 60% and 99% also being part of the invention.
  - In a further specific embodiment, the invention relates to an purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin, which polypeptide is immunologically reactive with antibodies raised against a polypeptide of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
  - The invention further provides a purified nucleic acid molecule comprising a nucleotide sequence
    - a) as given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94;
    - b) having substantial similarity to (a);
    - c) capable of hybridizing to (a) or the complement thereof;
    - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, SEQ ID

NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof;

- e) complementary to (a), (b) or (c);
- f) which is the reverse complement of (a), (b) or (c); or
- g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin.

In certain embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least between 66%, and 99% identical to SEQ ID NO:1, with each individual number within this range of between 66%, and 99% also being part of the invention..

- In certain embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least between 70%, and 99% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94, with each individual number within this range of between 70%, and 99% also being part of the invention..
- In some embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 80% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.
- In certain embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 90% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.

- In certain embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 95% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.
- In some embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence selected from the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, and SEQ ID NO:94.
- In particular embodiments, the nucleic acid molecule is isolated from a *Streptomyces* strain. In certain embodiments, the *Streptomyces* strain is selected from the group consisting of *Streptomyces tubercidicus*, *Streptomyces lydicus*, *Streptomyces platensis*, *Streptomyces chattanoogensis*, *Streptomyces kasugaensis*, and *Streptomyces rimosus* and *Streptomyces albofaciens*.
- In some embodiments of this aspect, the nucleic acid molecule further comprises a nucleic acid sequence encoding a tag which is linked to the P450 monooxygenase via a covalent bond. In certain embodiments, the tag is selected from the group consisting of a His tag, a GST tag, an HA tag, a HSV tag, a Myc-tag, and VSV-G-Tag.
- In another aspect, the invention provides a purified polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin.
- In some embodiments, the polypeptide comprises or consists essentially of an amino acid sequence that is encoded by a nucleic acid molecule
  - a) as given in SEQ ID NO:1 or the complement thereof;
  - b) having substantial similarity to (a);
  - c) capable of hybridizing to (a) or the complement thereof;
  - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, or the complement thereof;
  - e) complementary to (a), (b) or (c);

- f) which is the reverse complement of (a), (b) or (c); or.
- g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin.

In some embodiments, the polypeptide comprises or consists essentially of an amino acid sequence that is between at least 50%, and 99% identical to SEQ ID NO:2, with each individual number within this range of between 50% and 99% also being part of the invention..

- In some embodiments, the polypeptide comprises or consists essentially of an amino acid sequence that is encoded by a nucleic acid molecule
  - a) as given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof;
  - b) having substantial similarity to (a);
  - c) capable of hybridizing to (a) or the complement thereof;
  - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof, or the complement thereof;
  - e) complementary to (a), (b) or (c);
  - f) which is the reverse complement of (a), (b) or (c); or
  - g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin.
- In some embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is between at least 60%, and 99% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24,

SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95, with each individual number within this range of between 60% and 99% also being part of the invention..

- In certain embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 70% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In some embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 80% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In some embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 90% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In certain embodiments, the P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 95% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In some embodiments of this aspect of the invention, the P450 monooxygenase comprises or consists essentially of an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, and SEQ ID NO:95.
- In certain embodiments, the polypeptide according to the invention exhibiting an enzymatic activity of a P450 monooxygenase further comprises a tag. In some



embodiments, the tag is selected from the group consisting of a His tag, a GST tag, an HA tag, a HSV tag, a Myc-tag, and VSV-G-Tag.

- In another aspect, the invention provides a binding agent that specifically binds to a polypeptide according to the invention exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin. In some embodiments, the binding agent is an antibody. In certain embodiments, the antibody is a polyclonal antibody or a monoclonal antibody.
- In yet another aspect, the invention provides a family of P450 monooxygenase polypeptides, wherein each member of the family regioselectively oxidizes avermectin to 4''-keto-avermectin.
- In certain embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is between at least 50%, and 99% identical to SEQ ID NO:2, with each individual number within this range of between 50% and 99% also being part of the invention..
- In certain embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is between at least 60%, and 99% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95, with each individual number within this range of between 60% and 99% also being part of the invention..
- In some embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is at least 70% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In certain embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is at least 80% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95. In

some embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is at least 90% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.

- In certain embodiments, each member of the family comprises or consists essentially of an amino acid sequence that is at least 95% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- In some embodiments of this aspect of the invention, each member of the family comprises or consists essentially of an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, and SEQ ID NO:95.
- In still another aspect, the invention provides a cell genetically engineered to comprise a nucleic acid molecule encoding a polypeptide which exhibits an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin.
- In some embodiments, the nucleic acid molecule is positioned for expression in the cell. In certain embodiments, the cell further comprises a nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin protein.
- In some embodiments, the cell further comprises a nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase protein.
- In certain embodiments, the cell is a genetically engineered *Streptomyces* strain. In certain embodiments, the cell is a genetically engineered *Streptomyces lividans* strain. In particular embodiments, the genetically engineered *Streptomyces lividans* strain has NRRL Designation No. B-30478. In some embodiments, the cell is a genetically engineered *Pseudomonas* strain. In some embodiments, the cell is a genetically engineered

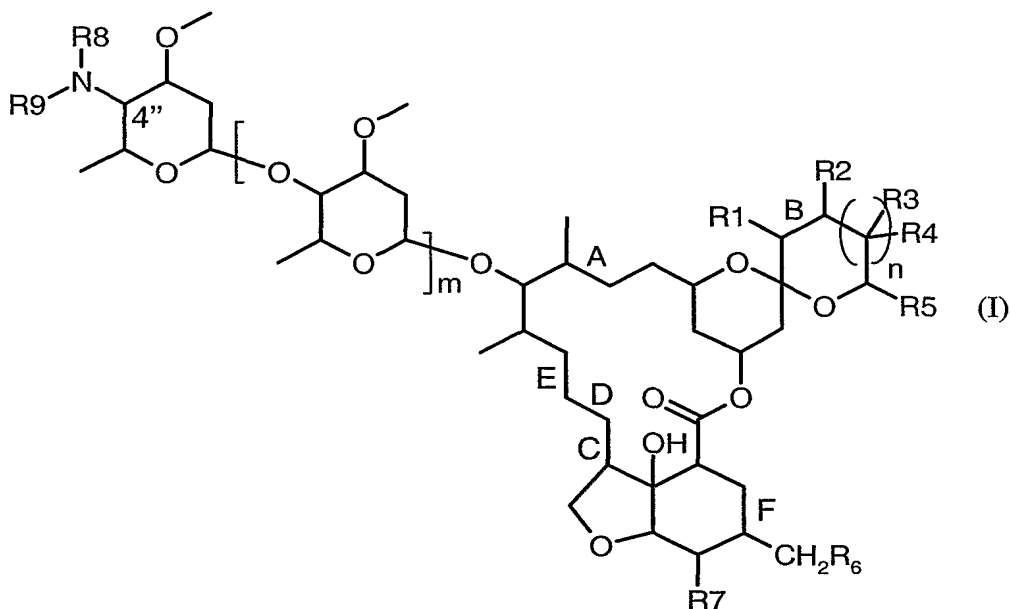
*Pseudomonas putida* strain. In certain embodiments, the genetically engineered *Pseudomonas putida* strain has NRRL Designation No. B-30479. In some embodiments, the cell is a genetically engineered *Escherichia coli* strain.

- In another aspect, the invention provides a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin.
- In a specific embodiment, the invention relates to a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits the enzymatic activity of a ferredoxin, which polypeptide is substantially similar, and preferably has between at least 80%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:36 or SEQ ID NO: 38, with each individual number within this range of between 80% and 99% also being part of the invention.
- In still a further specific embodiment, the invention relates to a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits the enzymatic activity of a ferredoxin, which polypeptide is immunologically reactive with antibodies raised against a polypeptide of SEQ ID NO: 36 or SEQ ID NO: 38.
- The invention further provides a purified nucleic acid molecule comprising a nucleotide sequence
  - a) as given in SEQ ID NO:35 or SEQ ID NO: 37;
  - b) having substantial similarity to (a);
  - c) capable of hybridizing to (a) or the complement thereof;
  - d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO: 35 or SEQ ID NO: 37, or the complement thereof;
  - e) complementary to (a), (b) or (c);
  - f) which is the reverse complement of (a), (b) or (c); or
  - g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a ferredoxin and regioselectively oxidizes avermectin to 4"-keto-avermectin.

- In certain embodiments, the nucleic acid molecule encoding a ferredoxin of the invention comprises or consists essentially of a nucleic acid sequence that is at least 81% identical to SEQ ID NO:35 or SEQ ID NO:37. In some embodiments, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:35 or SEQ ID NO:37. In certain embodiments, the nucleic acid molecule encoding a ferredoxin of the invention comprises or consists essentially of the nucleic acid sequence of SEQ ID NO:35 or SEQ ID NO:37.
- In yet another aspect, the invention provides a purified ferredoxin protein, wherein the ferredoxin protein is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin. In certain embodiments, the ferredoxin of the invention comprises or consists essentially of an amino acid sequence that is at least 80% identical to SEQ ID NO:36 or SEQ ID NO:38. In some embodiments, the nucleic acid molecule comprises or consists essentially of an amino acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:36 or SEQ ID NO:38.
- In particular embodiments, the ferredoxin of the invention comprises or consists essentially of the amino acid sequence of SEQ ID NO:36 or SEQ ID NO:38.
- In another aspect, the invention provides a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin.
- In certain embodiments, the nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase comprises or consists essentially of the nucleic acid sequence of SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:104.
- In yet another aspect, the invention provides a purified polypeptide exhibiting an enzymatic activity of a ferredoxin reductase protein, wherein the said polypeptide is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin. In certain embodiments, the

polypeptide of the invention comprises or consists essentially of the amino acid sequence of SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, or SEQ ID NO:105.

- In another aspect, the invention provides a process for the preparation a compound of the formula



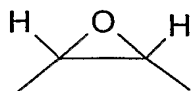
in which

$R_1$ - $R_9$  represent, independently of each other hydrogen or a substituent;

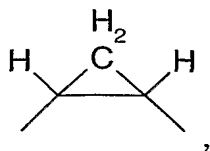
$m$  is 0, 1 or 2;

$n$  is 0, 1, 2 or 3; and

the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



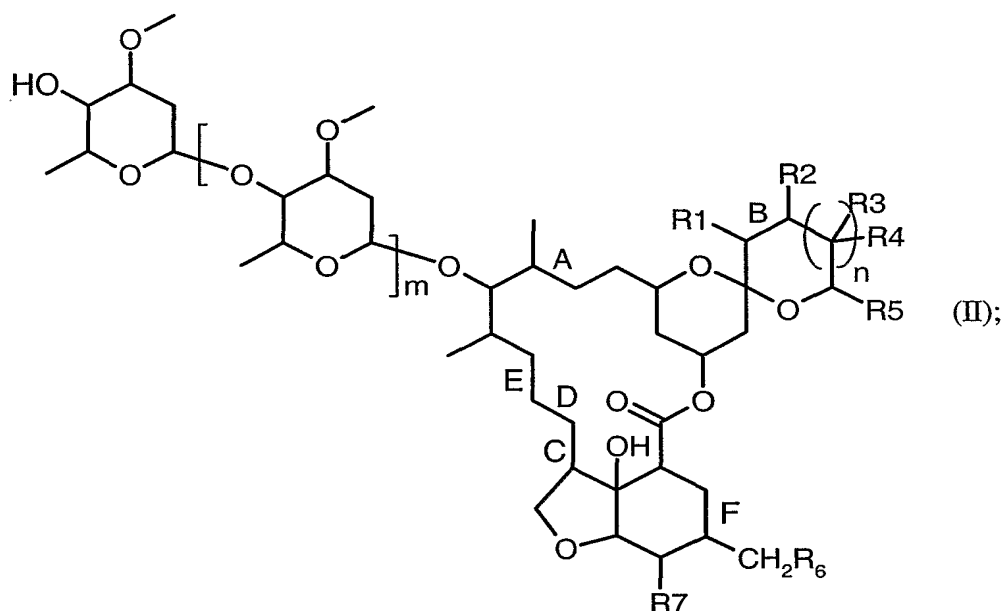
, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,

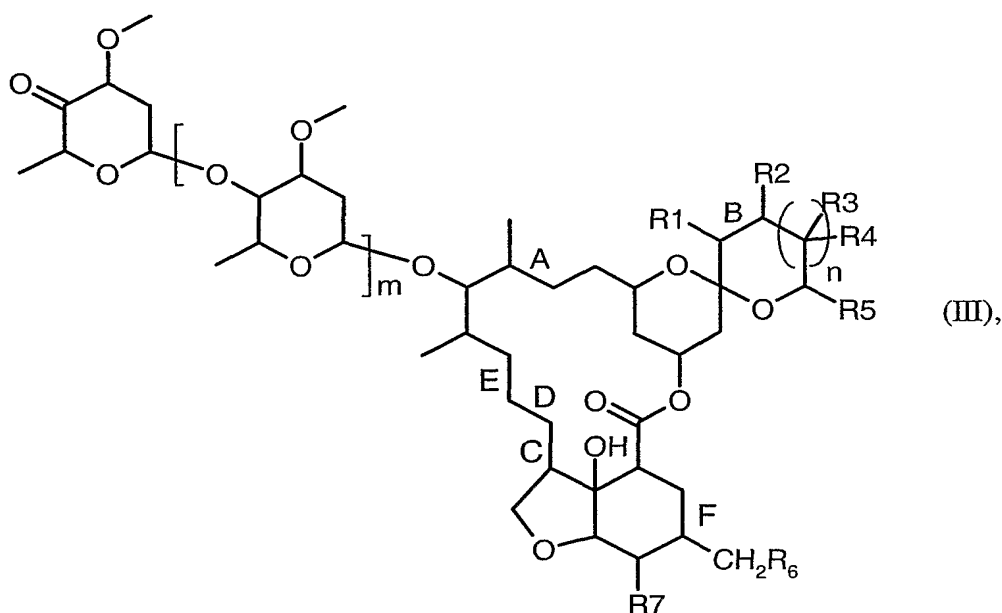
which process comprises

1) bringing a compound of the formula



wherein

R<sub>1</sub>-R<sub>7</sub>, m, n, A, B, C, D, E and F have the same meanings as given for formula (I) above, into contact with a polypeptide according to the invention that is capable of regioselectively oxidising the alcohol at position 4" in order to form a compound of the formula



in which  $R_1$ ,  $R_2$ ,  $R_3$ ,  $R_4$ ,  $R_5$ ,  $R_6$ ,  $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the meanings given for formula (I); and

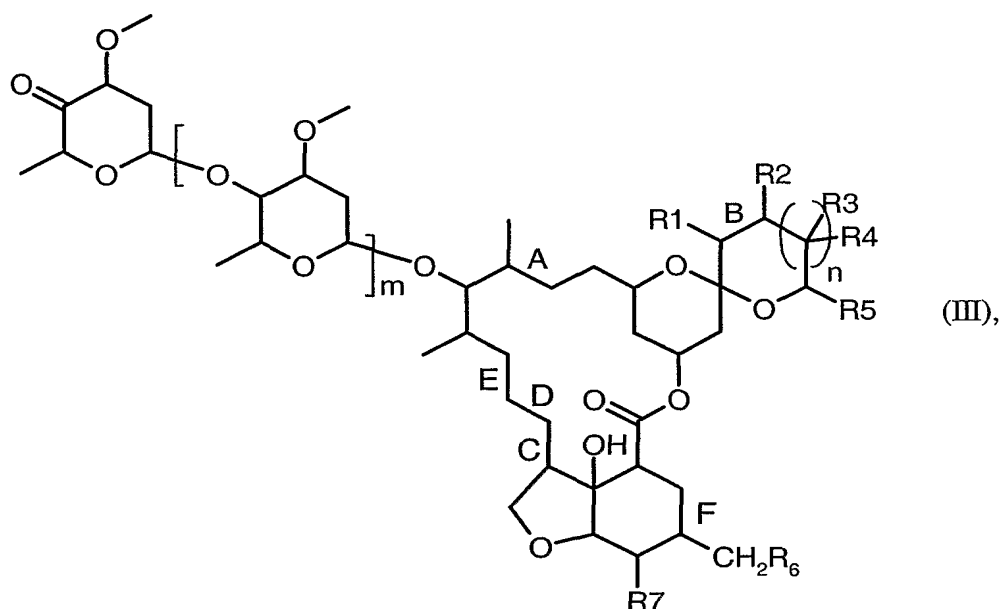
2) reacting the compound of the formula (III) with an amine of the formula  $\text{HN}(R_8)R_9$ , wherein  $R_8$  and  $R_9$  have the same meanings as given for formula (I), and which is known, in the presence of a reducing agent;

and, in each case, if desired, converting a compound of formula (I) obtainable in accordance with the process or by another method, or an E/Z isomer or tautomer thereof, in each case in free form or in salt form, into a different compound of formula (I) or an E/Z isomer or tautomer thereof, in each case in free form or in salt form, separating a mixture of E/Z isomers obtainable in accordance with the process and isolating the desired isomer, and/or converting a free compound of formula (I) obtainable in accordance with the process or by another method, or an E/Z isomer or tautomer thereof, into a salt or converting a salt, obtainable in accordance with the process or by another method, of a compound of formula (I) or of an E/Z isomer or tautomer thereof into the free compound of formula (I) or an E/Z isomer or tautomer thereof or into a different salt.

- In some embodiments, the compound of formula (II) is further brought into contact with a polypeptide according to the invention exhibiting an enzymatic activity of a

ferredoxin. In certain embodiments, the compound of formula (II) is further brought into contact with a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase. In some embodiments, the compound of formula (II) is further brought into contact with a reducing agent (*e.g.*, NADH or NADPH).

- In still a further embodiment, the invention provides a process for the preparation of a compound of the formula

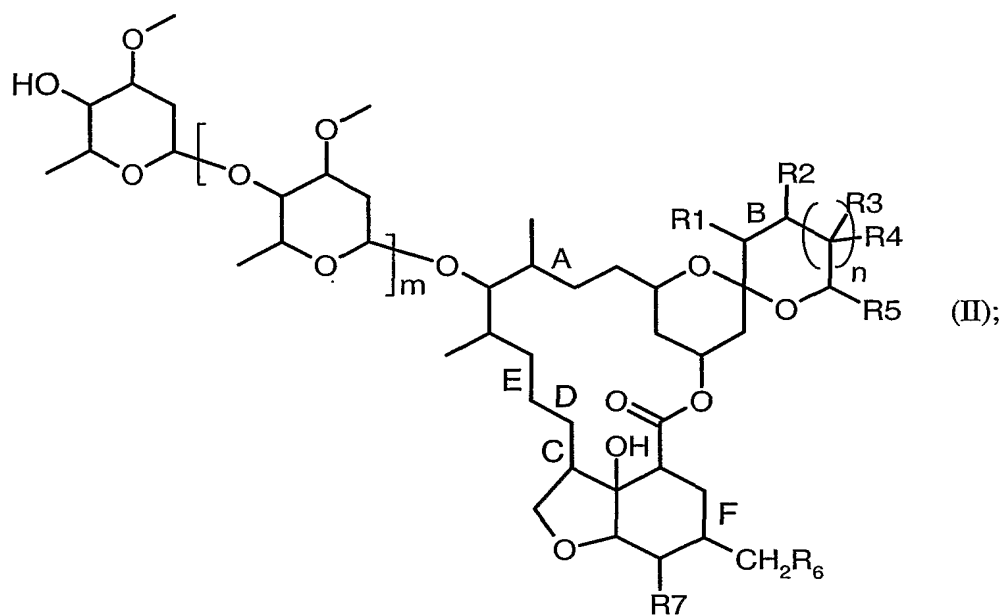


in which  $R_1$ ,  $R_2$ ,  $R_3$ ,  $R_4$ ,  $R_5$ ,  $R_6$ ,  $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the meanings given for formula (I) of claim 1,

which process comprises

- 1) bringing a compound of the formula





wherein

$R_1$ - $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the same meanings as given for formula (I) above, into contact with a polypeptide according to the invention that is capable of regioselectively oxidising the alcohol at position 4", maintaining said contact for a time sufficient for the oxidation reaction to occur and isolating and purifying the compound of formula (II).

- In yet another embodiment, the invention provides a process according to the invention for the preparation of a compound of the formula (I), in which

$n$  is 1;

$m$  is 1;

$A$  is a double bond;

$B$  is single bond or a double bond,

$C$  is a double bond,

$D$  is a single bond,

$E$  is a double bond,

$F$  is a double bond; or a single bond and a epoxy bridge; or a single bond and a methylene bridge;

$R_1$ ,  $R_2$  and  $R_3$  are H;

R<sub>4</sub> is methyl;

R<sub>5</sub> is C<sub>1</sub>-C<sub>10</sub>-alkyl, C<sub>3</sub>-C<sub>8</sub>-cycloalkyl or C<sub>2</sub>-C<sub>10</sub>-alkenyl;

R<sub>6</sub> is H;

R<sub>7</sub> is OH;

R<sub>8</sub> and R<sub>9</sub> are independently of each other H; C<sub>1</sub>-C<sub>10</sub>-alkyl or C<sub>1</sub>-C<sub>10</sub>-acyl; or together form -(CH<sub>2</sub>)<sub>q</sub>-; and

q is 4, 5 or 6.

- In still another embodiment, the invention provides a process according to the invention for the preparation of a compound of the formula (I), in which
  - n is 1;
  - m is 1;
  - A, B, C, E and F are double bonds;
  - D is a single bond;
  - R<sub>1</sub>, R<sub>2</sub>, and R<sub>3</sub> are H;
  - R<sub>4</sub> is methyl;
  - R<sub>5</sub> is s-butyl or isopropyl;
  - R<sub>6</sub> is H;
  - R<sub>7</sub> is OH;
  - R<sub>8</sub> is methyl
  - R<sub>9</sub> is H.
- In still another embodiment, the invention provides a process according to the invention for the preparation of 4''-deoxy-4''-N-methylamino avermectin B<sub>1a</sub>/B<sub>1b</sub> benzoate salt.
- In another aspect, the invention provides a method for making emamectin. The method comprises adding a polypeptide according to the invention exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin to a reaction mixture comprising avermectin and incubating the reaction mixture under conditions that allow the polypeptide to regioselectively oxidize avermectin to 4''-keto-avermectin. In some embodiments, the reaction mixture further comprises a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin. In

certain embodiments, the reaction mixture further comprises a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase. In some embodiments, the reaction mixture further comprises a reducing agent (*e.g.*, NADH or NADPH).

- In still another aspect, the invention provides a formulation for making a compound of formula (I) comprising a polypeptide according to the invention exhibiting a P450 monooxygenase activity that is capable of regioselectively oxidising the alcohol at position 4" in order to form a compound of formula (II). In some embodiments, the formulation further comprises a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin (*e.g.*, a ferredoxin from cell or strain from which the P450 monooxygenase was isolated or derived).
- In still another aspect, the invention provides a formulation for making emamectin comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. In some embodiments, the formulation further comprises a ferredoxin (*e.g.*, a ferredoxin from cell or strain from which the P450 monooxygenase was isolated or derived).
- In certain embodiments, the formulation further comprises a polypeptide according to the invention exhibiting an enzymatic activity of a ferredoxin reductase (*e.g.*, a ferredoxin from cell or strain from which the P450 monooxygenase was isolated or derived). In some embodiments, the formulation further comprises a reducing agent (*e.g.*, NADH or NADPH).

#### Brief Description of the Drawings

Figure 1 is a diagrammatic representation showing a map of plasmid pTBBKA. Recognition sites by the indicated restriction endonucleases are shown, along with the location of the site in the nucleotide sequence of the plasmid. Also shown are genes (*e.g.*, kanamycin resistance "KanR"), and other functional aspects (*e.g.*, Tip promoter) contained in the plasmid.

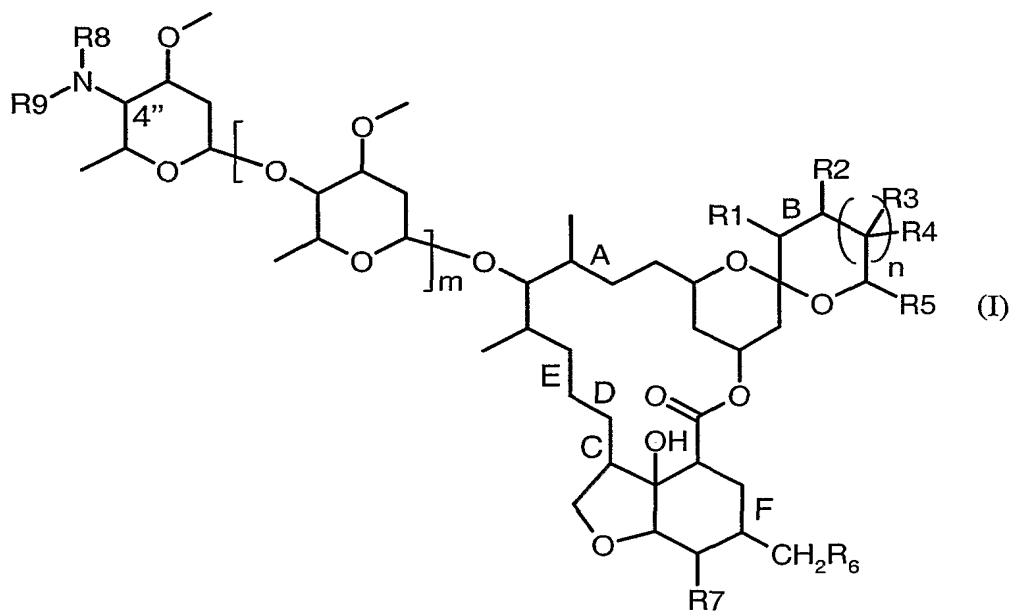
Figure 2 is a diagrammatic representation showing a map of plasmid pTUA1A. Recognition sites by the indicated restriction endonucleases are shown, along with the

location of the site in the nucleotide sequence of the plasmid. Also shown are genes (*e.g.*, ampicillin resistance "AmpR") and other functional aspects (*e.g.*, Tip promoter) contained in the plasmid.

Figure 3 is a diagrammatic representation showing a map of plasmid pRK-*ema1/fd233*. This plasmid was derived by ligating a BglII fragment containing the *ema1* and *fd233* genes organized on a single transcriptional unit into the BglII site of the broad host-range plasmid pRK290. The *ema1/fd233* genes are expressed by the tac promoter (Ptac), and they are followed by the tac terminator (Ttac). Restriction endonuclease recognition sites shown are BglII "B"; EcoRI "E"; PacI "Pc"; PmeI "Pm"; and SalI "S."

The present invention provides a family of polypeptides which exhibit an enzymatic activity of a P450 monooxygenases and are capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II) such as avermectin in order to produce a compound of the formula (III), but especially 4"-keto-avermectin.

More particularly, the family of polypeptides according to the invention may be used in a process for the preparation a compound of the formula



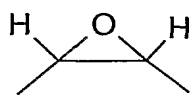
in which

R<sub>1</sub>-R<sub>9</sub> represent, independently of each other hydrogen or a substituent;

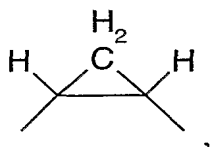
m is 0, 1 or 2;

n is 0, 1, 2 or 3; and

the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



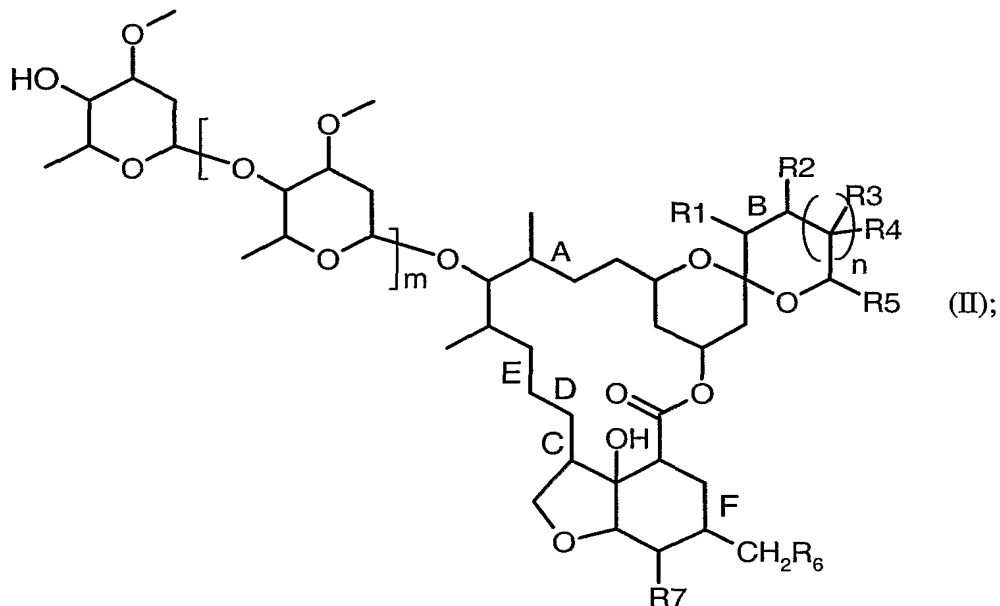
, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,

which process comprises

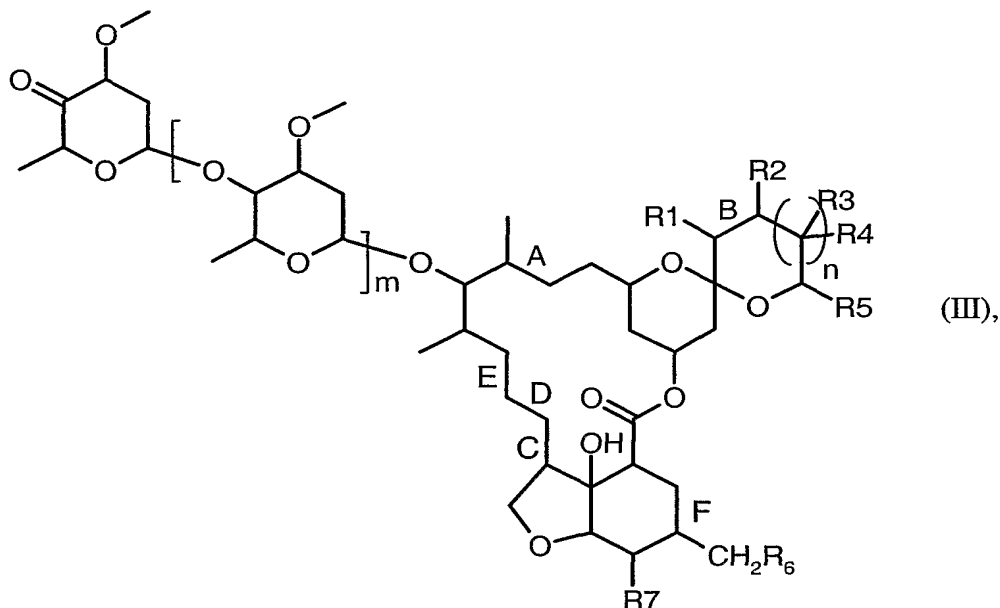
1) bringing a compound of the formula



wherein

R<sub>1</sub>-R<sub>7</sub>, m, n, A, B, C, D, E and F have the same meanings as given for formula (I) above,

into contact with a polypeptide according to the invention which exhibits an enzymatic activity of a P450 monooxygenases and is capable of regioselectively oxidizing the alcohol at position 4" of formular (II) in order to produce a compound of the formula (III)



in which  $R_1$ ,  $R_2$ ,  $R_3$ ,  $R_4$ ,  $R_5$ ,  $R_6$ ,  $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the meanings given for formula (I); and

2) reacting the compound of the formula (III) with an amine of the formula  $HN(R_8)R_9$ , wherein  $R_8$  and  $R_9$  have the same meanings as given for formula (I), and which is known, in the presence of a reducing agent;

and, in each case, if desired, converting a compound of formula (I) obtainable in accordance with the process or by another method, or an *E/Z* isomer or tautomer thereof, in each case in free form or in salt form, into a different compound of formula (I) or an *E/Z* isomer or tautomer thereof, in each case in free form or in salt form, separating a mixture of *E/Z* isomers obtainable in accordance with the process and isolating the desired isomer, and/or converting a free compound of formula (I) obtainable in accordance with the process or by another method, or an *E/Z* isomer or tautomer thereof, into a salt or converting a salt, obtainable in accordance with the process or by another method, of a compound of formula (I) or of an *E/Z* isomer or tautomer thereof into the free compound of formula (I) or an *E/Z* isomer or tautomer thereof or into a different salt.

Methods of synthesis for the compounds of formula (I) are described in the literature. It has been found, however, that the processes known in the literature cause considerable problems during production basically on account of the low yields and the tedious procedures which have to be used. Accordingly, the known processes are not satisfactory in that respect, giving rise to the need to make available improved preparation processes for those compounds.

The compounds (I), (II) and (III) may be in the form of tautomers. Accordingly, hereinbefore and hereinafter, where appropriate the compounds (I), (II) and (III) are to be understood to include corresponding tautomers, even if the latter are not specifically mentioned in each case.

The compounds (I), (II) and (III) are capable of forming acid addition salts. Those salts are formed, for example, with strong inorganic acids, such as mineral acids, for example perchloric acid, sulfuric acid, nitric acid, nitrous acid, a phosphoric acid or a hydrohalic acid, with strong organic carboxylic acids, such as unsubstituted or substituted, for example halo-substituted, C<sub>1</sub>-C<sub>4</sub>alkanecarboxylic acids, for example acetic acid, saturated or unsaturated dicarboxylic acids, for example oxalic, malonic, succinic, maleic, fumaric or phthalic acid, hydroxycarboxylic acids, for example ascorbic, lactic, malic, tartaric or citric acid, or benzoic acid, or with organic sulfonic acids, such as unsubstituted or substituted, for example halo-substituted, C<sub>1</sub>-C<sub>4</sub>alkane- or aryl-sulfonic acids, for example methane- or p-toluene-sulfonic acid. Furthermore, compounds of formula (I), (II) and (III) having at least one acidic group are capable of forming salts with bases. Suitable salts with bases are, for example, metal salts, such as alkali metal or alkaline earth metal salts, for example sodium, potassium or magnesium salts, or salts with ammonia or an organic amine, such as morpholine, piperidine, pyrrolidine, a mono-, di- or tri-lower alkylamine, for example ethyl-, diethyl-, triethyl- or dimethyl-propyl-amine, or a mono-, di- or tri-hydroxy-lower alkylamine, for example mono-, di- or tri-ethanolamine. In addition, corresponding internal salts may also be formed. Preference is given within the scope of the invention to agrochemically advantageous salts. In view of the close relationship between the compounds of formula (I), (II) and (III) in free form and in the form of their salts, any reference hereinbefore or hereinafter to the free compounds of formula (I), (II) and (III) or to their respective salts is to be understood as including also the corresponding salts or the free compounds of formula (I), (II) and (III), where appropriate and

expedient. The same applies in the case of tautomers of compounds of formula (I), (II) and (III) and the salts thereof. The free form is generally preferred in each case.

Preferred within the scope of this invention is a process for the preparation of compounds of the formula (I), in which

n is 1;

m is 1;

A is a double bond;

B is single bond or a double bond,

C is a double bond,

D is a single bond,

E is a double bond,

F is a double bond; or a single bond and an epoxy bridge; or a single bond and a methylene bridge;

R<sub>1</sub>, R<sub>2</sub> and R<sub>3</sub> are H;

R<sub>4</sub> is methyl;

R<sub>5</sub> is C<sub>1</sub>-C<sub>10</sub>-alkyl, C<sub>3</sub>-C<sub>8</sub>-cycloalkyl or C<sub>2</sub>-C<sub>10</sub>-alkenyl;

R<sub>6</sub> is H;

R<sub>7</sub> is OH;

R<sub>8</sub> and R<sub>9</sub> are independently of each other H; C<sub>1</sub>-C<sub>10</sub>-alkyl or C<sub>1</sub>-C<sub>10</sub>-acyl; or together form -(CH<sub>2</sub>)<sub>q</sub>-; and

q is 4, 5 or 6.

Especially preferred within the scope of this invention is a process for the preparation of a compound of the formula (I) in which

n is 1;

m is 1;

A, B, C, E and F are double bonds;

D is a single bond;

R<sub>1</sub>, R<sub>2</sub>, and R<sub>3</sub> are H;

R<sub>4</sub> is methyl;

R<sub>5</sub> is s-butyl or isopropyl;



R<sub>6</sub> is H;

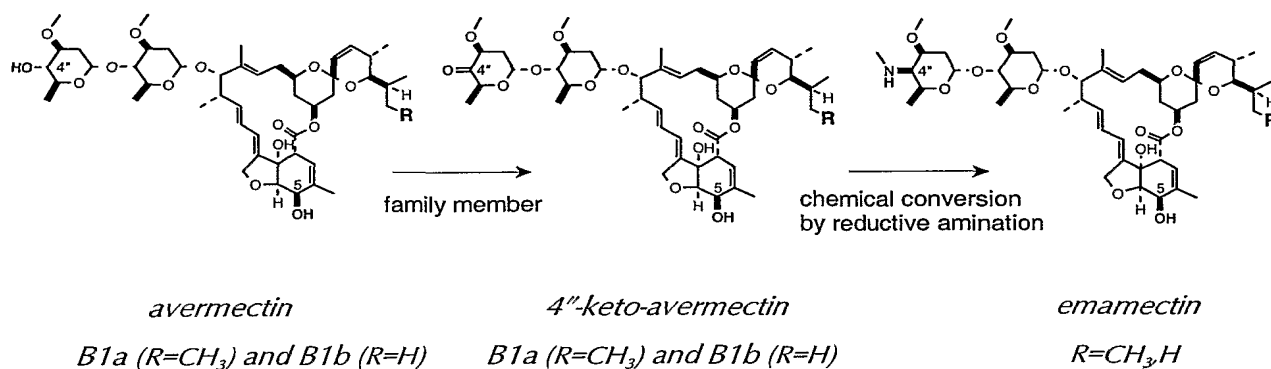
R<sub>7</sub> is OH;

R<sub>8</sub> is methyl

R<sub>9</sub> is H.

Very especially preferred is a process for the preparation of emamectin, more particularly the benzoate salt of emamectin. Emamectin is a mixture of 4''-deoxy-4''-N-methylamino avermectin B<sub>1a</sub>/B<sub>1b</sub> and is described in US-P-4,487,749 and as MK-244 in *Journal of Organic Chemistry*, Vol. 59 (1994), 7704-7708. Salts of emamectin that are especially valuable agrochemically are described in US-P-5,288,710. Each member of this family of peptides exhibiting an enzymatic activity of a P450 monooxygenases as described hereinbefore is able to oxidize unprotected avermectin regioselectively at position 4'', thus opening a new and more economical route for the production of emamectin.

The family members each catalyze the following reaction:



Accordingly, the invention provides a purified nucleic acid molecule encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and is capable of regioselectively oxidizing the alcohol at position 4'' of a compound of formula (II) such as avermectin in order to produce a compound of formula (III), but especially 4''-keto-avermectin.

In particular, the invention provides a purified nucleic acid molecule encoding a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin. A "nucleic

acid molecule” refers to single-stranded or double-stranded polynucleotides, such as deoxyribonucleic acid (DNA), ribonucleic acid (RNA), or analogs of either DNA or RNA.

The invention also provides a purified polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II) such as avermectin in order to produce a compound of formula (III), but especially 4"-keto-avermectin.

In particular, the invention also provides a purified P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin.

As used herein, by “purified” is meant a nucleic acid molecule or polypeptide (*e.g.*, an enzyme or antibody) that has been separated from components which naturally accompany it. An example of such a nucleotide sequence or segment of interest “purified” from a source, would be nucleotide sequence or segment that is excised or removed from said source by chemical means, *e.g.*, by the use of restriction endonucleases, so that it can be further manipulated, *e.g.*, amplified, for use in the invention, by the methodology of genetic engineering. Such a nucleotide sequence or segment is commonly referred to as “recombinant.” In one specific aspect, the purified nucleic acid molecule may be separated from nucleotide sequences, such as promoter or enhancer sequences, that flank the nucleic acid molecule as it naturally occurs in the chromosome.

In the case of a protein or a polypeptide, the purified protein and polypeptide, respectively, is separated from components, such as other proteins or fragments of cell membrane, that accompany it in the cell. Of course, those of ordinary skill in molecular biology will understand that water, buffers, and other small molecules may additionally be present in a purified nucleic acid molecule or purified protein preparation. A purified nucleic acid molecule or purified polypeptide (*e.g.*, enzyme) of the invention that is at least 95% by weight, or at least 98% by weight, or at least 99% by weight, or 100% by weight free of components which naturally accompany the nucleic acid molecule or polypeptide.

According to the invention, a purified nucleic acid molecule may be generated, for example, by excising the nucleic acid molecule from the chromosome. It may then be ligated into an expression plasmid. Other methods for generating a purified nucleic acid molecule encoding a P450 monooxygenase of the invention are available and include, without limitation, artificial synthesis of the nucleic acid molecule on a nucleic acid synthesizer.

Similarly, a purified P450 monooxygenase of the invention may be generated, for example, by recombinant expression of a nucleic acid molecule encoding the P450 monooxygenase in a cell in which the P450 monooxygenase does not naturally occur. Of course, other methods for obtaining a purified P450 monooxygenase of the invention include, without limitation, artificial synthesis of the P450 monooxygenase on a polypeptide synthesizer and isolation of the P450 monooxygenase from a cell in which it naturally occurs using, *e.g.*, an antibody that specifically binds the P450 monooxygenase.

Biotransformations of secondary alcohols to ketones by *Streptomyces* bacteria are known to be catalyzed by dehydrogenases or oxidases. However, prior to the present discovery of the cytochrome P450 monooxygenase from *Streptomyces tubercidicus* strain R-922 responsible for the regioselective oxidation of avermectin to 4''-keto-avermectin, no experimental data of another cytochrome P450 monooxygenase from *Streptomyces* to oxidize a secondary alcohol to a ketone had been reported.

According to some embodiments of the invention, the nucleic acid molecule and/or the polypeptide encoded by the nucleic acid molecule are isolated from a *Streptomyces* strain. Thus, the nucleic acid molecule (or polypeptide encoded thereby) may be isolated from, without limitation, *Streptomyces tubercidicus*, *Streptomyces lydicus*, *Streptomyces platensis*, *Streptomyces chattanoogensis*, *Streptomyces kasugaensis*, *Streptomyces rimosus*, and *Streptomyces albobaciens*.

As mentioned above and described in more detail below, an entire family of polypeptides exhibiting an enzymatic activity of P450 monooxygenases capable of regioselectively oxidizing avermectin to 4''-keto-avermectin are provided herein. All of these family members are related by at least 60% identity at the amino acid level. A useful nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase comprises or consists essentially of a nucleic acid sequence that is at least 70% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94. In certain embodiments, the nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase

comprises or consists essentially of a nucleic acid sequence that is at least 80% identical; or at least 85% identical; or at least 90% identical; or at least 95% identical; or at least 98% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.

Similarly, the invention provides a purified polypeptide exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin which, in some embodiments, comprises or consists essentially of an amino acid sequence that is at least 60% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95. In certain embodiments, the purified polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase comprises or consists essentially of an amino acid sequence that is at least 70% identical; or at least 80% identical; or at least 90% identical; or at least 95% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.

In some embodiments, the nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase comprises or consists essentially of the nucleic acid sequence of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94. Similarly, the purified polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase, in some embodiments, comprises or consists essentially of the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.

To describe the sequence relationships between two or more nucleic acids or polynucleotides the following terms are used: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

(a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full length cDNA or gene sequence, or the complete cDNA or gene sequence.

(b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent identity between any two sequences can be accomplished using a mathematical algorithm. Preferred, non-limiting examples of such mathematical algorithms are the algorithm of Myers and Miller, 1988; the local homology algorithm of Smith et al. 1981; the homology alignment algorithm of Needleman and Wunsch 1970; the search-for-similarity-method of Pearson and Lipman 1988; the algorithm of Karlin and Altschul, 1990, modified as in Karlin and Altschul, 1993.

Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Such implementations include, but are not limited to: CLUSTAL in the PC/Gene program (available from Intelligenetics, Mountain View, California); the ALIGN program (Version 2.0) and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Version 8 (available from Genetics Computer Group (GCG), 575 Science Drive, Madison, Wisconsin, USA). Alignments using these programs can be performed using the default parameters. The

CLUSTAL program is well described by Higgins et al. 1988; Higgins et al. 1989; Corpet et al. 1988; Huang et al. 1992; and Pearson et al. 1994. The ALIGN program is based on the algorithm of Myers and Miller, *supra*. The BLAST programs of Altschul et al., 1990, are based on the algorithm of Karlin and Altschul *supra*.

Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length  $W$  in the query sequence, which either match or satisfy some positive-valued threshold score  $T$  when aligned with a word of the same length in a database sequence.  $T$  is referred to as the neighborhood word score threshold (Altschul et al., 1990). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters  $M$  (reward score for a pair of matching residues; always  $> 0$ ) and  $N$  (penalty score for mismatching residues; always  $< 0$ ). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when the cumulative alignment score falls off by the quantity  $X$  from its maximum achieved value, the cumulative score goes to zero or below due to the accumulation of one or more negative-scoring residue alignments, or the end of either sequence is reached.

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul (1993). One measure of similarity provided by the BLAST algorithm is the smallest sum probability ( $P(N)$ ), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a test nucleic acid sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid sequence to the reference nucleic acid sequence is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

To obtain gapped alignments for comparison purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul et al. 1997. Alternatively, PSI-BLAST (in BLAST 2.0) can be used to perform an iterated search that detects distant relationships

between molecules. See Altschul et al., *supra*. When utilizing BLAST, Gapped BLAST, PSI-BLAST, the default parameters of the respective programs (e.g. BLASTN for nucleotide sequences, BLASTX for proteins) can be used. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, 1989). See <http://www.ncbi.nlm.nih.gov>. Alignment may also be performed manually by inspection. For purposes of the present invention, comparison of nucleotide sequences for determination of percent sequence identity to the nucleotide sequences disclosed herein is preferably made using the BlastN program (version 1.4.7 or later) with its default parameters or any equivalent program. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide or amino acid residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by the preferred program.

(c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences makes reference to the residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g., charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences that differ by such conservative substitutions are said to have "sequence similarity" or "similarity." Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero

and 1. The scoring of conservative substitutions is calculated, e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California).

(d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

(e)(i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, or 79%, preferably at least 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more preferably at least 90%, 91%, 92%, 93%, or 94%, and most preferably at least 95%, 96%, 97%, 98%, or 99% sequence identity, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 70%, more preferably at least 80%, 90%, and most preferably at least 95%.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions (see below). Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point ( $T_m$ ) for the specific sequence at a defined ionic strength and pH. However, stringent conditions encompass temperatures in the range of about 1°C to about 20°C, depending upon the desired degree of stringency as otherwise qualified herein. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two



nucleic acid sequences are substantially identical is when the polypeptide encoded by the first nucleic acid is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

(e)(ii) The term "substantial identity" in the context of a polypeptide indicates that a polypeptide comprises a sequence with at least 50%, 60%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, or 79%, preferably 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, or 89%, more preferably at least 90%, 91%, 92%, 93%, or 94%, or even more preferably, 95%, 96%, 97%, 98% or 99%, sequence identity to the reference sequence over a specified comparison window. Preferably, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch (1970). An indication that two polypeptide sequences are substantially identical is that one polypeptide is immunologically reactive with antibodies raised against the second polypeptide. Thus, a polypeptide is substantially identical to a second polypeptide, for example, where the two peptides differ only by a conservative substitution.

For sequence comparison, typically one sequence acts as a reference sequence to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are input into a computer, subsequence coordinates are designated if necessary, and sequence algorithm program parameters are designated. The sequence comparison algorithm then calculates the percent sequence identity for the test sequence(s) relative to the reference sequence, based on the designated program parameters.

As noted above, another indication that two nucleic acid sequences are substantially identical is that the two molecules hybridize to each other under stringent conditions. The phrase "hybridizing specifically to" refers to the binding, duplexing, or hybridizing of a molecule only to a particular nucleotide sequence under stringent conditions when that sequence is present in a complex mixture (e.g., total cellular) DNA or RNA. "Bind(s) substantially" refers to complementary hybridization between a probe nucleic acid and a target nucleic acid and embraces minor mismatches that can be accommodated by reducing the stringency of the hybridization media to achieve the desired detection of the target nucleic acid sequence.

"Stringent hybridization conditions" and "stringent hybridization wash conditions" in the context of nucleic acid hybridization experiments such as Southern and Northern

hybridization are sequence dependent, and are different under different environmental parameters. The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the  $T_m$  can be approximated from the equation of Meinkoth and Wahl, 1984;  $T_m = 81.5^\circ\text{C} + 16.6 (\log M) + 0.41 (\%GC) - 0.61 (\% \text{ form}) - 500/L$ ; where  $M$  is the molarity of monovalent cations,  $\%GC$  is the percentage of guanosine and cytosine nucleotides in the DNA,  $\% \text{ form}$  is the percentage of formamide in the hybridization solution, and  $L$  is the length of the hybrid in base pairs.  $T_m$  is reduced by about  $1^\circ\text{C}$  for each 1% of mismatching; thus,  $T_m$ , hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with  $>90\%$  identity are sought, the  $T_m$  can be decreased  $10^\circ\text{C}$ . Generally, stringent conditions are selected to be about  $5^\circ\text{C}$  lower than the thermal melting point  $T_m$  for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or  $4^\circ\text{C}$  lower than the thermal melting point  $T_m$ ; moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or  $10^\circ\text{C}$  lower than the thermal melting point  $T_m$ ; low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or  $20^\circ\text{C}$  lower than the thermal melting point  $T_m$ . Using the equation, hybridization and wash compositions, and desired  $T$ , those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a  $T$  of less than  $45^\circ\text{C}$  (aqueous solution) or  $32^\circ\text{C}$  (formamide solution), it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen, 1993. Generally, highly stringent hybridization and wash conditions are selected to be about  $5^\circ\text{C}$  lower than the thermal melting point  $T_m$  for the specific sequence at a defined ionic strength and pH.

An example of highly stringent wash conditions is 0.15 M NaCl at  $72^\circ\text{C}$  for about 15 minutes. An example of stringent wash conditions is a 0.2X SSC wash at  $65^\circ\text{C}$  for 15 minutes (see, Sambrook, *infra*, for a description of SSC buffer). Often, a high stringency wash is preceded by a low stringency wash to remove background probe signal. An example medium

stringency wash for a duplex of, e.g., more than 100 nucleotides, is 1X SSC at 45°C for 15 minutes. An example low stringency wash for a duplex of, e.g., more than 100 nucleotides, is 4-6X SSC at 40°C for 15 minutes. For short probes (e.g., about 10 to 50 nucleotides), stringent conditions typically involve salt concentrations of less than about 1.5 M, more preferably about 0.01 to 1.0 M, Na ion concentration (or other salts) at pH 7.0 to 8.3, and the temperature is typically at least about 30°C and at least about 60°C for long robes (e.g., >50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. In general, a signal to noise ratio of 2X (or higher) than that observed for an unrelated probe in the particular hybridization assay indicates detection of a specific hybridization. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the proteins that they encode are substantially identical. This occurs, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code.

Very stringent conditions are selected to be equal to the  $T_m$  for a particular probe. An example of stringent conditions for hybridization of complementary nucleic acids which have more than 100 complementary residues on a filter in a Southern or Northern blot is 50% formamide, e.g., hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C.

The following are examples of sets of hybridization/wash conditions that may be used to clone orthologous nucleotide sequences that are substantially identical to reference nucleotide sequences of the present invention: a reference nucleotide sequence preferably hybridizes to the reference nucleotide sequence in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 2X SSC, 0.1% SDS at 50°C, more desirably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 1X SSC, 0.1% SDS at 50°C, more desirably still in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.5X SSC, 0.1% SDS at 50°C, preferably in 7% sodium

dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 50°C, more preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO<sub>4</sub>, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 65°C.

One non-limiting source of a purified polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin is the cell-free extract described in the examples below. Another method for purifying a polypeptide exhibiting a P450 monooxygenase activity in accordance with the invention is to attach a tag to the protein, thereby facilitating its purification. Accordingly, the invention provides a purified polypeptide exhibiting an enzymatic activity of a P450 monooxygenase which regioselectively oxidizes avermectin to 4''-keto-avermectin, wherein the polypeptide is covalently bound to a tag. The invention further provides a nucleic acid molecule encoding such a tagged polypeptide.

As used herein, a "tag" is meant a polypeptide or other molecule covalently bound to a polypeptide of the invention, whereby a binding agent (*e.g.*, a polypeptide or molecule) specifically binds the tag. In accordance with the invention, by "specifically binds" is meant that the binding agent (*e.g.*, an antibody or Ni<sup>2+</sup> resin) recognizes and binds to a particular polypeptide or chemical but does not substantially recognize or bind to other molecules in the sample. In some embodiments, a binding agent that specifically binds a ligand forms an association with that ligand with an affinity of at least 10<sup>6</sup> M<sup>-1</sup>, or at least 10<sup>7</sup> M<sup>-1</sup>, or at least 10<sup>8</sup> M<sup>-1</sup>, or at least 10<sup>9</sup> M<sup>-1</sup> either in water, under physiological conditions, or under conditions which approximate physiological conditions with respect to ionic strength, *e.g.*, 140 mM NaCl, 5 mM MgCl<sub>2</sub>. For example, a His tag is specifically bound by nickel (*e.g.*, the Ni<sup>2+</sup>-charged column commercially available as His•Bind® Resin from Novagen Inc, Madison, WI). Likewise, a Myc tag is specifically bound by an antibody that specifically binds Myc.

As described below, a His tag is attached to the purified polypeptide of the invention exhibiting an enzymatic activity of a P450 monooxygenase by generating a nucleic acid molecule encoding the His-tagged polypeptide, and expressing the polypeptide in *E. coli*. These polypeptides, once expressed by *E. coli*, are readily purified by standard techniques (*e.g.*, using one of the His•Bind® Kits commercially available from Novagen or using the TALON™ Resin (and manufacturer's instructions) commercially available from Clontech Laboratories, Inc., Palo Alto, CA).

Additional tags may be attached to any or all of the polypeptides of the invention to facilitate purification. These tags include, without limitation, the HA-Tag (amino acid sequence: YPYDVPDYA (SEQ ID NO:39)), the Myc-tag (amino acid sequence: EQKLISEEDL (SEQ ID NO:40)), the HSV tag (amino acid sequence: QPELAPEDPED (SEQ ID NO:41)), and the VSV-G-Tag (amino acid sequence: YTDIEMNRLGK (SEQ ID NO:42)). Covalent attachment (*e.g.*, via a polypeptide bond) of these tags to a polypeptide of the invention allows purification of the tagged polypeptide using, respectively, an anti-HA antibody, an anti-Myc antibody, an anti-HSV antibody, or an anti-VSV-G antibody, all of which are commercially available (for example, from MBL International Corp., Watertown, MA; Novagen Inc.; Research Diagnostics Inc., Flanders, NJ).

The tagged polypeptides of the invention exhibiting a P450 monooxygenase activity may also be tagged by a covalent bond to a chemical, such as biotin, which is specifically bound by streptavidin, and thus may be purified on a streptavidin column. Similarly, the tagged P450 monooxygenases of the invention may be covalently bound (*e.g.*, via a polypeptide bond) to the constant region of an antibody. Such a tagged P450 monooxygenase may be purified, for example, on protein A sepharose.

The tagged P450 monooxygenases of the invention may also be tagged to a GST (glutathione-S-transferase) or the constant region of an immunoglobulin. For example, a nucleic acid molecule of the invention (*e.g.*, comprising SEQ ID NO:1) can be cloned into one of the pGEX plasmids commercially available from Amersham Pharmacia Biotech, Inc. (Piscataway NJ), and the plasmid expressed in *E. coli*. The resulting P450 monooxygenase encoded by the nucleic acid molecule is covalently bound to a GST (glutathione-S-transferase). These GST fusion proteins can be purified on a glutathione agarose column (commercially available from, *e.g.*, Amersham Pharmacia Biotech), and thus purified. Many of the pGEX plasmids enable easy removal of the GST portion from the fusion protein. For example, the pGEX-2T plasmid contains a thrombin recognition site between the inserted nucleic acid molecule of interest and the GST-encoding nucleic acid sequence. Similarly, the pGES-3T plasmid contains a factor Xa site. By treating the fusion protein with the appropriate enzyme, and then separating the GST portion from the P450 monooxygenase of the invention using glutathione agarose (to which the GST specifically binds), the P450 monooxygenase of the invention can be purified.

Yet another method to obtain a purified polypeptide of the invention exhibiting a P450 monooxygenase activity is to use a binding agent that specifically binds to such a polypeptide. Accordingly, the invention provides a binding agent that specifically binds to a P450 monooxygenase of the invention. This binding agent of the invention may be a chemical compound (*e.g.*, a protein), a metal ion, or a small molecule.

In particular embodiments, the binding agent is an antibody. The term “antibody” encompasses, without limitation, polyclonal antibody, monoclonal antibody, antibody fragments (*e.g.*, Fab, Fv, or Fab’ fragments), single chain antibody, chimeric antibody, bi-specific antibody, antibody of any isotype (*e.g.*, IgG, IgA, and IgE), and antibody from any species (*e.g.*, rabbit, mouse, and human).

In one non-limiting example, the binding agent of the invention is a polyclonal antibody. In another non-limiting example, the binding agent of the invention is a monoclonal antibody. Methods for making both monoclonal and polyclonal antibodies are well known (*see, e.g.*, Current Protocols in Immunology, ed. John E. Coligan, John Wiley & Sons, Inc. 1993; Current Protocols in Molecular Biology, eds. Ausubel *et al.*, John Wiley & Sons, Inc. 2000).

The polypeptides described herein exhibiting an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4”-keto-avermectin belong to a family of novel P450 monooxygenases. Accordingly, the invention also provides a family of P450 monooxygenase polypeptides, wherein each member of the family regioselectively oxidizes avermectin to 4”-keto-avermectin. In some embodiments, each member of the family comprises or consists of an amino acid sequence that is at least 50% identical to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95. In particular embodiments, each member of the family is encoded by a nucleic acid molecule comprising or consisting of a nucleic acid sequence that is at least 66% identical to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94.

The present invention, which provides an entire family of P450 monooxygenases, each member of which is able to regioselectively oxidize avermectin to 4''-keto-avermectin, allowed for the generation of an improved P450 monooxygenase, which may not be naturally occurring, but which regioselectively oxidizes avermectin to 4''-keto-avermectin with efficiency and with reduced undesirable side product. For instance, one of the members of the P450 monooxygenase family of the invention, P450<sub>Ema1</sub> enzyme catalyzes a further oxidation that is not desirable, since the formation of 3''-O-demethyl-4''-keto-avermectin has been detected in the reaction by *Streptomyces tubercidicus* strain R-922 and by *Streptomyces lividans* containing the *ema1* gene. The formation of 3''-O-demethyl-4''-keto-avermectin is brought about by the oxidation of the 3''-O-methyl group, whereby the hydrolytically labile 3''-O-hydroxymethyl group is formed which hydrolyzes to form formaldehyde and the 3''-hydroxyl group.

By providing a family of polypeptides exhibiting an enzymatic activity of P450 monooxygenases that regioselectively oxidize avermectin to 4''-keto-avermectin (*see, e.g.*, Table 3 below), individual members of the family can be subjected to family gene shuffling efforts in order to produce new hybrid genes encoding optimized P450 monooxygenases of the invention. In one non-limiting example, a portion of the *ema1* gene encoding the O<sub>2</sub> binding site of the P450<sub>Ema1</sub> protein can be swapped with the portion of the *ema2* gene encoding the O<sub>2</sub> binding site of the P450<sub>Ema2</sub> protein. Such a chimeric *ema1/2* protein is within definition of a P450 monooxygenase of the invention.

Site-directed mutagenesis or directed evolution technologies may also be employed to generate derivatives of the *ema1* gene that encode enzymes with improved properties, including higher overall activity and/or reduced side product formation. One method for deriving such a mutant is to mutate the *Streptomyces* strain itself, in a manner similar to the UV mutation of *Streptomyces tubercidicus* strain R-922 described below.

Additional derivatives may be made by making conservative or non-conservative changes to the amino acid sequence of a P450 monooxygenase. Conservative and non-conservative amino acid substitutions are well known (*see, e.g.*, Stryer, *Biochemistry*, 3<sup>rd</sup> Ed., W.H. Freeman and Co., NY 1988). Similarly, truncations of a P450 monooxygenase of the invention may be generated by truncating the protein at its N-terminus (*e.g.*, *see the ema1A*

gene described below), at its C-terminus, or truncating (*i.e.*, removing amino acid residues) from the middle of the protein.

Such a mutant, derivative, or truncated P450 monooxygenase is a P450 monooxygenase of the invention as long as the mutant, derivative, or truncated P450 monooxygenase is able to regioselectively oxidize avermectin to 4"-keto-avermectin.

In another aspect, the invention provides a cell genetically engineered to comprise a nucleic acid molecule encoding a polypeptide which exhibits an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4"-keto-avermectin. By "genetically engineered" is meant that the nucleic acid molecule is exogenous to the cell into which it is introduced. Introduction of the exogenous nucleic acid molecule into the genetically engineered cell may be accomplished by any means, including, without limitation, transfection, transduction, and transformation.

In certain embodiments, the nucleic acid molecule is positioned for expression in the genetically engineered cell. By "positioned for expression" is meant that the exogenous nucleic acid molecule encoding the polypeptide is linked to a regulatory sequence in such a way as to permit expression of the nucleic acid molecule when introduced into a cell. By "regulatory sequence" is meant nucleic acid sequences, such as initiation signals, polyadenylation (polyA) signals, promoters, and enhancers, which control expression of protein coding sequences with which they are operably linked. By "expression" of a nucleic acid molecule encoding a protein or polypeptide fragment is meant expression of that nucleic acid molecule as protein and/or mRNA.

A genetically engineered cell of the invention may be a prokaryotic cell (*e.g.*, *E. coli*) or a eukaryotic cell (*e.g.*, *Saccharomyces cerevisiae* or mammalian cell (*e.g.*, HeLa)). According to some embodiments of the invention, the genetically engineered cell is a cell wherein the wild-type (*i.e.*, not genetically engineered) cell does not naturally contain the inserted nucleic acid molecule and does not naturally express the protein encoded by the inserted nucleic acid molecule. Accordingly, the cell may be a genetically engineered *Streptomyces* strain, such as a *Streptomyces lividans* or a *Streptomyces avermitilis* strain. Alternatively, the cell may be a genetically engineered *Pseudomonas* strain, such as a *Pseudomonas putida* strain or a *Pseudomonas fluorescens* strain. In another alternative, the cell may be a genetically engineered *Escherichia coli* strain.



Note that in some types of cells genetically engineered to comprise a nucleic acid molecule encoding a polypeptide which exhibits an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin, the actual genetically engineered cell, itself, may not be able to convert avermectin into 4''-keto-avermectin. Rather, the P450 monooxygenase heterologously expressed by such a genetically engineered cell may be purified from that cell, where the purified P450 monooxygenase of the invention can be used to regioselectively oxidize avermectin to 4''-keto-avermectin. Thus, the genetically engineered cell of the invention need not, itself, be able to regioselectively convert avermectin to 4''-keto-avermectin; rather, the genetically engineered cell of the invention need only comprise a nucleic acid molecule encoding a polypeptide which exhibits an enzymatic activity of a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin, regardless of whether the polypeptide is active inside that cell.

In addition, a cell (*e.g.*, *E. coli*) genetically engineered to comprise a nucleic acid molecule encoding a polypeptide of the invention which exhibits an enzymatic activity of a P450 monooxygenase may not be able to regioselectively oxidize avermectin to 4''-keto-avermectin, although the P450 monooxygenase purified from the genetically engineered cell is able to regioselectively oxidize avermectin to 4''-keto-avermectin. However, if the same cell were genetically engineered to comprise a polypeptide of the invention which exhibits an enzymatic activity of a P450 monooxygenase, a ferredoxin of the invention, and/or a ferredoxin reductase of the invention, then the P450 monooxygenase together with the ferredoxin and the ferredoxin reductase, all purified from that cell, and in the presence of a reducing agent (*e.g.*, NADH or NADPH), would be able to regioselectively oxidize avermectin to 4''-keto-avermectin. Furthermore the genetically engineered cell comprising a P450 monooxygenase of the invention, a ferredoxin of the invention, and a ferredoxin reductase of the invention, itself, might be able to carry out this oxidation.

Moreover, in a non-limiting example where a cell (*e.g.*, *E. coli*) is genetically engineered to express P450 monooxygenase, a ferredoxin, and a ferredoxin reductase proteins of the invention, all three of these proteins, when purified from the genetically engineered *E. coli*, are together and in the presence of a reducing agent (*e.g.*, NADH or NADPH) would be active and able to regioselectively oxidize avermectin to 4''-keto-avermectin, and so are useful in a method for making emamectin.

In accordance with the present invention, the following material has been deposited with the Agricultural Research Service, Patent Culture Collection (NRRL), 1815 North University Street, Peoria, Illinois 61604, under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure: (1) *Streptomyces lividans* ZX7 (*ema1/fd233*-TUA1A) NRRL Designation No. B-30478; and (2) *Pseudomonas putida* NRRL B-4067 containing plasmid pRK290-*ema1/fd233*, NRRL Designation No. B-30479

In identifying the novel family of polypeptides exhibiting an enzymatic activity of P450 monooxygenases that regioselectively oxidize avermectin to 4''-keto-avermectin, novel ferredoxins and novel ferredoxin reductases were also identified in the same strains of bacteria in which the P450 monooxygenases were found. Accordingly, in a further aspect, the invention provides a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a ferredoxin, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a polypeptide that regioselectively oxidizes avermectin to 4''-keto-avermectin. Similarly, the invention provides a purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a ferredoxin reductase, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a polypeptide that regioselectively oxidizes avermectin to 4''-keto-avermectin. The invention also provides a purified protein that exhibits an enzymatic activity of a ferredoxin, as well as a purified protein that exhibits an enzymatic activity of a ferredoxin reductase, wherein the ferredoxin protein and the ferredoxin reductase protein are isolated from a *Streptomyces* strain comprising a polypeptide that regioselectively oxidizes avermectin to 4''-keto-avermectin.

A useful nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a ferredoxin comprises or consists essentially of a nucleic acid sequence that is at least 81% identical to SEQ ID NO:35 or SEQ ID NO:37. Alternatively, the nucleic acid molecule comprises or consists essentially of a nucleic acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:35 or SEQ ID NO:37. The nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide that exhibits an enzymatic activity of a ferredoxin

may comprise or consist essentially of the nucleic acid sequence of SEQ ID NO:35 or SEQ ID NO:37.

The protein of the invention exhibiting a ferredoxin activity may comprise or consist essentially of an amino acid sequence that is at least 80% identical to SEQ ID NO:36 or SEQ ID NO:38. In some embodiments, the nucleic acid molecule comprises or consists essentially of an amino acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:36 or SEQ ID NO:38. The ferredoxin of the invention may comprise or consist essentially of the amino acid sequence of SEQ ID NO:36 or SEQ ID NO:38.

A useful nucleic acid molecule comprising a nucleotide sequence encoding a protein of the invention exhibiting a ferredoxin reductase comprises or consists essentially of the nucleic acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:104. In a particular embodiment of the invention, the nucleic acid molecule encoding a ferredoxin reductase of the invention may comprise or consist essentially of the amino acid sequence of SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:102, or SEQ ID NO:104.

The ferredoxin reductase of the invention may comprise or consist essentially of the amino acid sequence that is at least 85%, or at least 90%, or at least 95%, or at least 99% identical to SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, or SEQ ID NO:105. In a particular embodiment of the invention, the ferredoxin reductase of the invention may comprise or consist essentially of the amino acid sequence of SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, or SEQ ID NO:105.

. Methods for purifying ferredoxin and ferredoxin reductase proteins and nucleic acid molecules encoding such ferredoxin and ferredoxin reductase proteins are known in the art and are the same as those described above for purifying P450 monooxygenases of the invention and nucleic acid molecules encoding P450 monooxygenases of the invention.

In one non-limiting example to obtain a purified P450 monooxygenase of the invention with a purified ferredoxin, a *S. lividans* strain (or *P. putida* strain, or any other cell in which the P450 monooxygenase of the invention does not naturally occur) may be genetically engineered to contain a first nucleic acid molecule encoding a P450 monooxygenase of the invention and a second nucleic acid molecule encoding a ferredoxin protein, where both the first and second nucleic acid molecules are positioned for expression in the genetically

engineered cell. The first and the second nucleic acid molecules can be on separate plasmids, or can be on the same plasmid. Thus, the same engineered cell or strain will produce both the P450 monooxygenase of the invention and the ferredoxin protein of the invention.

In a further non-limiting example to obtain a purified P450 monooxygenase of the invention with a purified ferredoxin and with a purified ferredoxin reductase of the invention, a *S. lividans* strain (or *P. putida* strain, or any other cell in which the P450 monooxygenase of the invention does not naturally occur) may be genetically engineered to contain a first nucleic acid molecule encoding a P450 monooxygenase of the invention and a second nucleic acid molecule encoding a ferredoxin protein of the invention and a third nucleic acid molecule encoding a ferredoxin reductase protein of the invention, where all the first and second and third nucleic acid molecules are positioned for expression in the genetically engineered cell. The first and the second and the third nucleic acid molecules may be provided on separate plasmids, or on the same plasmid. Thus, the same engineered cell or strain will produce all the P450 monooxygenase of the invention and the ferredoxin and the ferredoxin reductase proteins of the invention.

As described above for the P450 monooxygenases of the invention, the ferredoxin protein and/or the ferredoxin reductase protein may further comprise a tag. Moreover, the invention contemplates binding agents (*e.g.*, antibodies) that specifically bind to the ferredoxin protein, and binding agents that specifically bind to the ferredoxin reductase proteins of the invention. Methods for generating tagged ferredoxin protein, tagged ferredoxin reductase protein, and binding agents (*e.g.*, antibodies) that specifically bind to ferredoxin or ferredoxin reductase are the same as those as described above for generating tagged P450 monooxygenases of the invention and generating binding agents that specifically bind P450 monooxygenases of the invention.

The invention also provides a method for making emamectin. In this method, a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin is added to a reaction mixture containing avermectin. The reaction mixture is then incubated under conditions that allow the P450 monooxygenase to regioselectively oxidize avermectin to 4''-keto-avermectin. The reaction mixture may further comprise a ferredoxin, such as a ferredoxin of the present invention. In particular embodiments, the reaction mixture further

comprises a ferredoxin reductase such as a ferredoxin of the present invention. The reaction mixture may further comprise a reducing agent, such as NADH or NADPH.

Additionally, the invention provides a method for making 4''-keto-avermectin. The method comprises adding a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin to a reaction mixture comprising avermectin and incubating the reaction mixture under conditions that allow the P450 monooxygenase to regioselectively oxidize avermectin to 4''-keto-avermectin. In some embodiments, the reaction mixture further comprises a ferredoxin, such as a ferredoxin of the present invention. The reaction mixture may also further comprise a ferredoxin reductase such as a ferredoxin of the present invention. In particular embodiments, the reaction mixture further comprises a reducing agent, such as NADH or NADPH.

The invention also provides a formulation for making emamectin comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin. In some embodiments, the formulation further comprises a ferredoxin, such as a ferredoxin of the present invention. In particular embodiments, the ferredoxin is isolated from the same species of cell or strain from which the P450 monooxygenase was isolated or derived. The formulation may further comprise a ferredoxin reductase, such as a ferredoxin reductase of the present invention. In particular embodiments, the ferredoxin reductase is isolated from the same species of cell or strain from which the P450 monooxygenase was isolated or derived. In some embodiments, the formulation further comprises a reducing agent, such as NADH or NADPH.

In addition, the invention provides a formulation for making 4''-keto-avermectin comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4''-keto-avermectin. In some embodiments, the formulation further comprises a ferredoxin, such as a ferredoxin of the present invention. In particular embodiments, the ferredoxin is isolated from the same species of cell or strain from which the P450 monooxygenase was isolated or derived. In some embodiments, the formulation further comprises a ferredoxin reductase, such as a ferredoxin reductase of the present invention. In particular embodiments, the ferredoxin reductase is isolated from the same species of cell or strain from which the P450 monooxygenase was isolated or derived. The formulation may further comprise a reducing agent, such as NADH or NADPH.

The following examples are intended to further illustrate certain preferred embodiments of the invention and are not limiting in nature.

### EXAMPLE I

#### Optimized Growth Conditions for *Streptomyces tubercidicus* Strain R-922

In one non-limiting example the fermentation conditions needed to provide a steady supply of cells of *Streptomyces tubercidicus* strain R-922 highly capable of regioselectively oxidizing avermectin to 4''-keto-avermectin were optimized.

First, the following solutions were made. For ISP-2 agar, 4 g of yeast extract (commercially available from Oxoid Ltd, Basingstoke, UK), 4 g of D(+)-glucose, 10 g of bacto malt extract (Difco No. 0186-17-7 (Difco products commercially available from, *e.g.*, Voigt Global Distribution, Kansas City, MO)), and 20 g of agar (Difco No. 0140-01) were dissolved in one liter of demineralized water, and the pH is adjusted to 7.0. The solution was sterilized at 121°C for 20 min., cooled down, and kept at 55°C for the time needed for the immediate preparation of the agar plates.

For PHG medium, 10 g of peptone (Sigma 0521; commercially available from Sigma Chemical Co., St. Louis, MO), 10 g of yeast extract (commercially available from Difco), 10 g of D(+)-glucose, 2 g of NaCl, 0.15 g of  $\text{MgSO}_4 \times 7 \text{H}_2\text{O}$ , 1.3 g of  $\text{NaH}_2\text{PO}_4 \times \text{H}_2\text{O}$ , and 4.4 g of  $\text{K}_2\text{HPO}_4$  were dissolved in 1 liter of demineralized water, and the pH was adjusted to 7.0.

*Streptomyces tubercidicus* strain R-922 was grown in a Petri dish on ISP-2 agar at 28°C. This culture was used to inoculate four 500 ml shaker flasks with a baffle, each containing 100 ml PHG medium. These pre-cultures were grown on an orbital shaker at 120 rpm at 28°C for 72 hours and then used to inoculate a 10-liter fermenter equipped with a mechanical stirrer and containing 8 liters of PHG medium. This main culture was grown at 28°C with stirring at 500 rpm and with aeration of 1.75 vvm (14 l/min.) and a pressure of 0.7 bar. At the end of the exponential growth, after about 20 hours, the cells were harvested by centrifugation. The yield of wet cells was 70-80 g/l culture.

### EXAMPLE II

### Whole Cell Biocatalysis Assay

As determined in accordance with the present invention, the following whole cell biocatalysis assay was employed to determine that the activity from *Streptomyces* cells capable of regioselectively oxidizing avermectin to 4''-keto-avermectin is catalyzed by a P450 monooxygenase.

*Streptomyces tubercidicus* strain R-922 was grown in PHG medium, and *Streptomyces tubercidicus* strain I-1529 was grown in M-17 or PHG medium. PHG medium contains 10 g/l Peptone (Sigma, 0.521), 10 g/l Yeast Extract (Difco, 0127-17-9), 10 g/l D-Glucose, 2 g/l NaCl, 0.15 g/l MgSO<sub>4</sub> x 7 H<sub>2</sub>O, 1.3 g/l NaH<sub>2</sub>PO<sub>4</sub> x 1 H<sub>2</sub>O, and 4.4 g/l K<sub>2</sub>HPO<sub>4</sub> at pH 7.0. M-17 medium contains 10 g/l glycerol, 20 g/l Dextrin white, 10 g/l Soytone (Difco 0437-17), 3 g/l Yeast Extract (Difco 0127-17-9), 2 g/l (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, and 2 g/l CaCO<sub>3</sub> at pH 7.0

To grow the cells, an ISP2 agar plate (not older than 1-2 weeks) was inoculated and incubated for 3-7 days until good growth was achieved. Next, an overgrown agar piece was transferred (with an inoculation loop) to a 250ml Erlenmeyer flask with 1 baffle containing 50 ml PHG medium. This pre-culture is incubated at 28°C and 120 rpm for 2-3 days. Next, 5 ml of the pre-culture were transferred to a 500 ml Erlenmeyer flask with 1 baffle containing 100 ml PHG medium. The main culture was incubated at 28°C and 120 rpm for 2 days. Next, the culture was centrifuged for 10 min. at 8000 rpm on a Beckman Rotor JA-14. The cells were next washed once with 50 mM potassium phosphate buffer, pH 7.0.

To perform the whole cell biocatalysis assay, 500 mg wet cells were placed into a 25 ml Erlenmeyer flask, to which were added 10 ml of 50 mM potassium phosphate buffer, pH 7.0. The cells were stirred with a magnetic stir bar to distribute the cells. Next, 15 µl of a solution of avermectin B1a in isopropanol (30 mg/ml) were added, and the mixture shaken on an orbital shaker at 160 rpm and 28°C. Strain R-922 was reacted for 2 hours, and strain I-1529 was reacted for 30 hours.

To work up the cultures in the whole cell biocatalysis assay, 10 ml methyl-t-butyl-ether was added to an Erlenmeyer flask containing the resting cells and the entire cell mixture was transferred to a 30 ml-centrifuge tube, shaken vigorously, and then centrifuged at 16000 rpm for 10 min. The ether phase was pipetted into a 50 ml pear flask, and evaporated in vacuo by means of a rotary evaporator (≤0.1 mbar). The residue was re-dissolved in 1.2 ml acetonitrile

and transferred to an HPLC-sample vial. The conversion of avermectin B1a to 4''-hydroxy-avermectin B1a and 4''-keto-avermectin B1a (also called 4''-oxo-avermectin B1a) and the formation of a side product from the biocatalysis reaction could be observed by HPLC analysis using HPLC protocol I.

For HPLC protocol I, the following parameters were used:

#### Hardware

Pump:	L-6250 Merck-Hitachi
Autosampler:	AS-2000A Merck-Hitachi
Interface Module:	D-6000 Merck-Hitachi
Channel 1-Detector:	L-7450A UV-Diode Array Merck-Hitachi
Column Oven:	none
Column:	70mm x 4mm
Adsorbent:	Kromasil 100Å-3.5µ-C18

#### Gradient Mode: Low

<i>Pressure Limit:</i>	5-300bar		
<i>Column Temperature</i>	ambient ( $\approx 20^{\circ}\text{C}$ )		
Solvent A:	acetonitrile		
Solvent B:	water		
Flow:	1.5 ml/min		
Detection:	243 nm		
Pump Table:	0.0 min	75% A	25% B
<i>linear gradient</i>	7.0 min	100% A	0% B
	9.0 min	100% A	0% B
<i>jump</i>	9.1 min	75% A	25% B
	12.0 min	75% A	25% B
Stop time:	12 min		
Sampling Period:	every 200 msec		
Retention time table:	<i>time</i>	<i>References</i>	
	2.12 min	4''-hydroxy- avermectin B1a	



3.27 min	avermectin B1a
3.77 min	3''-O-demethyl-4''-keto-avermectin B1a
4.83 min	4''-keto-avermectin B1a

### EXAMPLE III

#### Biotransformation With Cell-Free Extract From *Streptomyces* Strain R-922

To prepare an active cell-free extract from *Streptomyces tubercidicus* strain R-922 capable of regioselective oxidation of avermectin to 4''-keto-avermectin, the following solutions were made, stored at 4°C, and kept on ice when used.

Solution	Formula
PP-buffer	50 mM K <sub>2</sub> HPO <sub>4</sub> /KH <sub>2</sub> PO <sub>4</sub> (pH 7.0)
Disruption buffer	50 mM K <sub>2</sub> HPO <sub>4</sub> /KH <sub>2</sub> PO <sub>4</sub> (pH 7.0), 5 mM benzamidine, 2 mM dithiothreitol, and 0.5 mM Pefabloc (from Roche Diagnostics)
Substrate	10 mg avermectin were dissolved in 1 ml isopropanol

Six grams of wet cells from *Streptomyces* strain R-922 were washed in PP-buffer and then resuspended in 35 ml disruption buffer and disrupted in a French press at 4°C. The resulting suspension was centrifuged for 1 hour at 35000 x g. The supernatant of the cell free extract was collected. One µl substrate was added to 499µl of cleared cell free extract and incubated at 30°C for 1 hour. Then, 1 ml methyl-t-butyl ether was added to the reaction mixture and thoroughly mixed. The mixture was next centrifuged for 2 min. at 14000 rpm, and the methyl-t-butyl ether phase was transferred into a 10 ml flask and evaporated in vacuo

by means of a rotary evaporator. The residue was dissolved in 200  $\mu$ l acetonitrile and transferred into an HPLC-sample vial.

For HPLC, the HPLC protocol I was used.

When 1  $\mu$ l substrate was added to 499  $\mu$ l of cleared cell free extract and incubated at 30°C, no conversion of avermectin to 4"-keto-avermectin was observed by HPLC analysis using HPLC protocol I.

However, the possibility of addition of spinach ferredoxin and spinach ferredoxin reductase and NADPH to the cell free extract to restore the biocatalytic activity was explored (*see, generally, D.E. Cane and E.I. Graziani, J. Amer. Chem. Soc. 120:2682, 1998*).

Accordingly, the following solutions were made:

Solution	Formula
Substrate	10 mg avermectin were dissolved in 1 ml isopropanol
Ferredoxin	5 mg ferredoxin (from spinach), solution 1-3 mg/ml in Tris/HCl-buffer (from Fluka) or 5 mg ferredoxin (from <i>Clostridium pasteurianum</i> ), solution of 1-3 mg/ml in Tris/HCl-buffer (from Fluka) or 5 mg ferredoxin (from <i>Porphyra umbilicalis</i> ), solution of 1-3 mg/ml in Tris/HCl-buffer (from Fluka)
Ferredoxin Reductase	1 mg freeze-dried ferredoxin reductase (from spinach), solution of 3.9 U/mg in 1 ml H <sub>2</sub> O (from Sigma)
NADPH	100 mM NADPH in H <sub>2</sub> O (from Roche Diagnostics)

The substrate solution was stored at 4°C, the other solutions were stored at -20°C, and kept on ice when used.

Thus, to 475  $\mu$ l of cleared cell free extract the following solutions were added: 10  $\mu$ l ferredoxin, 10  $\mu$ l ferredoxin reductase and 1  $\mu$ l substrate. After the addition of substrate to the cells, the mixture was immediately and thoroughly mixed and aerated. Then, 5  $\mu$ l of NADPH were added and the mixture incubated at 30°C for 30 min. Then, 1 ml methyl-t-butyl ether was added to the reaction mixture and thoroughly mixed. The mixture was next centrifuged for 2 min. at 14000 rpm, and the methyl-t-butyl ether phase was transferred into a 10 ml flask

and evaporated in vacuo by means of a rotary evaporator. The residue was dissolved in 200  $\mu$ l acetonitrile and transferred into an HPLC-sample vial, and HPLC analysis performed using HPLC protocol I.

Formation of 4"-keto-avermectin was observable by HPLC analysis. Thus, addition of spinach ferredoxin and spinach ferredoxin reductase and NADPH to the cell free extract restored the biocatalytic activity.

Upon injection of a 30  $\mu$ l sample, a peak appeared at 4.83 min., indicating the presence of 4"-keto-avermectin B1a. A mass of 870 D could be assigned to this peak by HPLC-mass spectrometry which corresponds to the molecular weight of 4"-keto-avermectin B1a.

Note that when analyzing product formation by HPLC and HPLC-mass spectrometry, in addition to the 4"-keto-avermectin, the corresponding ketohydrate 4"-hydroxy-avermectin was also found giving a peak at 2.12 min. This finding indicated that the P450 monooxygenase converts avermectin by hydroxylation to 4"-hydroxy-avermectin, from which 4"-keto-avermectin is formed by dehydration. Interestingly, when the spinach ferredoxin was replaced by ferredoxin from the bacterium *Clostridium pasteurianum* or from the red alga *Porphyra umbilicalis*, the biocatalytic conversion of avermectin to 4"-keto-avermectin still took place, indicating that the enzyme does not depend on a specific ferredoxin for receiving reduction equivalents.

#### EXAMPLE IV

##### Isolation of a Mutant *Streptomyces* Strain R-922 With Enhanced Activity

To obtain strains of *Streptomyces* strain R-922 that have an enhanced ability to regioselectively oxidize avermectin to 4"-keto-avermectin, UV mutants were generated. To do this, spores of *Streptomyces* strain R-922 were collected and stored in 15% glycerol at -20°C. This stock solution contained  $2 \times 10^9$  spores.

The spore stock solution was next diluted and transferred to petri plates containing 10ml of sterile water, and the suspension was exposed to UV light in a Stratalinker UV crosslinker 2400 (commercially available from Stratagene, La Jolla, CA). The Stratalinker UV crosslinker uses a 254-nm light source and the amount of energy used to irradiate a sample can be set in the "energy mode."

Applicant's or agent's file reference PB/5-60016A	International application No. PCT/EP 02/05363
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OR OTHER BIOLOGICAL MATERIAL**

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B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution Agricultural Research Service, Patent Culture Collection (NRRL)	
Address of depositary institution (including postal code and country) 1815 North University Street Peoria Illinois 61604 USA	
Date of deposit May 08, 2001	Accession Number NRRL B-30479
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Pseudomonas putida NRRL B-4067 containing plasmid pRK290-ema1/fd233	
D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE (if the indications are not for all designated States)	
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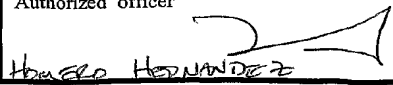
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Name of depositary institution Agricultural Research Service, Patent Culture Collection (NRRL)	
Address of depositary institution (including postal code and country) 1815 North University Street Peoria Illinois 61604 USA	
Date of deposit May 08, 2001	Accession Number NRRL B-30478
<b>C. ADDITIONAL INDICATIONS</b> (leave blank if not applicable) This information is continued on an additional sheet <input type="checkbox"/>	
Streptomyces lividans ZX7 (ema1/fd233-TUA1A)	
<b>D. DESIGNATED STATES FOR WHICH INDICATIONS ARE MADE</b> (if the indications are not for all designated States)	
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Through experimentation, it was determined that an exposure of 8000 microjoules of UV irradiation (254 nm) was required to kill 99.9% of the spores. This level of UV exposure was used in the mutagenesis.

Surviving UV-mutagenized spores were plated, cultured, and transferred to minimal media. Approximately 0.3-0.4% of the viable spores were determined to be auxotrophic, indicating a good level of mutagenesis in the population.

The mutagenized clones were screened for activity in the whole cell biocatalysis assay described in Example II. As shown in an HPLC chromatogram, one mutant ("R-922 UV mutant") showed a two to three fold increase in an ability to regioselectively oxidize avermectin to 4"-keto-avermectin as compared to wild-type strain R-922. Although the gene encoding the P450 monooxygenase responsible for the regioselectively oxidation activity, *ema1*, is not mutated in the R-922 UV mutant, this mutant nonetheless provides an excellent source for a cell-free extract containing *ema1* protein.

#### EXAMPLE V

##### Isolation of the P450 Monooxygenase from *Streptomyces* Strain R-922

To enrich the P450 enzyme, 35 ml of active cell free extract were filtered through a 45 µm filter and fractionated by anion exchange chromatography. Anion exchange chromatography conditions were as follows:

FPLC instrument: Äkta prime (from Pharmacia Biotech)

FPLC-column: HiTrap<sup>TM</sup>Q (5 ml) stacked onto Resource® Q (6 ml) (from Pharmacia Biotech)

eluent	buffer A: 25 mM Tris/HCl (pH 7.5)
	buffer B: 25 mM Tris/HCl (pH 7.5) containing 1 M KCl
temperature	eluent bottles and fractions in ice bath,
flow	3 ml/min
detection	UV 280nm
Pump table:	0.0 min    100% A    0% B
<i>linear gradient to</i>	2.0 min    90% A    10% B
	5.0 min    90% A    10% B

<i>linear gradient to</i> 30.0 min	50% A	50% B
<i>linear gradient to</i> 40.0 min	0% A	100% B
50.0min	0% A	100% B

Enzyme activity eluted with 35%-40% buffer B. The active fractions were pooled and concentrated by centrifugal filtration through Biomax™ filters with an exclusion limit of 5kD (commercially available from Millipore Corp., Bedford, MA) at 5000 rpm and then rediluted in disruption buffer containing 20% glycerol to a volume of 5 ml containing 3-10 mg/ml protein. This enriched enzyme solution contained at least 25% of the original enzyme activity.

The enzyme was further purified by size exclusion chromatography. Size exclusion chromatography conditions were as follows:

FPLC instrument:	Äkta prime (from Pharmacia Biotech)
FPLC-column:	HiLoad 26/60 Superdex® 200 prep grade (from Pharmacia Biotech)
sample:	3-5 ml enriched enzyme solution from the anion chromatography step
sample preparation:	filtered through 45 µm filter
eluent buffer:	PP-buffer (pH 7.0) + 0.1 M KCl
temperature:	4°C
flow:	2 ml/min
detection:	UV 280nm

Enzyme activity eluted between 205-235 ml eluent buffer. The active fractions were pooled, concentrated by centrifugal filtration through Biomax™ filters with an exclusion limit of 5 kD (from Millipore) at 5000 rpm, and rediluted in disruption buffer containing 20% glycerol to form a solution of 0.5-1 ml containing 2-5mg/ml protein. This enriched enzyme solution contained 10% of the original enzyme activity. This enzyme preparation, when checked for purity by SDS page, (*see*, generally, Laemmli, U.K., *Nature* **227**:680-685, 1970 and Current Protocols in Molecular Biology, *supra*) and stained with Coomassie blue, showed one dominant protein band with a molecular weight of 45-50 kD, according to reference proteins of known molecular weight.

#### EXAMPLE VI

Attempted Isolation of P450 Monooxygenase Genes  
From *Streptomyces* Strains R-922 and I-1529

Based on results described above that suggested the enzyme from strain R-922 that is responsible for the regiospecific oxidation of avermectin to 4''-keto-avermectin is a P450 monooxygenase, a direct PCR-based approach to clone P450 monooxygenase genes from this strain was initiated (*see, generally, Hyun et al., J. Microbiol. Biotechnol.* **8**(3):295-299, 1998). This approach is based on the fact that all P450 monooxygenase enzymes contain highly conserved oxygen-binding and heme-binding domains that are also conserved at the nucleotide level. PCR primers were designed to prime to these conserved domains and to amplify the DNA fragment from P450 genes using R-922 or I-1529 genomic DNA as a template. The PCR primers used are shown in Table 1.

Table 1

O <sub>2</sub> -Binding Domain Primers (5' to 3')*	Degeneracy	SEQ ID NOS
I A G H E T T ATC GCS GGS CAC GAG ACS AC	8	43
		44
V A G H E T T GTS GCS GGS CAC GAG ACS AC	16	45
		46
L A G H E T T CTS GCS GGS CAC GAG ACS AC	16	47
		48
L L L I A G H E T TS CTS CTS ATC GCS GGS CAC GAG AC <sup>&amp;</sup>	32	49
		50
Heme-Binding Domain Primers (3' to 5')*		
H Q C L G Q N L A GTG GTC ACG GAS CCS TGC TTG GAS CG <sup>&amp;</sup>	8	51
		52
F G H G V H Q C AAG CCS GTG CCS CAS GTG GTC ACG	8	53
		54
F G F G V H Q C AAG GCS AAG CCS CAS GTG GTC ACG	8	55
		56
F G H G I H Q C AAG CCS GTG CCS TAG GTG GTC ACG	4	57
		58



F	G	H	G	V	H	F	C		59
AAG	CCS	GTG	CCS	CAS	GTG	AAG	ACG	8	60

\* The amino acid sequence is shown on the top line and the corresponding nucleotide sequence is shown below on the second line; S=G or C.

\* This primer was described by Hyun *et al.*, *supra*

PCR amplification using any of the primers specific to nucleotide sequences encoding the O<sub>2</sub>-binding domain with any of the primers specific to the nucleotide sequences encoding the heme-binding domain and genomic DNA from *Streptomyces* strains R-922 or I-1529 resulted in the amplification of an approximately 350 bp DNA fragment. This is exactly the size that would be expected from this PCR amplification due to the approximately 350 bp separation in P450 genes of the gene segments encoding the O<sub>2</sub>-binding and heme-binding sites.

The 350 bp PCR fragments were cloned into the pCR2.1-TOPO TA cloning plasmid (commercially available Invitrogen, Carlsbad, CA) and transformed into *E. coli* strain TOP10 (Invitrogen, Carlsbad, CA). Approximately 150 individual clones from strains R-922 and I-1529 were sequenced to determine how many unique P450 gene fragments were represented. Analysis of the sequences revealed that they included 8 unique P450 gene fragments from strain R-922 and 7 unique fragments from I-1529.

Blast analysis (alignment of the deduced amino acid sequences of P450 gene-specific PCR fragments derived from *Streptomyces tubercidicus* strain R-922 and *Streptomyces* strain I-1529, respectively, and the P450 monooxygenase from *S. thermotolerans* that is involved in the synthesis of carbomycin (Stol-ORFA) (GenBank Accession No. D30759) by the program Pretty from the University of Wisconsin Package version 10.1 (Altschul *et al.*, *Nucl. Acids Res.* **25**:3389-3402). demonstrated that all of the unique P450 gene fragments from both the R-922 and I-1529 strains were derived from P450 genes and encoded the region between the O<sub>2</sub>-binding and heme-binding domains.

Next, in order to clone the full-length genes from which the PCR fragments were derived, the DNA fragments cloned by PCR were used as hybridization probes to gene libraries containing genomic DNA from strains R-922 and I-1529. To do this, genomic DNA from the R-922 and I-1529 strains was partially digested with Sau3A I, dephosphorylated with

calf intestinal alkaline phosphatase (CIP) and ligated into the cosmid pPEH215, a modified version of SuperCos 1 (commercially available from Stratagene, La Jolla, CA). Ligation products were packaged using the Gigapack III XL packaging extract and transfected into *E. coli* XL1 Blue MR host cells. Twelve cosmids that strongly hybridized to the PCR-generated P450 gene fragments were identified from the R-922 library, from which three unique P-450 genes were subcloned and sequenced. The hybridizations were performed at high stringency conditions according to the protocol of Church and Gilbert (Church and Gilbert, *Proc. Natl. Acad. Sci. USA* **81**:1991-1995, 1984). In brief, these high stringency conditions include Hybrid Buffer containing 500 mM Na-phosphate, 1 mM EDTA, 7% SDS, 1% BSA; Wash Buffer 1 containing 40 mM Na-phosphate, 1 mM EDTA, 5% SDS, 0.5% BSA; and Wash Buffer 2 containing 40 mM Na-phosphate, 1 mM EDTA, 1% SDS (Note that other high stringency hybridizations conditions are described, for example, in Current Protocols in Molecular Biology, *supra*.) Nineteen strongly hybridizing cosmids were identified from the I-1529 library, and from these, four unique P-450 genes were subcloned and sequenced.

In yet a further approach to isolate diverse P450 monooxygenase genes from strains R-922 and I-1529, a known P450 gene from another bacterium was used as a hybridization probe to identify cosmid clones containing homologous P450 genes from strains R-922 and I-1529. The *epoF* P450 gene from *Sorangium cellulosum* strain So ce90 that is involved in the synthesis of epothilones (Molnar *et al.*, *Chem Biol.* **7**(2):97-109, 2000) was used as a probe in this effort. Using the *epoF* P450 gene probe, one cosmid was identified from strain R-922 (clone LC), and three were identified from strain I-1529 (clones LA, LB, and EA). In each case, the homologous gene fragment was subcloned and sequenced, and found to code for P450 monooxygenase enzymes.

However, a comparison of the 17 polypeptide sequences identified in Example VII (below) failed to match any of these cloned genes. Two of the polypeptide sequences (namely, LVKDDPALLPR and AVHELMR) mapped to the region between the O<sub>2</sub> and heme binding domains, and so these should have identified any of the partial gene fragments derived by the PCR approach. Thus, the standard approaches based on the known PCR technique of Hyun *et al.*, *supra*, and using known P450 genes as hybridization probes failed to identify the gene that encodes the specific P450 monooxygenase responsible for the regioselective

oxidation of avermectin. Accordingly, it was determined that additional experimentation was required to isolate the gene encoding the P450 monooxygenase of the invention.

### EXAMPLE VII

#### Partial Sequencing of the P450 Monooxygenase from *Streptomyces* Strain R-922

Partial amino acid sequencing of the P450 monooxygenase from *Streptomyces* strain R-922 was carried out by the Friedrich Miescher Institute, Basel Switzerland. The protein of the dominant band on the SDS page was tryptically digested and the formed peptides separated and sequenced by mass spectrometry and Edman degradation (*see, generally, Zerbe-Burkhardt et al., J. Biol. Chem.* **273**:6508, 1998). The sequence of the following 17 peptides were found:

<u>Sequence</u>	<u>Sequence I.D. No.</u>
HPGEPNVMDPALITDPFTGYGALR	(SEQ ID NO:61)
FVNNPASPSLNYAPEDNPLTR	(SEQ ID NO:62)
LLTHYPDISLGIAPHEHLER	(SEQ ID NO:63)
VYLLGSILNYDAPDHTR	(SEQ ID NO:64)
TWGADLISMDPDR	(SEQ ID NO:65)
EALTDDLLSELIR	(SEQ ID NO:66)
FMDDSPVWLVR	(SEQ ID NO:67)
LMEMGLGLPEHLR	(SEQ ID NO:68)
VEQIADALLAR	(SEQ ID NO:69)
LVKDDPALLPR	(SEQ ID NO:70)
DDPALLPR	(SEQ ID NO:71)
TPLPGNWR	(SEQ ID NO:72)
LNSLPVR	(SEQ ID NO:73)
ITDLRPR	(SEQ ID NO:74)
EQGPVVR	(SEQ ID NO:75)
AVHELMR	(SEQ ID NO:76)

AFTAR (SEQ ID NO:77)

FEEVR (SEQ ID NO:78)

Alignment of these peptides to a selection of actinomycete P450 monooxygenase sequences indicated that all the peptides were fragments of a single P450 mono-oxygenase.

### EXAMPLE VIII

#### Cloning the P450 Monooxygenase Gene from Strain R-922 that Encodes the Enzyme Responsible for the Oxidation of Avermectin to 4''-Keto-Avermectin

PCR primers were designed by reverse translation from the amino acid sequences of several of the peptides derived from the P450 enzyme of strain R-922 (*see* Example VII and Table 2 below). Each of five forward primers (2aF, 2bF, 3F, 1F, and 7F) was paired with one reverse primer (5R) in PCR reactions with R-922 genomic DNA as a template. In each reaction, a DNA fragment of the expected size was produced.

**Table 2**

Primer	Primer sequence and the amino acid sequence to which they were designed*	Degen-eracy	Expected size (bp) **	SEQ ID NO:
2aF	P G E D N V M 5'-CCS GGS GAR CCS AAY GTS ATG-3'	64	600	79 80
2bF	A L I T D P F 5'-GCS CTS ATY ACS GAC CCS TTC-3'	32	580	81 82
3F	F M D D S P V W 5'-TTC ATG GAC GAC WSS CCS GTS TGG-3'	32	549	83 84
1F	L N Y D A P D H 5'-CTS AAY TAY GAC GCS CCS GAC CAC-3'	32	350	85 86
7F	V E Q I A D A L 5'-GTS GAR CAG ATY GCS GAC GCS CTS-3'	32	300	87 88
5R	D L I S M D P D 3'-CTG GAS TAR WSS TAC CTG GGS CTG-5'	64	---	89 90

\* Ambiguity codes: Y=C or T; R=A or G; S=C or G; W=A or T

\*\* Expected size of PCR product when the primer is when paired with primer 5R

The 580 and 600 bp PCR fragments generated by using primers (2bF and 5R) and (2aF and 5R), respectively, were cloned into the pCR-Blunt II-TOPO cloning plasmid (commercially available from Invitrogen, Carlsbad, CA) and transformed into *E. coli* strain TOP10 (Invitrogen, Carlsbad, CA). The inserted DNA fragments were then sequenced. Examination of the sequences revealed that the 600 and 580 bp fragments were identical in the 580 bp of sequence that they have in common. Also, there was a perfect match between the deduced amino acid sequence (SEQ ID NO:2) derived from the nucleotide sequence of the 600 bp and 580 bp fragments and the amino acid sequences of peptides isolated from the purified P450<sub>Ema1</sub> enzyme that aligned in this region of the isolated gene. This result strongly suggested that the gene fragments isolated in these clones are derived from the gene that encodes the P450<sub>Ema1</sub> enzyme that is responsible for the oxidation of avermectin to 4"-keto-avermectin.

The 600 bp PCR fragment produced using primers 2aF (SEQ ID No:80) and 5R (SEQ ID No:90) was used as a hybridization probe to a cosmid library of genomic DNA isolated from strain R-922 (cosmid library described in Example VI). Two cosmids named pPEH249 and pPEH250 were identified that hybridized strongly with the probe. The portion of each cosmid encoding the P450 enzyme was sequenced and the sequences were found to be identical between the two cosmids. The complete coding sequence of the *ema1* gene was identified (SEQ ID NO:1). The amino acid sequence of all polypeptide fragments from P450<sub>Ema1</sub> matched perfectly with the deduced amino acid sequence from the *ema1* gene. Comparison of the deduced amino acid sequence of the protein encoded by the *ema1* gene using BLASTP (Altschul *et al.*, *supra*) determined that the closest match in the databases is to a P450 monooxygenase from *S. thermotolerans* that has a role in the biosynthesis of carbomycin (Arisawa *et al.*, *Biosci. Biotech. Biochem.* **59**(4):582-588, 1995) and whose identity with *ema1* is only 49% (Identities = 202/409 (49%), Positives = 271/409 (65%), Gaps = 2/409 (0%)). In the Blast analysis, the following settings were employed:

```

BLASTP 2.0.10
Lambda      K      H
  0.322      0.140    0.428
Gapped
Lambda      K      H
  0.270      0.0470   0.230
Matrix: BLOSUM62
Gap Penalties: Existence: 11, Extension: 1
Number of Hits to DB: 375001765
Number of Sequences: 1271323
Number of extensions: 16451653
Number of successful extensions: 46738
Number of sequences better than 10.0: 2211
Number of HSP's better than 10.0 without gapping: 628
Number of HSP's successfully gapped in prelim test: 1583
Number of HSP's that attempted gapping in prelim test: 43251
Number of HSP's gapped (non-prelim): 2577
length of query: 430
length of database: 409,691,007
effective HSP length: 55
effective length of query: 375
effective length of database: 339,768,242
effective search space: 127413090750
effective search space used: 127413090750

```

A similar comparison of the nucleotide sequences of these two genes demonstrated that they are 65% identical at the nucleotide level. These results demonstrate that P450<sub>Emal</sub> is a new enzyme.

### EXAMPLE IX

#### Heterologous Expression of the *emal* Gene in *Streptomyces lividans* Strain ZX7

The coding sequence of the *emal* gene was fused to the thiostrepton-inducible promoter (*tipA*) (Murakami *et al.*, *J. Bacteriol.* **171**:1459-1466, 1989). The *tipA* promoter was derived from plasmid pSIT151 (Herron and Evans, *FEMS Microbiology Letters* **171**:215-221, 1999).

The fusion of the *tipA* promoter and the *emal* coding sequence was achieved by first amplifying the *emal* coding sequence with the following primers to introduce a PacI cloning site at the 5' end and a PmeI compatible end on the 3' end.

Forward Primer: The underlined sequence is a PacI recognition sequence; the sequence in bold-face type is the start of the coding sequence of *emal*.

5'-AGATTAATTAATATGTCGGAATTAATGAACTGTCCGTT-3' (SEQ ID NO:91)

Reverse Primer: The underlined sequence is half of a PmeI recognition sequence; the bold-face type sequence is the reverse complement of the *emaI* translation stop codon followed by the 3' end of the *emaI* coding sequence.

5'-AAACTCACCCCAACCGCACCGGCAGCGAGTTC-3" (SEQ ID NO:92)

The PacI-digested PCR fragment containing the *emaI* coding sequence was cloned into plasmid pTBBKA (see Figure 1) that was restricted (*i.e.*, digested) with PacI and PmeI, and the ligated plasmid transformed into *E. coli*. Four clones were sequenced. Three of the four contained the complete and correct *emaI* coding sequence. The fourth *emaI* gene clone contained a truncated version of the *emaI* gene. The full-length *emaI* gene encodes a protein that begins with the amino acid sequence MSELMNS (SEQ ID NO:93). The truncated gene encodes a protein that lacks the first 4 amino acids and begins with the second methionine residue. This gene has been named *emaIA*. The nucleotide and amino acid sequence of *emaIA* are provided as SEQ ID NO:33 and SEQ ID NO:34, respectively. The *emaI* and *emaIA* genes in these plasmids, pTBBKA-*emaI* and pTBBKA-*emaIA*, are in the correct juxtaposition with the *tipA* promoter to cause expression of the genes from this promoter.

Plasmid pTBBKA contains a gene from the *Streptomyces* insertion element IS117 that encodes an integrase that catalyzes site-specific integration of the plasmid into the chromosome of *Streptomyces* species (Henderson *et al.*, *Mol. Microbiol.* **3**:1307-1318, 1989 and Lydiate *et al.*, *Mol. Gen. Genet.* **203**:79-88, 1986). Since plasmid pTBBKA has only an *E. coli* replication origin and contains a mobilization site, it can be transferred from *E. coli* to *Streptomyces* strains by conjugation where it will not replicate. However, it is able to integrate into the chromosome due to the IS117 integrase and *Streptomyces* clones containing chromosomal integrations can be selected by resistance to kanamycin due to the plasmid-borne kanamycin resistance gene.

The *emaI* coding sequence was also cloned into other plasmids that are either replicative in *Streptomyces* or, like pTBBKA, integrate into the chromosome upon introduction into a *Streptomyces* host. For example, *emaI* was cloned into plasmid pEAA, which is similar to plasmid pTBBKA but the KpnI/PacI fragment containing the *tipA* promoter was replaced with the *ermE* gene promoter (Schmitt-John and Engels, *Appl*

*Microbiol Biotechnol.* **36**(4):493-498, 1992). In addition, pEAA does not contain the kanamycin resistance gene. The *ema1* gene was cloned into pEAA as a *PacI*/*PmeI* fragment to create plasmid pEAA-*ema1* in which the *ema1* gene is expressed from the constitutive *ermE* promoter.

Plasmid pTUA1A is a *Streptomyces-E.coli* shuttle plasmid (see Figure 2) that contains the *tipA* promoter. The *ema1* gene was also cloned into the *PacI*/*PmeI* site in plasmid pTUA1A to create plasmid pTUA-*ema1*.

The *ema1A* gene fragment was also ligated as a *PacI*/*PmeI* fragment into plasmids pTUA1A, and pEAA in the same way as the *ema1* gene fragment to create plasmids pTUA-*ema1A*, and pEAA-*ema1A*, respectively.

The pTBBKA, pTUA1A, and pEAA based plasmids containing the *ema1* or *ema1A* genes were introduced into *S. lividans* ZX7 and in each case transformants were obtained and verified (*S. lividans* strains ZX7::pTBBKA-*ema1* or *ema1A*, ZX7 (pTUA-*ema1* or -*ema1A*), and ZX7::pEAA-*ema1* or -*ema1A*, respectively).

Wild-type *Streptomyces lividans* strain ZX7 was tested and found to be incapable of the oxidation of avermectin to 4''-keto-avermectin. Transformed *S. lividans* strains ZX7::pTBBKA-*ema1*, ZX7::pTBBKA-*ema1A*, ZX7 (pTUA-*ema1*), ZX7 (pTUA-*ema1A*), ZX7::pEAA-*ema1*, and ZX7::pEAA-*ema1A* were each tested for the ability to oxidize avermectin to 4''-keto-avermectin using resting cells. To do this, the whole cell biocatalysis assay described above (including analysis method) was performed. Note that for the whole cell biocatalysis assay, transformed *Streptomyces lividans*, like strain R-922, was grown in PHG medium and, again like strain R-922, had a reaction time of 16 hours (*i.e.*, during which time the 500 mg transformed *Streptomyces lividans* wet cells in 10 ml of 50 mM potassium phosphate buffer, pH 7.0, were shaken at 160 rpm at 28°C in the presence of 15 µl of a solution of avermectin in isopropanol (30 mg/ml)).

In the presence of the inducer, thiostrepton (5 ug/ml), the *ema1*- or *ema1A*-containing strains ZX7::pTBBKA-*ema1*, ZX7::pTBBKA-*ema1A*, ZX7 (pTUA-*ema1*), ZX7 (pTUA-*ema1A*) were found to oxidize avermectin to 4''-keto-avermectin as evidenced by the appearance of the oxidized 4''-keto-avermectin compound (see Table 3).

### Table 3



Beispiel 1: <u>Strain</u>	% Conversion of Avermectin	
	2 hour	16 hour
<b><i>Streptomyces lividans</i> ZX7 + Plasmid<sup>1</sup></b>		
None	0	0
pTBBKA- <i>ema1A</i>	0.5 ±0.059	1.17 ±0.112
pTBBKA- <i>ema1</i>	0.21 ±0.0356	0.65 ±0.079
pTUA- <i>ema1</i>	20.96 ±1.044	42.0 ±2.5
pEAA- <i>ema1</i>	3.0 ±0.232	24.1 ±0.358
pTBBKA- <i>ema2</i>	4.79 ±0.096	9.57 ±0.423
pTUA- <i>ema2</i>	0.77 ±0.138	2.05 ±0.537
pEAA- <i>ema2</i>	0.0	1.73 ±3.00
pTBBKA- <i>ema1/fd233</i>	8.89 ±0.720	30.99 ±0.880
pTUA- <i>ema1/fd233</i>	23.29 ±0.854	61.2 ±3.548
pEAA- <i>ema1/fd233</i>	8.26 ±0.845	10.66 ±0.858
pTUA- <i>ema2/fd233</i>	1.85 ±0.861	6.40 ±1.918
<b><i>Pseudomonas putida</i> S12 + Plasmid</b>		
None		0
pRK- <i>ema1</i>	ND <sup>2</sup>	18
pRK- <i>ema1/fd233</i>	ND	32

<sup>1</sup>pTBBKA= IS117 integrase, tipA promoter; pTUA= replicative plasmid, tipA promoter;  
pEAA= IS117 integrase, ermE promoter

<sup>2</sup>Not Determined

These results conclusively demonstrate that the P450<sub>Ema1</sub> enzyme encoded by the *ema1* gene is responsible for the oxidation of avermectin to 4''-keto-avermectin in *S. tubercidicus* strain R-922. Furthermore, the data demonstrates that the *ema1A* gene that is 4 amino acids shorter on the N-terminus than the native *ema1* gene also encodes an active P450<sub>Ema1</sub> enzyme. As can be demonstrated by HPLC analysis, oxidation of avermectin to 4''-keto-avermectin by *S.*

*lividans* strain ZX7::pTBBKA-*ema1* following induction of *ema1* expression with 0, 0.5, or 5.0 µg/ml thiostrepton. is variable depending upon the amount of thiostrepton used to induce expression of *ema1*. Note that *S. lividans* strains ZX7::pEAA-*ema1* and ZX7::pEAA-*ema1A* (see Table 3) demonstrated this oxidation activity in the absence of thiostrepton since in these strains the *ema1* or *ema1A* genes are expressed from the *ermE* promoter that does not require induction.

### EXAMPLE X

#### Isolation of an *ema1*-Homologous Gene From *Streptomyces tubercidicus* Strain I-1529

*Streptomyces tubercidicus* strain I-1529 was also found to be active in biocatalysis of avermectin to form the 4''-keto-avermectin derivative. The cosmid library from strain I-1529, described in Example VI, was probed at the high stringency conditions of Church and Gilbert (Church and Gilbert, *Proc. Natl. Acad. Sci. USA* **81**:1991-1995, 1984) with the 600 bp *ema1* PCR fragment produced using primers 2aF (SEQ ID No:80) and 5R (SEQ ID No:90) described previously to identify clones containing the *ema1* homolog from strain I-1529. Three strongly hybridizing cosmids were identified. The P450 gene regions in two of the cosmids, pPEH252 and pPEH253, were sequenced and found to be identical. Analysis of the DNA sequence revealed the presence of a gene with high homology to the *ema1* gene of strain R-922. A comparison of the deduced amino acid sequence of Ema2 (*i.e.*, P450<sub>Ema2</sub>), Ema1 (*i.e.*, P450<sub>Ema1</sub>), and a P450 monooxygenase from *Streptomyces thermotolerans* that is involved in the biosynthesis of carbomycin (Carb-450) (GenBank Accession No. D30759). demonstrated that all of the unique P450 gene fragments from both the R-922 and I-1529 strains were derived from P450 genes and encoded the region between the O<sub>2</sub>-binding and heme-binding domains.

The gene from *Streptomyces tubercidicus* strain I-1529, named *ema2*, encodes an enzyme with 90% identity at the amino acid level and 90.6% identity at the nucleotide level to the P450<sub>Ema1</sub> enzyme. The nucleotide sequence of the *ema2* gene and the deduced amino acid sequence of P450<sub>Ema2</sub> are provided in SEQ ID NO:3 and SEQ ID NO:4, respectively.

The *ema2* coding sequence was cloned in the same manner as the *ema1* and *ema1A* genes into plasmids pTBBKA, pTUA1A, and pEAA such that the coding sequence was

functionally fused to the *tipA* or *ermE*\* promoter in these plasmids. The resulting plasmids, pTBBKA-*ema2*, pTUA-*ema2*, and pEAA-*ema2* were transferred from *E. coli* to *S. lividans* ZX7 by conjugation to create strains ZX7::TBBKA-*ema2* and ZX7 (pTUA-*ema2*), and ZX7::pEAA-*ema2* containing the *ema2* gene integrated into the chromosome or maintained on a plasmid.

Strains ZX7::TBBKA-*ema2*, ZX7 (pTUA-*ema2*), and ZX7::pEAA-*ema2* were next tested for the ability to oxidize avermectin to 4''-keto-avermectin. The *ema2* gene was also shown to provide biocatalysis activity, although at a lower level compared to the *ema1* gene (see Table 3).

These results demonstrate that the *ema2* gene from *S. tubercidicus* strain I-1529 also encodes a P450 enzyme (P450<sub>Ema2</sub>) capable of oxidizing avermectin to 4''-keto-avermectin.

### EXAMPLE XI

#### Characterization of *ema1* Homologs From Other Biocatalysis Strains

Seventeen *Streptomyces* sp. strains, including strains R-922 and I-1529, were identified that are capable of catalyzing the regiospecific oxidation of the 4''-carbinol of avermectin to a ketone. Next, the isolation and characterization of the genes encoding the biocatalysis enzyme from all of these strains was accomplished.

To do this, genomic DNA was isolated from the strains and was evaluated by restriction with several restriction endonucleases and Southern hybridization with the *ema1* gene. A specific restriction endonuclease was identified for each DNA that would generate a single DNA fragment of a defined size to which the *ema1* gene hybridizes. For each strain, there was only one strongly hybridizing DNA fragment, thus suggesting that other P450 genes were not detected under the high stringency hybridization conditions used in these experiments. Each DNA was digested with the appropriate restriction endonuclease, and the DNA was subjected to agarose gel electrophoresis. DNA in a narrow size range that included the size of the *ema1*-hybridizing fragment was excised from the gel. The size selected DNA was ligated into an appropriate cloning plasmid and this ligated plasmid was used to transform *E. coli*. The *E. coli* clones from each experiment were screened by colony hybridization with the *ema1* gene fragment to identify clones containing the *ema1*-homologous DNA fragment.

The nucleotide sequence of the cloned DNA in each *ema1*-homologous clone was determined and examined for the presence of a gene encoding a P450 enzyme with homology to *ema1*. In this way, *ema1*-homologous genes were isolated from 14 of the 15 other active strains. The nucleotide and deduced amino acid sequences of these are referenced in Table 4 as SEQ ID NOS:5-32 and 94-95. The relationship of these enzymes can be shown in the form of a phylogenetic tree. Such a phylogenetic tree can be generated using the commercially available GCG Wisconsin software program version 1.0 (Madison, WI).

**Table 4**

Strain Number		Classification	SEQ ID NO (nucleotide and amino acid, respectively)
R-0922	<i>ema1</i>	<i>Streptomyces tubercidicus</i>	2. 1 and 2
I-1529	<i>ema2</i>	<i>Streptomyces tubercidicus</i>	3 and 4
1053	<i>ema3</i>	<i>Streptomyces rimosus</i>	5 and 6
R-0401	<i>ema4</i>	<i>Streptomyces lydicus</i>	7 and 8
I-1525	<i>ema5</i>	<i>Streptomyces sp.</i>	9 and 10
DSM-40241	<i>ema6</i>	<i>Streptomyces chattanoogensis</i> *	3. 11 and 12
IHS-0435	<i>ema7</i>	<i>Streptomyces sp.</i>	13 and 14
C-00083	<i>ema8</i>	<i>Streptomyces albofaciens</i>	15 and 16
MAAG-7479	<i>ema9</i>	<i>Streptomyces platensis</i>	17 and 18
A/96-1208710	<i>ema10</i>	<i>Streptomyces kasugaensis</i>	4. 19 and 20
R-2374	<i>ema11</i>	<i>Streptomyces rimosus</i>	21 and 22
MAAG-7027	<i>ema12</i>	<i>Streptomyces tubercidicus</i>	5. 23 and 24
Tue-3077	<i>ema13</i>	<i>Streptomyces platensis</i>	25 and 26
I-1548	<i>ema14</i>	<i>Streptomyces platensis</i>	27 and 28
NRRL-2433	<i>ema15</i>	<i>Streptomyces lydicus</i>	6. 29 and 30
MAAG-0114	<i>ema16</i>	<i>Streptomyces lydicus</i>	31 and 32
DSM-40261	<i>ema17</i>	<i>Streptomyces tubercidicus</i>	94 and 95

\* This strain was shown to be in the *chattanoogensis* species by 16s rDNA analysis; however, classical taxonomic methods used by the German culture collection (DSMZ) showed it to be *saraceticus*.

## EXAMPLE XII

### Construction of His-tagged *ema1* and *ema1* Homologs to Facilitate Enzyme Purification

In order to purify the P450<sub>Ema1</sub> enzyme and the P450 enzymes encoded by the *ema1* homologs from other biocatalysis strains, each of the P450 genes was cloned into the *E. coli* expression plasmid pET-28b(+) (commercially available from Novagen, Madison, WI). The pET-28 plasmids are designed to facilitate His-tag fusions at either the N-, or C-terminus and to provide strong expression of the genes in *E. coli* from the T7 phage promoter. In many cases, the coding sequence of the *ema* genes begins with the sequence ATGT. These genes were amplified by PCR such that the primers on the 5' end incorporated a PciI recognition site (5' ATATGT 3') at the 5' terminus. The last four bases of the PciI site correspond to the ATGT at the beginning of the *ema* gene coding sequence.

PCR primers at the 3' end of the genes were designed to remove the translation stop codon at the end of the *ema* gene coding sequence and to add an XhoI recognition site to the 3' terminus. The resulting PCR fragments were restricted with PciI and XhoI to generate PciI ends at the 5' termini and XhoI ends at the 3' termini, thereby facilitating cloning of the fragments into pET-28b(+) previously restricted with NcoI and XhoI. Since PciI and NcoI ends are compatible, the fragments were cloned into pET-28b(+) in the proper orientation to the T7 promoter and ribosome binding site in the plasmid to provide expression of the genes.

At the 3' end of each *ema* gene, the coding sequence was fused in frame at the XhoI site to the His-tag sequence followed by a translation stop codon. This results in the production of an Ema enzyme with six histidine residues added to the C-terminus to facilitate purification on nickel columns.

In the case of *ema* genes in which the ATG translation initiation codon is not followed by a T nucleotide, the *ema* genes were amplified by PCR using a different strategy for the 5' end. The primers at the 5' end were designed to incorporate a C immediately preceding the ATG translation initiation codon and the primers at the 3' end were the same as described above. The PCR fragments that were amplified were restricted with XhoI to create an XhoI end at the 3'-terminus and the 5' end was left as a blunt end. These fragments were cloned into pET-28b(+) that had been restricted with NcoI, but the NcoI ends were made blunt-ended by treatment with mung bean exonuclease, and restricted with XhoI.

In this manner, the *ema* genes were cloned into pET-28b(+) to create a functional fusion with the T7 promoter and the His-tag at the C-terminus as described previously. All His-tagged *ema* genes were sequenced to ensure that no errors were introduced by PCR.

Large amounts of the P450<sub>Ema1</sub> and P450<sub>Ema2</sub> enzymes were isolated and purified by standard protocols. *E. coli* strain BL21 DE3 (commercially available from Invitrogen; Carlsbad, CA) containing the T7 RNA polymerase gene under the control of the inducible *tac* promoter and the appropriate pET-28/*ema* plasmid was cultured and the cells were harvested and lysed. The lysates were applied to Ni-NTA columns (commercially available from Qiagen Inc., Valencia, CA) and the protein were purified according to the procedure recommended by the manufacturer.

Purified His-tagged P450<sub>Ema1</sub> and P450<sub>Ema2</sub> were highly active in *in vitro* activity assays as evidenced by a high rate of conversion of avermectin to 4''-keto-avemectin.

### EXAMPLE XIII

#### Expression of *ema1* in *Pseudomonas*

The *ema1* gene constructs were next introduced into *P. putida* (wildtype *P. putida* commercially available from the American Type Culture Collection, Manassas, Virginia; ATCC Nos. 700801 and 17453). The *ema1* and *ema1/fd233* gene fragments were cloned as PacI/PmeI fragments into the plasmid pUK21 (Viera and Messing, *Gene* **100**:189-194, 1991). The fragments were cloned into a position located between the *tac* promoter (P<sub>tac</sub>) and terminator (T<sub>tac</sub>) on pUK21 in the proper orientation for expression from the *tac* promoter. The P<sub>tac</sub>-*ema1*-T<sub>tac</sub> and P<sub>tac</sub>-*ema1/fd233*-T<sub>tac</sub> gene fragments were removed from pUK21 as BglII fragments and these were cloned into the broad host-range, transmissible plasmid, pRK290 (Ditta *et al.*, *Proc. Natl. Acad. Sci. USA* **77**:7347-7351, 1980) to create plasmids pRK-*ema1* and pRK-*ema1/fd233* (Figure 3). These plasmids were introduced into *P. putida* strains ATCC 700801 and ATCC 17453 by conjugal transfer from *E. coli* hosts by standard methodology (Ditta *et al.*, *Proc. Natl. Acad. Sci. USA* **77**:7347-7351, 1980).

*P. putida* ATCC 700801 and ATCC 17453 containing plasmids pRK-*ema1* or pRK-*ema1/fd233* were tested for the ability to catalyze the oxidation of avermectin. The results shown in Table 3 demonstrate that these strains are able to catalyze this reaction.

### EXAMPLE XIV

Identification of Genes Encoding Ferredoxins That Are Active With the  
P450<sub>Ema1</sub> Monooxygenase

P450 monooxygenases require two electrons for each hydroxylation reaction catalyzed (Mueller *et al.*, "Twenty-five years of P450<sub>cam</sub> research: Mechanistic Insights into Oxygenase Catalysis." Cytochrome P450, 2<sup>nd</sup> Edition, P.R. Ortiz de Montellano (ed.), pp. 83-124; Plenum Press, NY 1995). These electrons are transferred to the P450 monooxygenase one at a time by a ferredoxin. The electrons are ultimately derived from NAD(P)H and are passed to the ferredoxin by a ferredoxin reductase. Specific P-450 monooxygenase enzymes have a higher activity when they interact with a specific ferredoxin. In many cases, the gene encoding a ferredoxin that interacts specifically with a given P450 monooxygenase is located adjacent to the gene encoding the P450 enzyme.

As described above, in addition to the *ema1* gene, four P450 genes from strain R-922 and seven P450 genes from strain I-1529 (*see* Example VI) were isolated and sequenced. In some of these, there was sufficient sequence information about the DNA flanking the P-450 genes to look for the presence of associated ferredoxin genes. By this approach, two unique ferredoxin genes were identified from each of the two strains. Ferredoxin genes *fd229* and *fd230* were identified from strain R-922, and *fd233* and *fdEA* were identified from strain I-1529. In addition, a ferredoxin reductase gene was found to reside adjacent to the *fdEA* gene from strain I-1529.

In order to test the biological activity of each of these ferredoxins in combination with P450<sub>Ema1</sub>, each individual ferredoxin gene was amplified by PCR to produce a gene fragment that included a blunt 5'-end, the native ribosome-binding site and ferredoxin gene coding sequence, and a PmeI restriction site on the 3'-end. Each such ferredoxin gene fragment was cloned into the PmeI site located 3' to the *ema1* gene in plasmid pTUA-*ema1*. In this way, artificial operons consisting of the *ema1* gene and one of the ferredoxin genes operably linked to a functional promoter were created.

In the case of the *fdEA* ferredoxin gene in which a ferredoxin reductase gene, *freEA*, was found to be located adjacent to the *fdEA* gene, a DNA fragment containing both the *fdEA* and *freEA* genes was generated by a similar PCR strategy. This gene fragment was also cloned in the PmeI site of plasmid pTUA-*ema1* as described for the other ferredoxin genes.

Each *ema1*-ferredoxin gene combination was tested for biological activity by introduction of the individual *ema1*-ferredoxin gene plasmids into *S. lividans* strain ZX7. The biocatalysis activity derived from each plasmid in *S. lividans* was determined. Of the four different constructs, only the ferredoxin gene *fd233* derived from strain I-1529 provided increased activity when compared to the expression of *ema1* alone in the same plasmid and host background (see Table 3). The pTUA-*ema1/fd233* plasmid in *S. lividans* provided approximately 1.5 to 3- fold higher activity compared to the pTUA-*ema1* plasmid. The other three plasmids containing the other ferredoxin genes gave results essentially the same as the plasmid with only the *ema1* gene. Likewise, the pTUA-*ema1/fdEA/freEA* plasmid did not yield results different from those of pTUA-*ema1*. The nucleotide and deduced amino acid sequences of the *fd233* gene are shown in SEQ ID NOs:35 and 36, respectively.

A BLAST analysis of the nucleotide and amino acid sequences of *fd233* revealed that the closest matches were to ferredoxins from *S. coelicolor* (GenBank Accession AL445945) and *S. lividans* (GenBank Accession AF072709). At the nucleotide level, *fd233* shares 80 and 79.8 % identity with the ferredoxin genes from *S. coelicolor* and *S. lividans*, respectively. At the peptide level, *fd233* shares 79.4 and 77.8% identity with the ferredoxins from *S. coelicolor* and *S. lividans*, respectively.

Since *fd233* is derived from strain I-1529 and *ema1* is from strain R-922, the proteins encoded by the two genes cannot interact with each other in nature. In an approach designed to identify a ferredoxin gene from strain R-922 that is homologous to the *fd233* gene and that might encode a ferredoxin that interacts optimally with the P450<sub>Ema1</sub>, the *fd233* gene was used as a hybridization probe to a gene library of DNA from strain R-922. A strongly hybridizing cosmid, pPEH232, was identified and the hybridizing DNA was cloned and sequenced. Comparison of the deduced amino acid sequences from *fd233* and the ferredoxin gene on cosmid pPEH232, *fd232*, revealed that they differed in only a single amino acid.

In a similar manner, plasmid pTUA-*ema1-fd232* was constructed and tested in *S. lividans* ZX7. This plasmid gave similar results as those obtained with plasmid pTUA-*ema1-fd233* (see Table 3). The nucleotide and deduced amino acid sequences of *fd232* are shown in SEQ ID NOs:37 and 38, respectively.

The *ema1-fd233* operon was also subcloned, as a PacI-PmeI fragment, into pTBBKA and pEAA that had been digested with the same restriction enzymes. *S. lividans*



ZX7::pTBBKA-*ema1*-*fd233*, and *S. lividans* ZX7::pEAA-*ema1*-*fd233* were tested in the avermectin conversion assay and found to have higher activities than the strains harboring the *ema1* gene alone in the comparable plasmids (see Table 3).

### EXAMPLE XV

#### Heterologous Expression of P450<sub>Ema1</sub> and P450<sub>Ema2</sub> in Other Cells

The expression constructs pRK-*ema1* (Example XIII) and pRK-*ema2* (created in a way analogous to that described in Example XIII for pRK-*ema1*) were mobilized by conjugation into three fluorescent soil *Pseudomonas* strains. Conjugation was performed according to standard methods (Ditta *et al.*, *Proc. Natl. Acad. Sci. USA* **77**:7347-7351, 1980). The strains were: *P. fluorescens* MOCG134, *P. fluorescens* Pf-5, and *P. fluorescens* CHAO. Standard resting cell assays for the conversion of avermectin to 4'-ketoavermectin were conducted for each of the transconjugants. For strains Pf-5 and CHAO, the levels of conversion were below the detection limit. Strain MOCG134 yielded 3% conversion for *ema1* and 5% for *ema2*.

In addition, the constructs listed in the Table 5 were introduced into *Streptomyces avermitilis* MOS-0001 by protoplast-mediated transformation (Kieser, T.; Bibb, M.J.; Buttner, M.J.; Chater, K.F.; Hopwood, D.A. (eds.): *Practical Streptomyces Genetics*. The John Innes Foundation, Norwich (England), 2000), (Stutzman-Engwall, K. *et al.* (1999) *Streptomyces avermitilis* gene directing the ratio of B<sub>2</sub>:B<sub>1</sub> avermectins, WO 99/41389).

**Table 5**

<b>Construct</b>	<b>% Conversion of avermectin, 16 hrs</b>
None	0
pTBBKA- <i>ema1</i>	10.90 +/- 3.48
pTUA- <i>ema1</i>	5.326 +/- 2.19
pEAA- <i>ema1</i>	6.74 +/- 0.08
pTBBKA- <i>ema1A</i> / <i>fd233</i>	28.50 +/- 0.20
pTUA- <i>ema1A</i> / <i>fd233</i>	23.97 +/- 5.95

Wild-type *Str. avermitilis* MOS-0001 was tested and found to be incapable of the oxidation of avermectin to 4''-ketoavermectin.

Transformed *S. avermitilis* strains MOS-0001::pTBBKA-*ema1*, MOS-0001 (pTUA-*ema1*), MOS-0001::pEAA-*ema1*, MOS-0001::pTBBKA-*ema1A/fd233*, and MOS-0001 (pTUA-*ema1A/fd233*) were each tested for their ability to oxidize avermectin to 4''-keto-avermectin using resting cells. To do this, the whole cell biocatalysis assay described above (including analysis method) was performed. Note that for the whole cell biocatalysis assay, transformed *Streptomyces avermitilis*, like strain R-922, was grown in PHG medium and, again like strain R-922, had a reaction time of 16 hours (*i.e.*, during which time the 500 mg transformed *Streptomyces avermitilis* wet cells in 10 ml of 50 mM potassium phosphate buffer, pH 7.0, were shaken at 160 rpm at 28°C in the presence of 15 µl of a solution of avermectin in isopropanol (30 mg/ml)).

As shown in Table 5, in the presence of the inducer, thiostrepton (5 µg/ml), the *ema1*- or *ema1A/fd233*-containing strains MOS-0001::pTBBKA-*ema1*, MOS-0001::pTBBKA-*ema1A/fd233*, MOS-0001 (pTUA-*ema1*), MOS-0001 (pTUA-*ema1A/fd233*) were found to oxidize avermectin to 4''-keto-avermectin as evidenced by the appearance of the oxidized 4''-keto-avermectin compound. Note that the *S. avermitilis* strain MOS-0001::pEAA-*ema1* demonstrated this oxidation activity in the absence of thiostrepton since in this strain the *ema1* gene is expressed from the *ermE* promoter that does not require induction.

Thus, expression of the *ema1* P450 monooxygenase gene in various *Streptomyces* and *Pseudomonas* strains provided recombinant cells that were able to convert avermectin to 4''-ketoavermectin in resting cell assays.

Next, expression and activity of P450<sub>Ema1</sub> monooxygenase was tested in *E. coli*. To do this, the *ema1* gene was cloned into the *E. coli* expression plasmid pET-28b(+) (commercially available from Novagen, Madison, WI) as described previously. *E. coli* strain BL21 DE3 (commercially available from Invitrogen; Carlsbad, CA) that contains the T7 RNA polymerase gene under control of the inducible *tac* promoter and the pET-28/*ema1* plasmid was cultured in 50 ml LB medium containing 5 mg/l kanamycin in a 250-ml flask with one baffle, for 16 hours at 37°C, with shaking at 130 rpm. 0.5 ml of this culture was used to inoculate 500 ml LB medium with 5 mg/l kanamycin in a 2-liter flask with one baffle, and the

culture was incubated for 4 hours at 37°C followed by 4 hours and 30°C, with shaking at 130 rpm throughout. The cells were harvested by centrifugation, washed in 50 mM potassium phosphate buffer, and centrifuged again.

For the resting cell assays, 90 mg wet cells were weighed into deep-well plates in triplicate and resuspended in 0.5 ml 50 mM potassium phosphate buffer. For cell-free extracts, 4 grams wet cells in 8 ml disruption buffer were disrupted in French press.

For the resting cell assays, 5 µl of substrate (2.5 mg/ml in 2-propanol) was added to the cell suspension. The plate was sealed with air permeable foil, and the reaction was incubated on an orbital shaker at 1000 rpm at 28°C for 22 hours. No conversion of avermectin to 4''-ketoavermectin was detected.

For the cell-free assays, 100 µl cell free extract, 1µl substrate solution (20 mg/ml) in 2-propanol, 5 µl 100 mM NADPH, 10 µl ferredoxin, 10 µl ferredoxin reductase, and 374 µl potassium phosphate buffer pH 7.0 were added as described in Example III, and the assay was incubated at 30°C with shaking at 600 rpm for 20 hours. 9.2% +/- 0.3% of avermectin was converted to 4''-ketoavermectin.

Thus, expression of the *ema1* gene in *E. coli* resulted in the production of the active Ema1 P450 monooxygenase enzyme which, when purified from the cells, was able to convert avermectin to 4''-ketoavermectin.

## EXAMPLE XVI

### Identification and Cloning of Genes Encoding Ferredoxin Reductases that Support Increased Activity of the P450<sub>Ema1</sub> Monooxygenase

The electron transport pathway that supports the activity of P450 monooxygenases also includes ferredoxin reductases. These proteins donate electrons to the ferredoxin and, as is the case with ferredoxins and P450 monooxygenases, specific ferredoxin reductases are known to be better electron donors for certain ferredoxins than others.

According, a number of ferredoxin reductase genes from *Streptomyces* strains were cloned and were evaluated for their impacts on the biocatalysis reaction. To do this, numerous bacterial ferredoxin reductase (Fre) protein sequences were retrieved from NCBI and aligned with the program Pretty from the GCG package. Two conserved regions,

approximately 266 amino acid residues apart, were used to make degenerate oligonucleotides for PCR. The forward primer (CGSCCSCCCTSWSSAAS (SEQ ID NO:96; where “S” is C or G; and “W” is A or G)) and the reverse primer (SASSGCSTTSBCCCARTGYTC (SEQ ID NO:97; where “S” is C or G; “B” is C, G, or T; “R” is A or G; and “Y” is C or T)) were used to amplify 800 bp products from the biocatalytically active *Streptomyces* strains R-922 and I-1529. These pools of products were cloned into TOPO TA cloning vectors (commercially available from Invitrogen Inc., Carlsbad, CA), and 20 clones each from R922 and I-1529 were sequenced according to standard methods (see, *e.g.*, Current Protocols in Molecular Biology, eds. Ausubel *et al.*, John Wiley & Sons, Inc. 2000). Sequencing revealed that 4 unique *fre* gene fragments were isolated from the strains: three from R922 (*fre3*, *fre12*, *fre14*) and one from I-1529 (*fre16*). The *fre3*, *fre12*, *fre14*, and *fre16* gene fragments were used as probes to identify full-length ferredoxin reductases from genomic clone banks of *Streptomyces* strains R922 and I-1529. By this approach, the complete coding sequence of each of the 4 different *fre* genes was cloned and sequenced. The nucleic acid and amino acid sequences are provided as follows: *fre3* (SEQ ID NOs:98 and 99); *fre12* (SEQ ID NOs:100 and 101); *fre14* (SEQ ID NOs:102 and 103); and *fre16* (SEQ ID NOs:104 and 105).

In order to assess the biological activity of each *fre* gene in relation to the activity of Ema1, each gene was inserted into the *ema1/fd233* operon described above, 3' to the *fd233* gene. This resulted in the formation of artificial operons consisting of the *ema1*, *fd233*, and individual *fre* genes that were expressed from the same promoter. The *ema1/fd233/fre* operons were cloned into the *Pseudomonas* plasmid pRK290 and introduced into 3 different *P. putida* strains. These strains were then analysed for Ema1 biocatalysis activity using the whole cell assay and one of the genes, the *fre* gene *fre16* from strain I-1529, was found to increase the activity of P450<sub>Ema1</sub> monooxygenase by approximately 2-fold. This effect was strain specific, as it was seen only in one of the *P. putida* strains, ATCC Desposit No. 17453, and not in the other two. In *P. putida* strain ATCC 17453, the presence of *fre* gene *fre16* resulted in 44% conversion of avermectin to 4"-keto-avermectin, as compared to 23% without this gene. The other *fre* genes had no impact on the biocatalysis activity in any of the *P. putida* strains tested.

In a similar approach, each of the *ema1/fd233/fre* operons were cloned into the *Streptomyces* plasmids pTUA, pTBBKA, and pEAA, and introduced into *S. lividans* strain

ZX7. In each case there was no impact in *S. lividans* by any of the *fre* genes on biocatalysis activity.

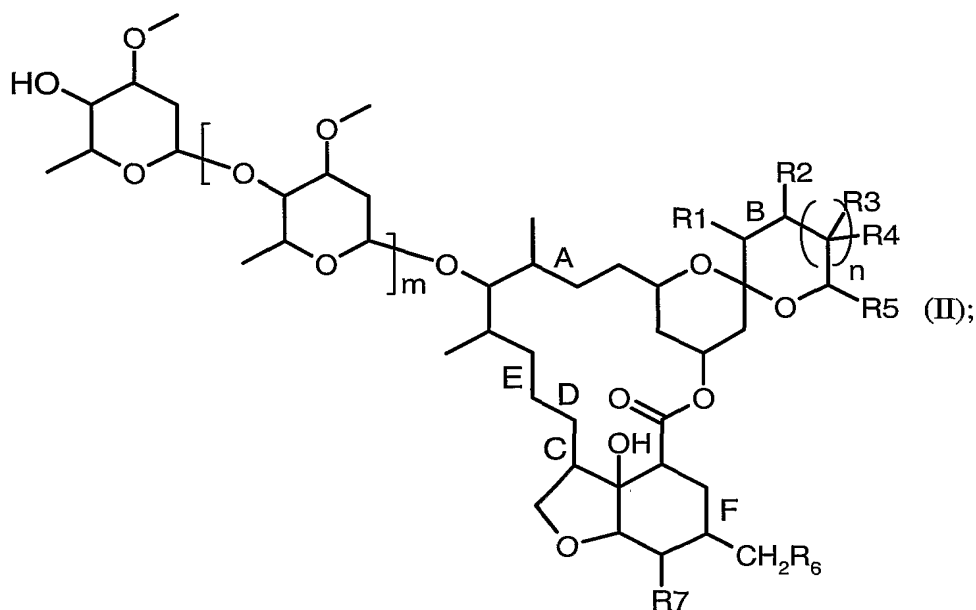
### EQUIVALENTS

Those skilled in the art will recognize, or be able to ascertain, using no more than routine experimentation, numerous equivalents to the specific substances and procedures described herein. Such equivalents are considered to be within the scope of this invention.

The patent and scientific literature referred to herein establishes knowledge that is available to those with skill in the art. The issued patents, applications, and references, including GenBank database sequences, that are cited herein are hereby incorporated by reference to the same extent as if each was specifically and individually indicated to be incorporated by reference. Any conflict between any reference cited herein and the specific teachings of this specification shall be resolved in favor of the latter. Likewise, any conflict between an art-understood definition of a word or phrase and a definition of the word or phrase as specifically taught in this specification shall be resolved in favor of the latter.

**What is claimed is:**

1. A purified nucleic acid molecule encoding a polypeptide that exhibits an enzymatic  
 5 activity of a P450 monooxygenase and is capable of regioselectively oxidizing the alcohol  
 at position 4" of a compound of formular (II)



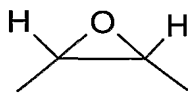
wherein

10  $R_1$ - $R_7$  represent, independently of each other hydrogen or a substituent;

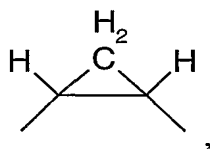
$m$  is 0, 1 or 2;

$n$  is 0, 1, 2 or 3; and

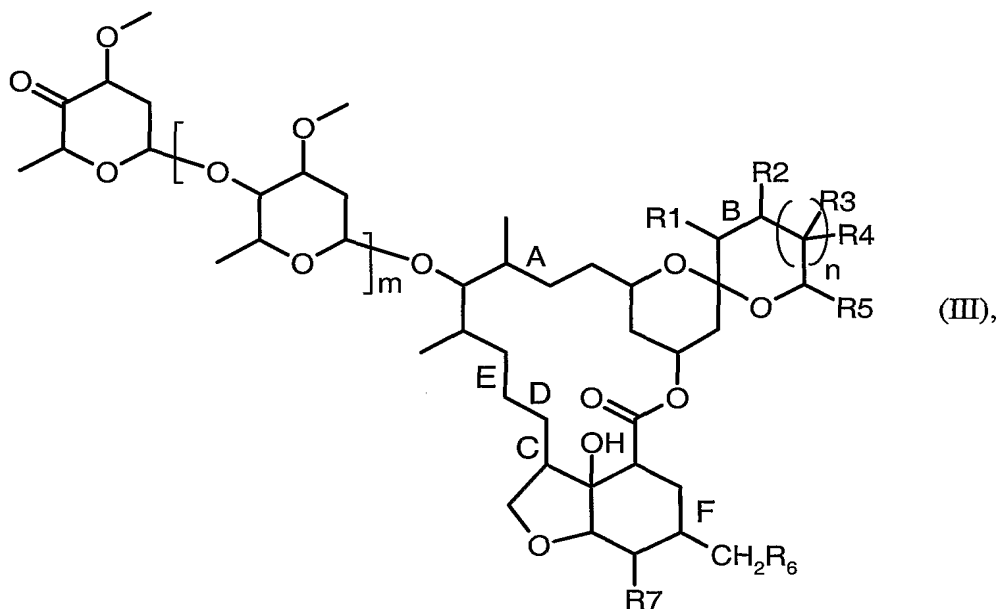
the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two  
 adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a  
 15 epoxide bridge of the formula



, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,  
in order to produce a compound of the formula (III)



wherein

R<sub>1</sub>-R<sub>7</sub>, m, n, A, B, C, D, E and F have the same meanings as given for formula (II) above.

2. The nucleic acid molecule of claim 1, comprising a nucleic acid sequence that encodes a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''keto-avermectin.
3. The nucleic acid molecule of claims 1 or 2, comprising a nucleic acid sequence that encodes a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase, which polypeptide is substantially similar, and has between at least 50%, and 99% amino acid sequence identity to the polypeptide of SEQ ID NO:2.

4. The nucleic acid molecule of claim 3 comprising a nucleotide sequence
- a) as given in SEQ ID NO:1;
  - b) having substantial similarity to (a);
  - c) capable of hybridizing to (a) or the complement thereof;
  - 5 d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, or the complement thereof;
  - e) complementary to (a), (b) or (c);
  - f) which is the reverse complement of (a), (b) or (c); or
  - 10 g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin. .
5. The nucleic acid molecule of claims 1 or 2, comprising a nucleic acid sequence that is
- 15 at least 66 % identical to SEQ ID NO:1.
6. The nucleic acid molecule of claims 1 or 2, comprising a nucleic acid sequence that encodes a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase, which polypeptide is substantially similar, and has at least between 60%, and 99%
- 20 amino acid sequence identity to the polypeptide of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.
- 25
7. The nucleic acid molecule of claims 1 or 2, comprising a nucleic acid sequence that encodes a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase, which polypeptide is immunologically reactive with antibodies raised against a polypeptide of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8,
- 30 SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18,



SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28,  
SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, or SEQ ID NO:95.

8. The nucleic acid molecule of claims 1 or 2 comprising a nucleotide sequence

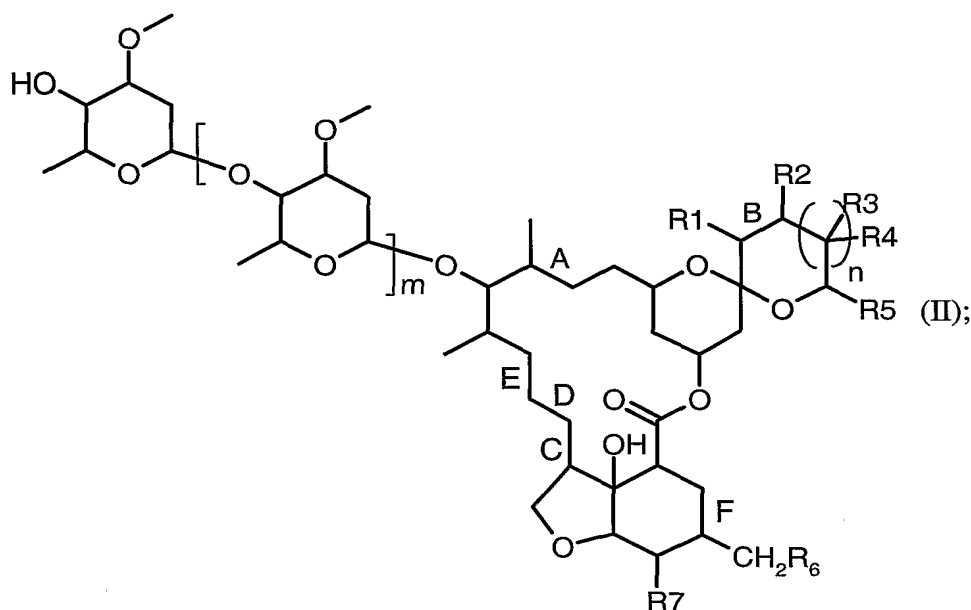
- 5 a) as given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID  
NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID  
NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID  
NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94;
- b) having substantial similarity to (a);
- 10 c) capable of hybridizing to (a) or the complement thereof;
- d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more  
consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, SEQ ID  
NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13,  
SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23,  
15 SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or  
SEQ ID NO:94 or the complement thereof;
- e) complementary to (a), (b) or (c); and
- f) which is the reverse complement of (a), (b) or (c).
- g) . which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that  
20 still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively  
oxidizes avermectin to 4"-keto-avermectin.

9. The nucleic acid molecule of claim 8, comprising a nucleic acid sequence selected from  
the group consisting of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ  
25 ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID  
NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID  
NO:29, SEQ ID NO:31, SEQ ID NO:33, and SEQ ID NO:94.

10. The nucleic acid molecule of anyone of claims 1 to 9, wherein the nucleic acid molecule  
30 is isolated from a *Streptomyces* strain.

11. The nucleic acid molecule of anyone of claims 1 to 10 further comprising a nucleic acid sequence encoding a tag which is linked to the P450 monooxygenase via a covalent bond.

12. A polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II)



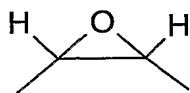
10            wherein

R<sub>1</sub>-R<sub>7</sub> represent, independently of each other hydrogen or a substituent;

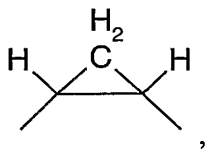
$m$  is 0, 1 or 2;

$n$  is 0, 1, 2 or 3; and

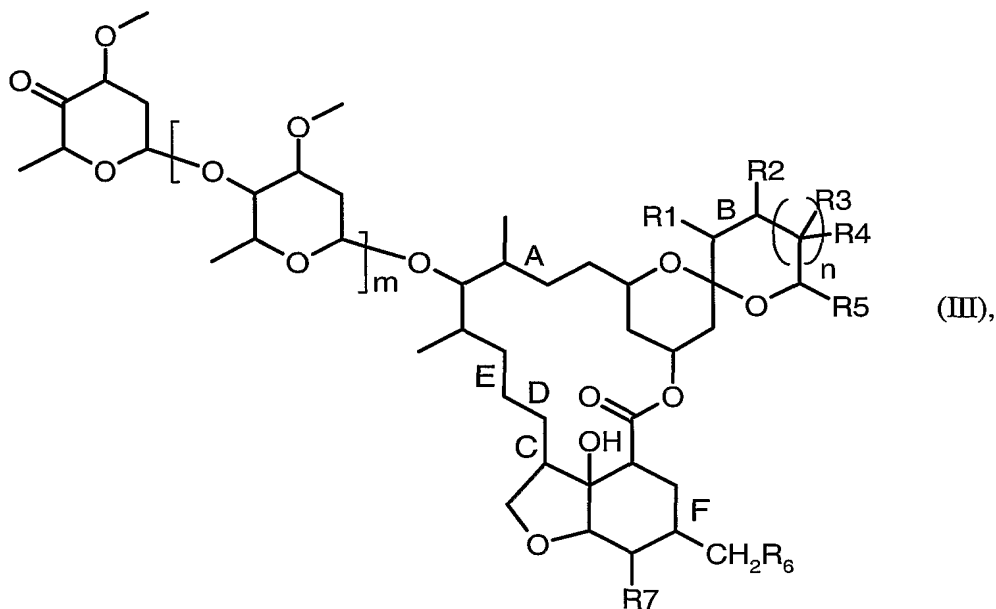
the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,  
in order to produce a compound of the formula (III)



wherein

$R_1$ - $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the same meanings as given for formula (II) above.

13. A polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''keto-avermectin.

14. The polypeptide of claims 12 or 13 that comprises an amino acid sequence that is encoded by a nucleic acid molecule

a) as given in SEQ ID NO:1 or the complement thereof;

b) having substantial similarity to (a);

c) capable of hybridizing to (a) or the complement thereof;

- d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, or the complement thereof;
- e) complementary to (a), (b) or (c);
- 5 f) which is the reverse complement of (a), (b) or (c); or
- g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''-keto-avermectin.
- 10 15. The polypeptide of claims 12 to 14, comprising an amino acid sequence that is at least 50% identical to SEQ ID NO:2.
16. The polypeptide of claims 12 or 13 comprising an amino acid sequence that is encoded by a nucleic acid molecule
- 15 a) as given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof;
- b) having substantial similarity to (a);
- 20 c) capable of hybridizing to (a) or the complement thereof;
- d) capable of hybridizing to a nucleic acid molecule comprising 50 to 200 or more consecutive nucleotides of a nucleotide sequence given in SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:15, SEQ ID NO:17, SEQ ID NO:19, SEQ ID NO:21, SEQ ID NO:23, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:31, SEQ ID NO:33, or SEQ ID NO:94 or the complement thereof, or the complement thereof;
- 25 e) complementary to (a), (b) or (c);
- f) which is the reverse complement of (a), (b) or (c); or

g) which is a functional part of (a), (b), (c), (d), (e) or (f) encoding a polypeptide that still exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4"-keto-avermectin.

- 5      17. The polypeptide of claim 16, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, SEQ ID NO:34, and SEQ ID NO:95.

10

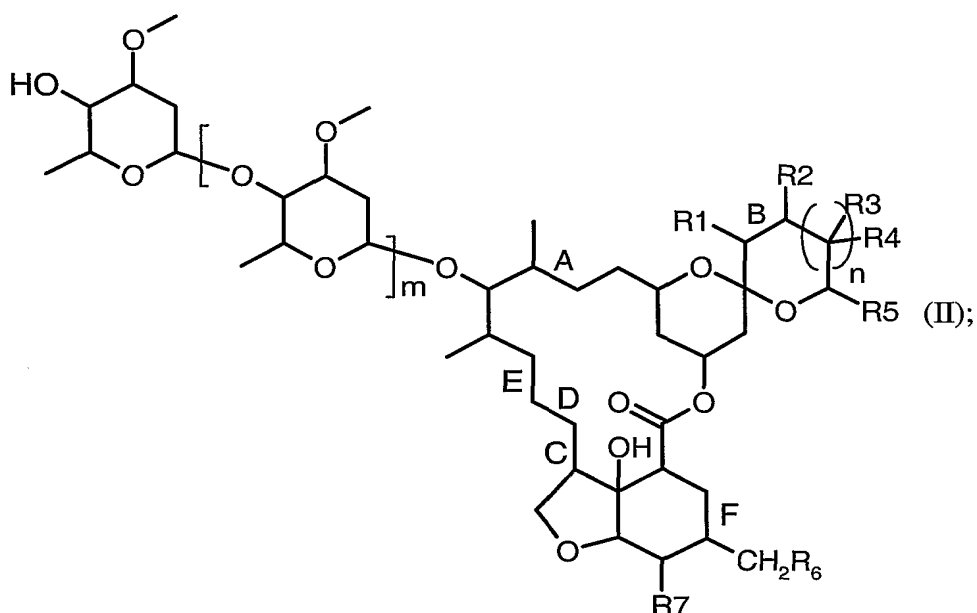
18. The polypeptide of anyone of claims 12 to 17, further comprising a tag.

19. A binding agent that specifically binds to the polypeptide of anyone of claims 12 to 18.

- 15      20. The binding agent of claim 20, wherein the binding agent is an antibody.

21. A family of polypeptides exhibiting an enzymatic activity of a P450 monooxygenase, wherein each member of the family is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formular (II)

20



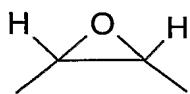
wherein

$R_1$ - $R_7$  represent, independently of each other hydrogen or a substituent;

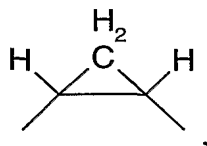
$m$  is 0, 1 or 2;

5  $n$  is 0, 1, 2 or 3; and

the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula

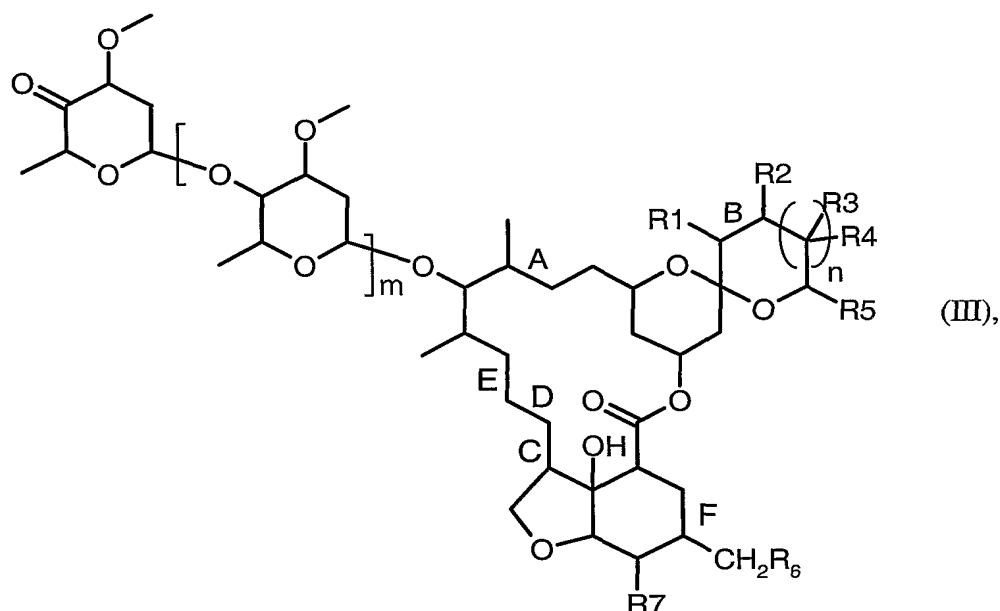


, or a single bond and a methylene bridge of the formula



10

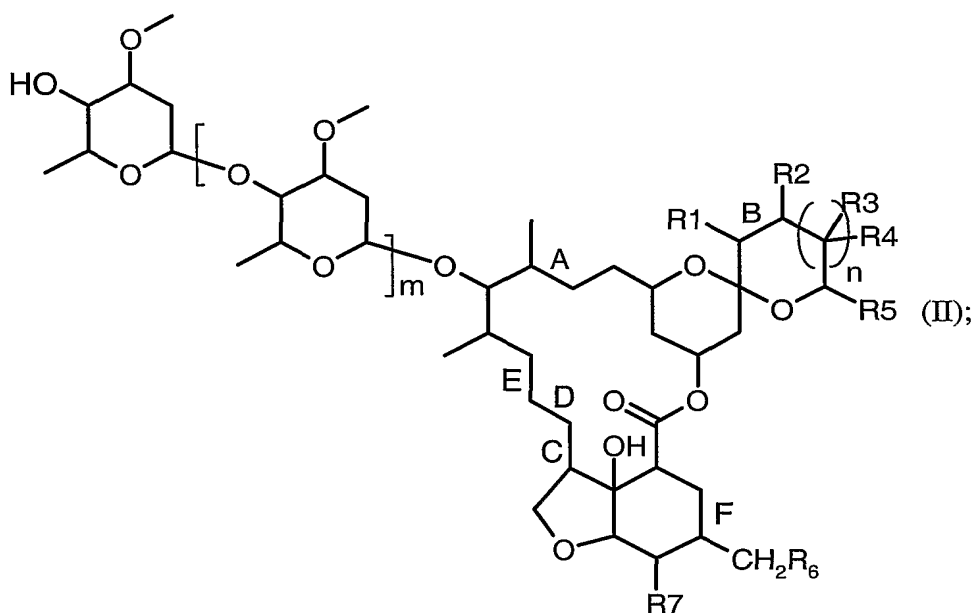
including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,  
in order to produce a compound of the formula (III)



wherein

$R_1$ - $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the same meanings as given for formula (II) above.

22. A family of polypeptides exhibiting an enzymatic activity of a P450 monooxygenase, wherein each member of the family oxidizes avermectin to 4''keto-avermectin.
23. The family of claims 21 or 22, wherein each member of the family is comprises an amino acid sequence that is at least 50% identical to SEQ ID NO:2.
24. A purified nucleic acid molecule comprising a nucleotide sequence encoding a polypeptide exhibiting an enzymatic activity of a ferredoxin and a ferredoxin reductase, respectively, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that is capable of regioselectively oxidizing the alcohol at position 4'' of a compound of formular (II)



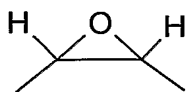
wherein

$R_1$ - $R_7$  represent, independently of each other hydrogen or a substituent;

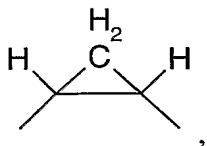
$m$  is 0, 1 or 2;

5  $n$  is 0, 1, 2 or 3; and

the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



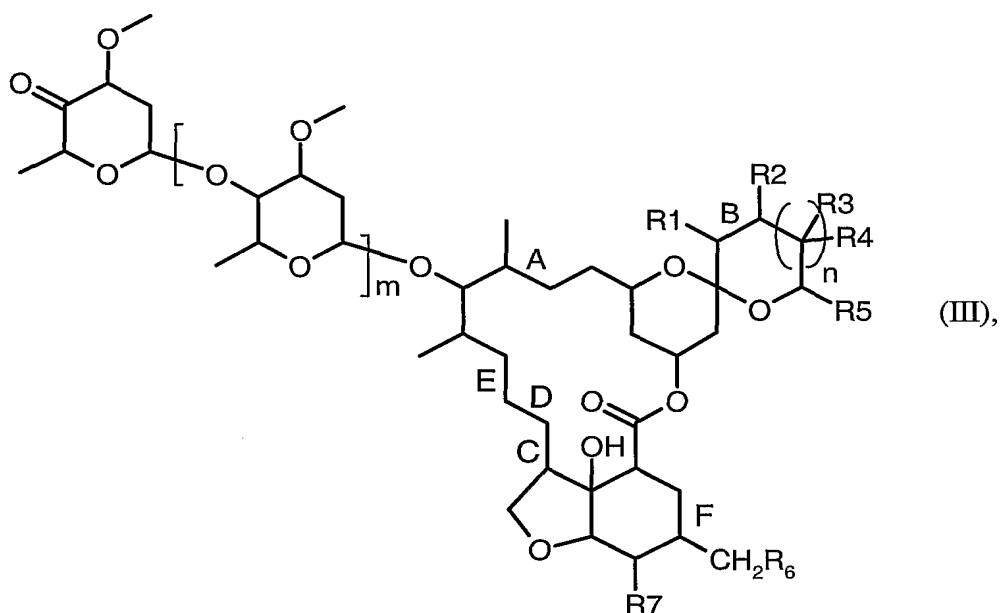
, or a single bond and a methylene bridge of the formula



10

including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form, in order to produce a compound of the formula (III)





wherein

$R_1$ - $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the same meanings as given for formula (II) above.

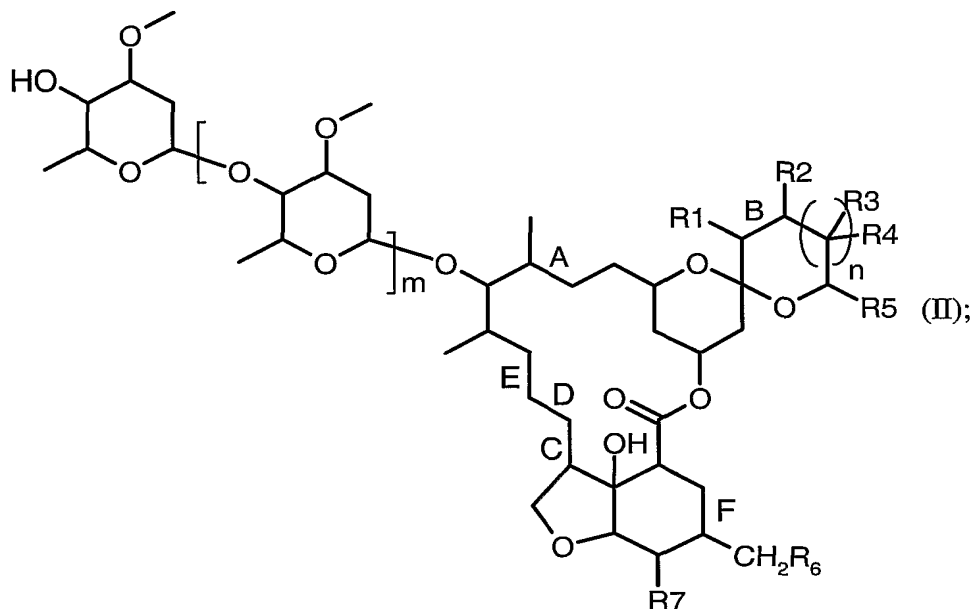
25. A purified nucleic acid molecule according to claim 24 comprising a nucleotide sequence encoding a polypeptide exhibiting an enzymatic activity of a ferredoxin and a ferredoxin reductase, respectively, wherein the nucleic acid molecule is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4''keto-avermectin.

26. The nucleic acid molecule of claim 25, comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:35 and SEQ ID NO:37.

27. The nucleic acid molecule of claim 25, comprising a nucleic acid sequence selected from the group consisting of SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:102, and SEQ ID NO:104.

28. A polypeptide exhibiting an enzymatic activity of a ferredoxin and a ferredoxin reductase, respectively, wherein the polypeptide is isolated from a *Streptomyces* strain comprising a

P450 monooxygenase that is capable of regioselectively oxidizing the alcohol at position 4" of a compound of formula (II)



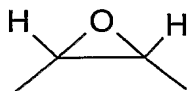
5 wherein

$R_1$ - $R_7$  represent, independently of each other hydrogen or a substituent;

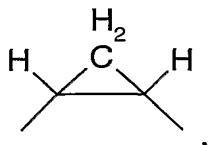
$m$  is 0, 1 or 2;

$n$  is 0, 1, 2 or 3; and

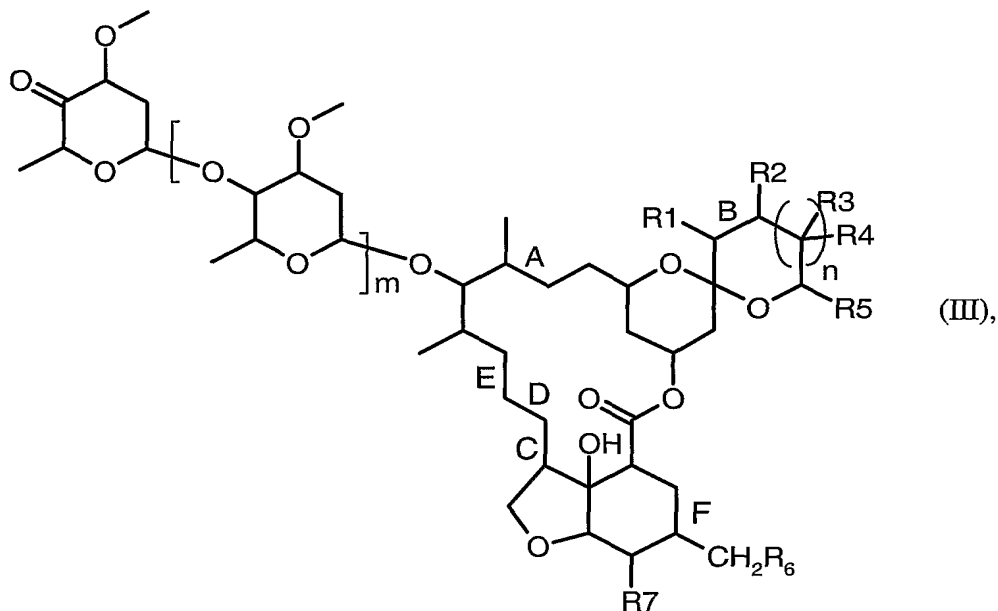
10 the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula



, or a single bond and a methylene bridge of the formula



including, where applicable, an E/Z isomer, a mixture of E/Z isomers, and/or a tautomer thereof, in each case in free form or in salt form,  
in order to produce a compound of the formula (III)



wherein

$R_1$ - $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the same meanings as given for formula (II) above.

29. A polypeptide exhibiting an enzymatic activity of a ferredoxin and a ferredoxin reductase, respectively, wherein the ferredoxin protein is isolated from a *Streptomyces* strain comprising a P450 monooxygenase that regioselectively oxidizes avermectin to 4''keto-  
avermectin.
30. The ferredoxin protein of claim 29, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:36 and SEQ ID NO:38.
31. The ferredoxin reductase protein of claim 29, comprising an amino acid sequence selected from the group consisting of SEQ ID NO:99, SEQ ID NO:101, SEQ ID NO:103, and SEQ ID NO:105.

32. A cell genetically engineered to comprise a nucleic acid molecule encoding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase according to anyone of claims 1 to 11.

5 33. The cell of claim 32 further comprising a nucleic acid molecule encoding a ferredoxin protein and a ferredoxin reductase protein, respectively, or a combination thereof.

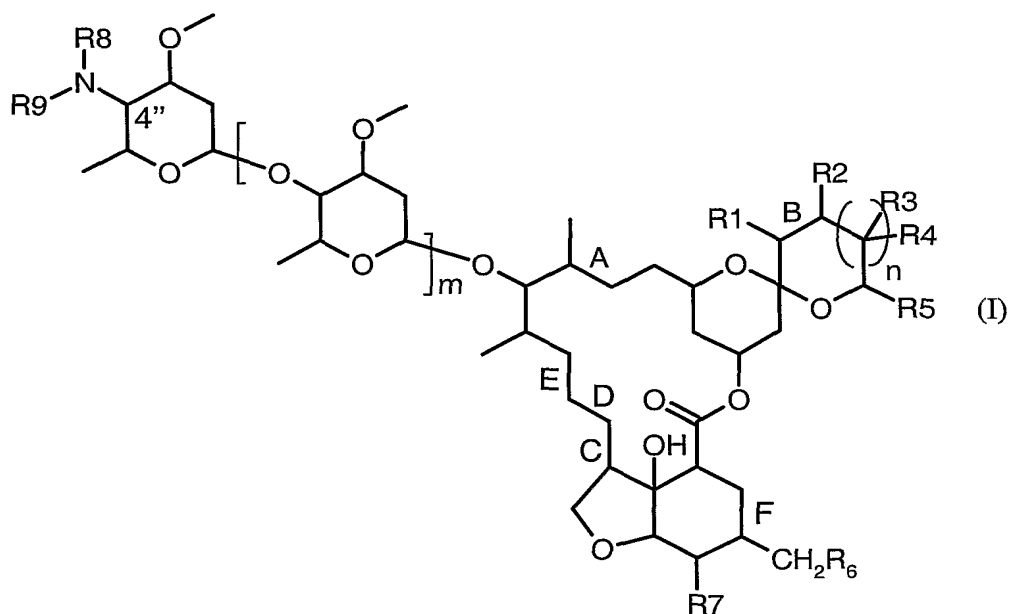
34. The cell of claims 32 or 33, wherein the nucleic acid molecule is positioned for expression in the cell.

10

35. The cell of anyone of claims 32 to 34, wherein the cell is a genetically engineered cell selected from the group consisting of a *Streptomyces* strain cell and a *Pseudomona* strain cell, and an *Escherichia coli* strain cell.

15 36. The cell of claim 35, wherein the cell has NRRL Designation No. B-30478 and NRRL Designation No.B-30479, respectively.

37. A method for the preparation a compound of the formula



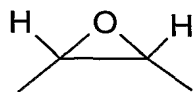
in which

$R_1$ - $R_9$  represent, independently of each other hydrogen or a substituent;

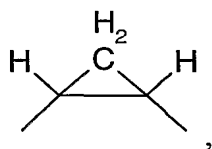
$m$  is 0, 1 or 2;

$n$  is 0, 1, 2 or 3; and

- 5 the bonds marked with A, B, C, D, E and F indicate, independently of each other, that two adjacent carbon atoms are connected by a double bond, a single bond, a single bond and a epoxide bridge of the formula

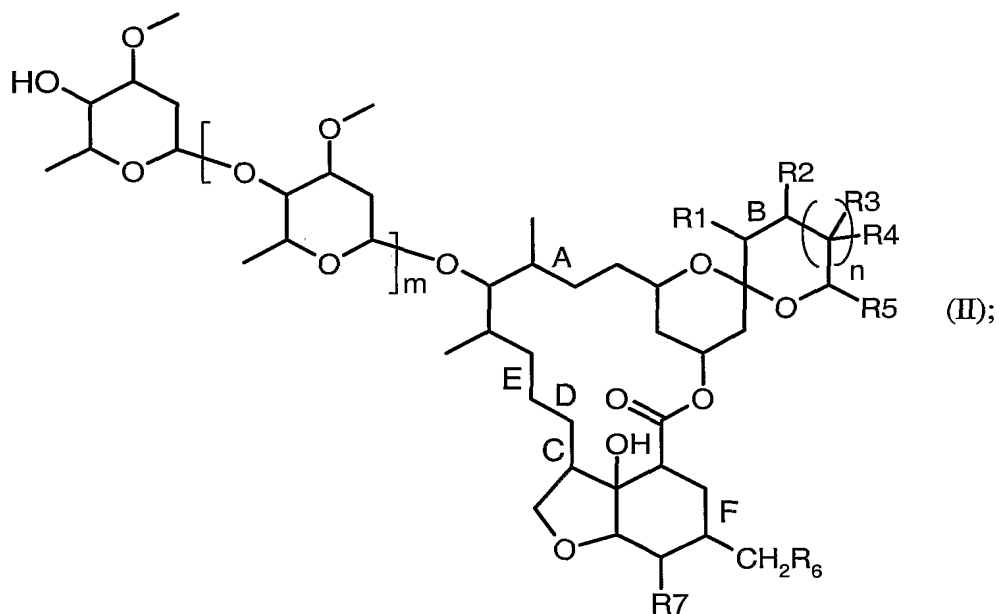


, or a single bond and a methylene bridge of the formula



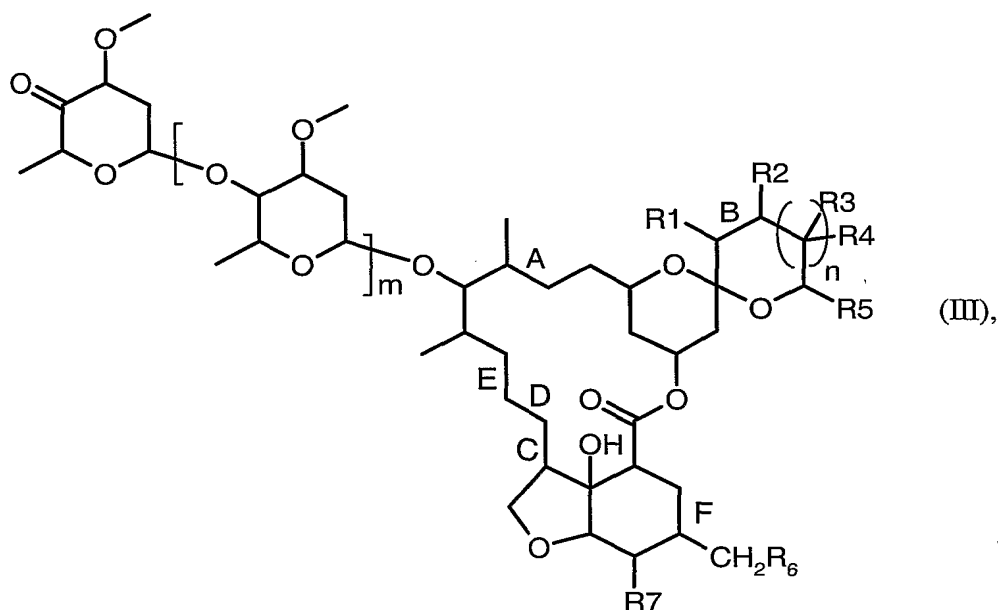
- 10 including, where applicable, an *E/Z* isomer, a mixture of *E/Z* isomers, and/or a tautomer thereof, in each case in free form or in salt form,  
which process comprises

1) bringing a compound of the formula



wherein

$R_1$ - $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the same meanings as given for formula (I) above, into contact with a polypeptide according to the invention that is capable of regioselectively oxidising the alcohol at position 4" in order to form a compound of the formula



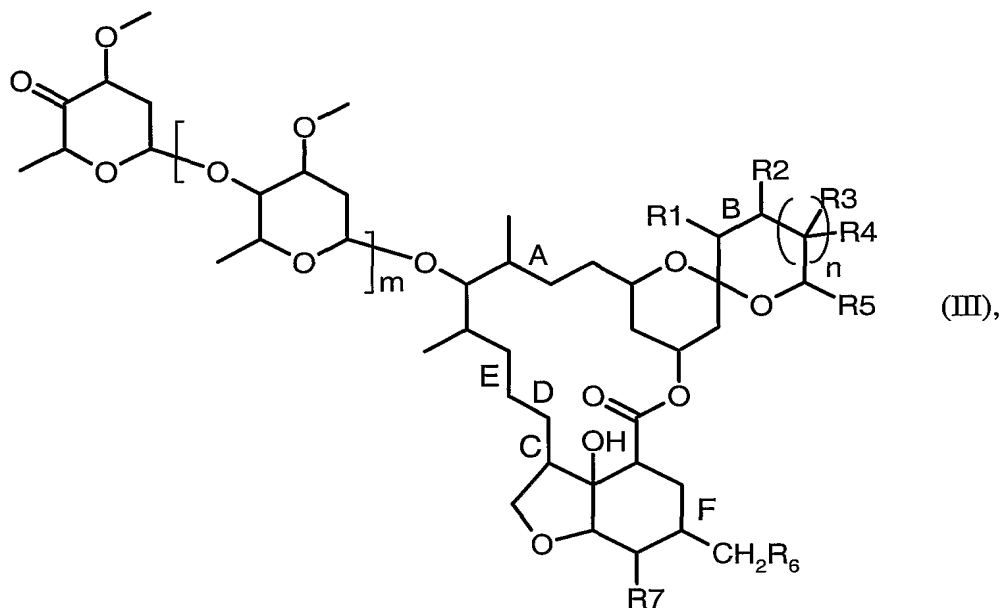
in which  $R_1$ ,  $R_2$ ,  $R_3$ ,  $R_4$ ,  $R_5$ ,  $R_6$ ,  $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the meanings given for formula (I); and

2) reacting the compound of the formula (III) with an amine of the formula  $HN(R_8)R_9$ , wherein  $R_8$  and  $R_9$  have the same meanings as given for formula (I), and which is known, in the presence of a reducing agent;

and, in each case, if desired, converting a compound of formula (I) obtainable in accordance with the process or by another method, or an  $E/Z$  isomer or tautomer thereof, in each case in free form or in salt form, into a different compound of formula (I) or an  $E/Z$  isomer or tautomer thereof, in each case in free form or in salt form, separating a mixture of  $E/Z$  isomers obtainable in accordance with the process and isolating the desired isomer, and/or converting a free compound of formula (I) obtainable in accordance with the process or by another method, or an  $E/Z$  isomer or tautomer thereof, into a salt or converting a salt, obtainable in accordance with the process or by another

method, of a compound of formula (I) or of an E/Z isomer or tautomer thereof into the free compound of formula (I) or an E/Z isomer or tautomer thereof or into a different salt.

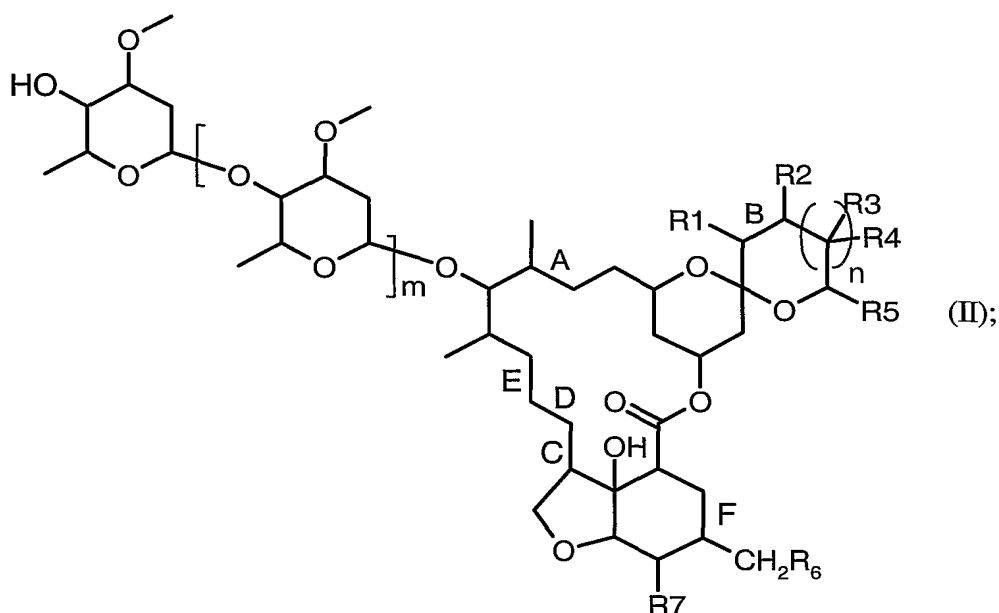
- 5 38. A method for the preparation of a compound of the formula



in which  $R_1$ ,  $R_2$ ,  $R_3$ ,  $R_4$ ,  $R_5$ ,  $R_6$ ,  $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the meanings given for formula (III) of claim 37,

10 which process comprises

- 1) bringing a compound of the formula



wherein

$R_1$ - $R_7$ ,  $m$ ,  $n$ ,  $A$ ,  $B$ ,  $C$ ,  $D$ ,  $E$  and  $F$  have the same meanings as given for formula (I) above, into contact with a polypeptide according to the invention that is capable of regiospecifically oxidising the alcohol at position 4", maintaining said contact for a time sufficient for the oxidation reaction to occur and isolating and purifying the compound of formula (II).

39. A method according to anyone of claims 37 or 38 for making emamectin, comprising adding a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regiospecifically oxidizes avermectin to 4''keto-avermectin to a reaction mixture comprising avermectin and incubating the reaction mixture under conditions that allow the polypeptide to regiospecifically oxidize avermectin to 4''keto-avermectin.

40. The method of anyone of claims 37 to 39, wherein the reaction mixture further comprises a ferredoxin protein.

41. The method of anyone of claims 37 to 40, wherein the reaction mixture further comprises a ferredoxin reductase protein.

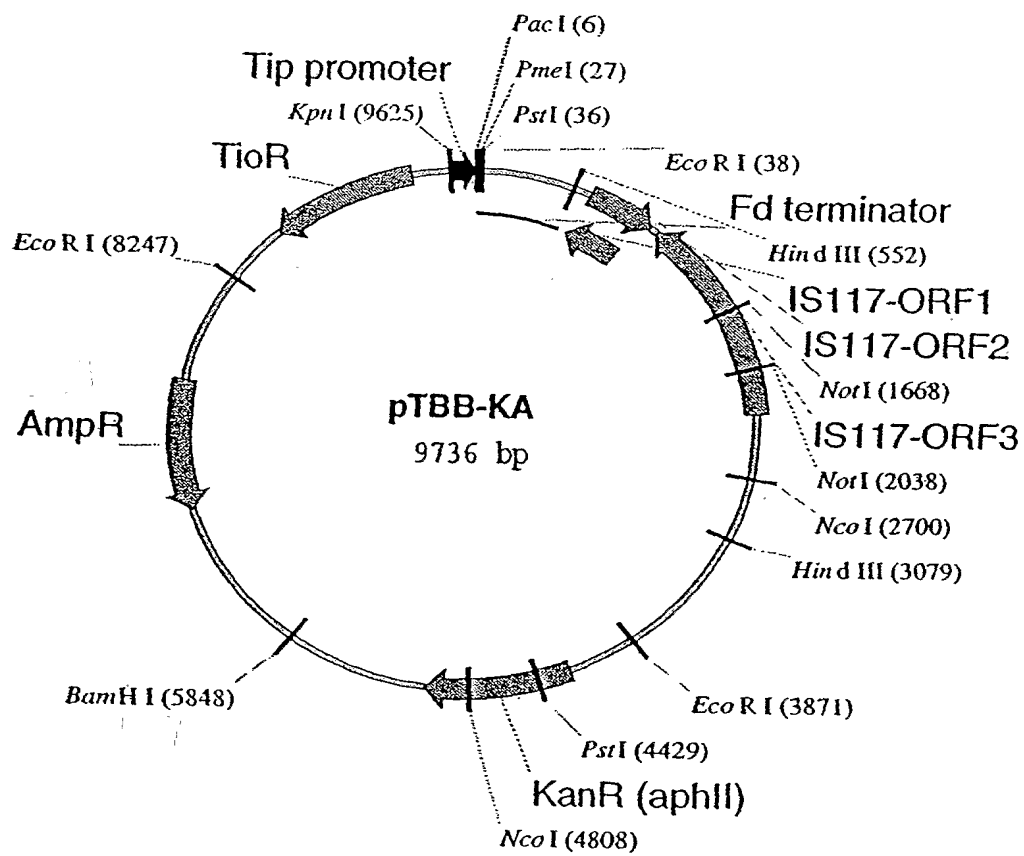


42. A formulation for making emamectin comprising a polypeptide that exhibits an enzymatic activity of a P450 monooxygenase and regioselectively oxidizes avermectin to 4''keto-avermectin.

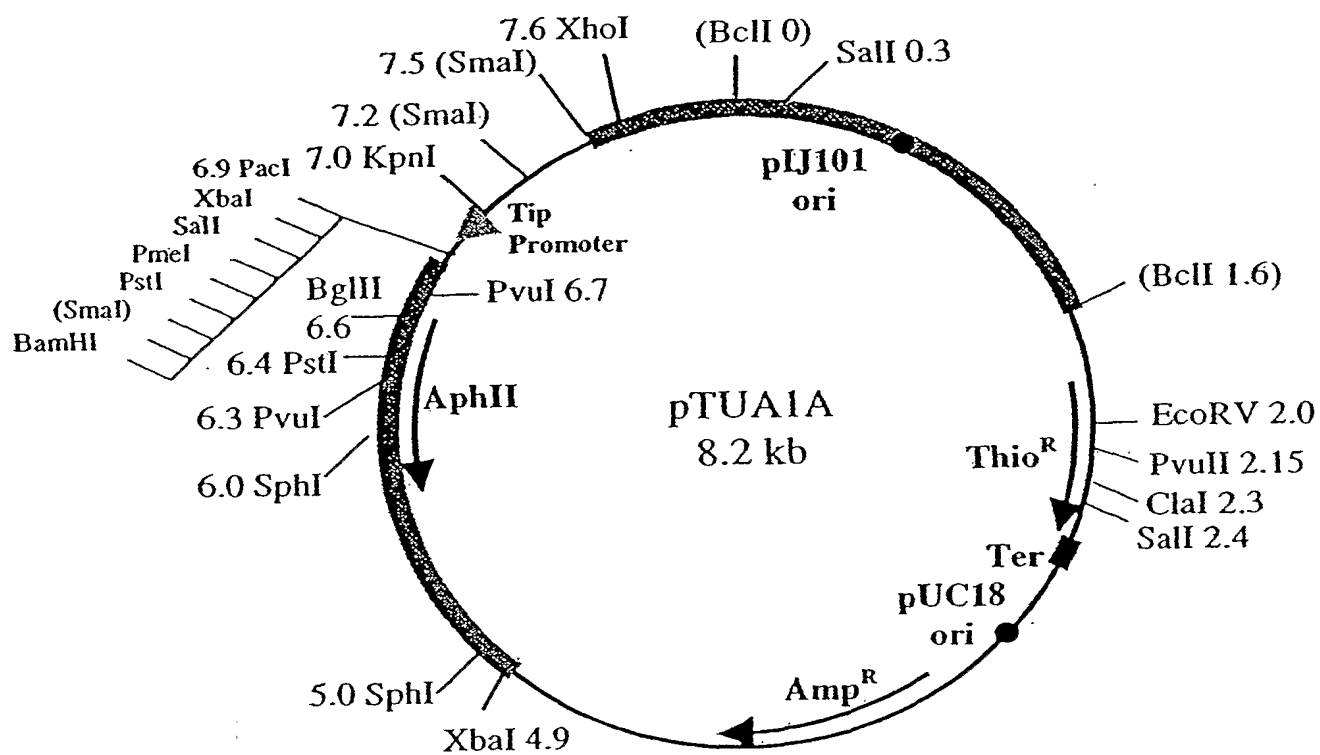
5

43. The formulation of claims 42 further comprising a ferredoxin protein.

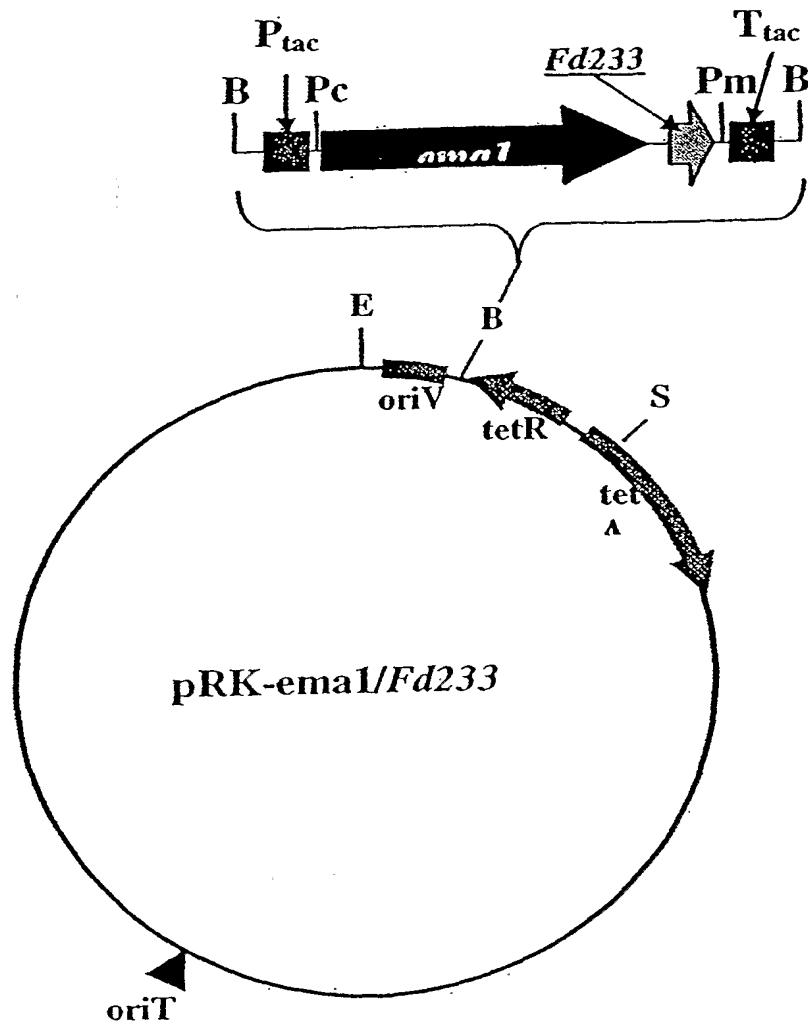
44. The formulation of claim 42 or 43 further comprising a ferredoxin reductase protein.



**Figure 1**



**Figure 2**



**Figure 3**

## SEQUENCE LISTING

<110> Syngenta Participations AG

<120> METHODS AND COMPOSITIONS FOR MAKING EMAMECTIN

<130> PB/5-60016A

<140>

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85      90      95
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Leu Asn Tyr Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser
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340     345     350
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His
355     360     365
Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala
370     375     380
Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Leu Thr His Tyr Pro
385     390     395     400
Asp Ile Ser Leu Gly Ile Ala Pro Glu His Leu Glu Arg Thr Pro Leu
405     410     415
Pro Gly Asn Trp Arg Leu Asn Ser Leu Pro Val Arg Leu Gly

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420

425

430

<210> 3  
 <211> 1293  
 <212> DNA  
 <213> *Streptomyces tubercidicus*

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 cccctggccc catcggccga ggagaacccg ctgaccaggc tgatggacat gctgggcctc 300  
 cccgagcacc tccgcgtcta catgctcggg tcgattctca actacgacgc ccccgaccac 360  
 acccggtgc gccgtctggt gtcgcgcgcg ttcacggcgc ggaagatcac cgatctgcga 420  
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 tcgatggacc cggaccgggt cggcgcaacg ttcccggcga tgatcgagca catccatgag 660  
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 catgacgacg atggcgccg gctcagcgac gtcgagatgg tcacatgat cctcacgctc 780  
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<210> 4  
 <211> 430  
 <212> PRT  
 <213> *Streptomyces tubercidicus*

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 35 40 45  
 Phe Val Asp Asp Ser Pro Val Trp Phe Val Thr Arg Phe Glu Glu Val  
 50 55 60  
 Arg Gln Val Leu Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Ala Pro  
 65 70 75 80  
 Pro Leu Ala Pro Ser Ala Glu Glu Asn Pro Leu Thr Arg Leu Met Asp  
 85 90 95  
 Met Leu Gly Leu Pro Glu His Leu Arg Val Tyr Met Leu Gly Ser Ile  
 100 105 110  
 Leu Asn Tyr Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser

115	120	125
Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Glu		
130	135	140
Gln Ile Ala Asp Glu Leu Leu Ala Arg Leu Pro Glu Tyr Ala Glu Asp		
145	150	155
Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr		
165	170	175
Val Ile Cys Glu Leu Val Gly Ile Pro Glu Ala Asp Arg Pro Gln Trp		
180	185	190
Arg Lys Trp Gly Ala Asp Leu Ile Ser Met Asp Pro Asp Arg Leu Gly		
195	200	205
Ala Thr Phe Pro Ala Met Ile Glu His Ile His Glu Met Val Arg Glu		
210	215	220
Arg Arg Ala Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr		
225	230	235
His Asp Asp Asp Gly Gly Arg Leu Ser Asp Val Glu Met Val Thr Met		
245	250	255
Ile Leu Thr Leu Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile		
260	265	270
Ser Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu		
275	280	285
Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met		
290	295	300
Arg Trp Cys Gly Pro Val Gln Met Thr Gln Leu Arg Tyr Ala Ala Ala		
305	310	315
Asp Val Asp Leu Ala Gly Thr Arg Ile His Lys Gly Asp Ala Val Gln		
325	330	335
Leu Leu Leu Val Ala Ala Asn Phe Asp Pro Arg His Tyr Thr Asp Pro		
340	345	350
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His		
355	360	365
Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala		
370	375	380
Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Leu Ala His Tyr Pro		
385	390	395
Glu Met Ser Leu Gly Ile Glu Pro Glu Arg Leu Glu Arg Leu Pro Leu		
405	410	415
Pro Gly Asn Trp Arg Leu Asn Ser Leu Pro Leu Arg Leu Gly		
420	425	430

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 <211> 1413  
 <212> DNA  
 <213> Streptomyces rimosus

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 cgcgcccggt tcatcgacga ctgcgccatc tggctggtga cccgcttcga cgtggtgcgc 300  
 gaggtgatgc gtgaccagcg gttcgtcaac aaccgaccc tgggtgcccg catcggcgcg 360  
 gacaaggacc cgcgtgcccc gctgatcgag ctgttcggca tccccgagga cctggccccg 420



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gaggccgagg tggcctacgg gaagctgctc acccgctacc cggacctggc gctcgccctc 1320
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<210> 6  
 <211> 470  
 <212> PRT  
 <213> Streptomyces rimosus

<400> 6

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			20					25					30		
Ala	Ala	Ser	Pro	Asp	Thr	Thr	Asp	Arg	Thr	Thr	Leu	Pro	Ser	Tyr	Val
		35					40					45			
Gly	Leu	His	Pro	Gly	Glu	Pro	Asn	Leu	Met	Glu	Pro	Glu	Leu	Leu	Glu
	50					55					60				
Asn	Pro	Tyr	Thr	Gly	Tyr	Gly	Thr	Leu	Arg	Glu	Gln	Ala	Pro	Leu	Val
65					70					75				80	
Arg	Ala	Arg	Phe	Ile	Asp	Asp	Ser	Pro	Ile	Trp	Leu	Val	Thr	Arg	Phe
				85					90					95	
Asp	Val	Val	Arg	Glu	Val	Met	Arg	Asp	Gln	Arg	Phe	Val	Asn	Asn	Pro
			100					105					110		
Thr	Leu	Val	Pro	Gly	Ile	Gly	Ala	Asp	Lys	Asp	Pro	Arg	Ala	Arg	Leu
		115					120					125			
Ile	Glu	Leu	Phe	Gly	Ile	Pro	Glu	Asp	Leu	Ala	Pro	Tyr	Leu	Thr	Asp
	130					135					140				
Asn	Ile	Leu	Thr	Ser	Asp	Pro	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu
145					150					155					160
Val	Ser	Arg	Ala	Phe	Thr	Ala	Arg	Arg	Ile	Gln	Asp	Leu	Arg	Pro	Arg
				165					170					175	
Val	Glu	Arg	Ile	Thr	Asp	Glu	Leu	Leu	Glu	Arg	Leu	Pro	Asp	His	Ala
			180					185					190		
Glu	Asp	Gly	Val	Val	Asp	Leu	Val	Glu	His	Phe	Ala	Tyr	Pro	Leu	Pro
		195					200					205			
Ile	Thr	Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Asp	Glu	Glu	Asp	Arg	Ala
	210					215					220				
Leu	Trp	Arg	Arg	Phe	Gly	Ala	Asp	Leu	Ala	Ser	Leu	Asn	Pro	Lys	Arg

225		230		235		240
Ile Gly Ala Thr Met	Pro Glu Met Ile Ser His Ile His Glu Leu Ile					
	245		250		255	
Asp Glu Arg Arg Ala Ala Leu Arg Asp Asp Leu Leu Ser Gly Leu Ile						
	260		265		270	
Arg Ala Gln Asp Asp Asp Gly Gly Arg Leu Ser Asp Val Glu Met Val						
	275		280		285	
Thr Leu Val Leu Thr Leu Val Leu Ala Gly His Glu Thr Thr Ala His						
	290		295		300	
Leu Ile Ser Asn Gly Thr Leu Ala Leu Leu Thr His Pro Asp Gln Arg						
305		310		315		320
Arg Leu Ile Asp Glu Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu						
	325		330		335	
Leu Met Arg Trp Cys Gly Pro Ile Gln Ala Thr Gln Leu Arg Tyr Ala						
	340		345		350	
Leu Glu Asp Thr Glu Val Ala Gly Val Gln Val Arg Gln Gly Glu Ala						
	355		360		365	
Leu Met Phe Ser Leu Val Ala Ala Asn His Asp Pro Arg His Tyr Thr						
370		375		380		
Gly Pro Glu Arg Leu Asp Leu Thr Arg Gln Pro Ala Gly Arg Ala Glu						
385		390		395		400
Asp His Val Gly Phe Gly His Gly Met His Tyr Cys Leu Gly Ala Ser						
	405		410		415	
Leu Ala Arg Gln Glu Ala Glu Val Ala Tyr Gly Lys Leu Leu Thr Arg						
	420		425		430	
Tyr Pro Asp Leu Ala Leu Ala Leu Thr Pro Glu Gln Leu Glu Asp Gln						
	435		440		445	
Glu Arg Leu Arg Gln Pro Gly Thr Trp Arg Leu Arg Arg Leu Pro Leu						
450		455		460		
Arg Leu His Ala Gln Ser						
465		470				

<210> 7  
 <211> 1293  
 <212> DNA  
 <213> Streptomyces lydicus

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 catgacgacg acggcagccg gctcagcgac gtcgagatgg tcaccatggt cctcaccgtc 780  
 gtccctggccg gccacgagac caccgcgcac ctcatcgcca acggcacggc ggccctgctc 840  
 acccaccaccg accagctgcg gctgctcaag gacgaccgg cgctgctgcc gcgcgcgggtg 900

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cacgagttga tgcgctggtg cggcccggtg cacatgaccc agctgcgcta cgccgccgag      960
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tcggcgaaac gcgacccgcg cactacacc gaccccgacc ggctggacct gaccgggcac      1080
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gagctgtcgc tggccgtcgc gccggaggcc ctggagcgca caccggtacc gggcagctgg      1260
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<210> 8  
 <211> 430  
 <212> PRT  
 <213> Streptomyces lydicus

<400> 8

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Pro	Gly	Glu	Pro	Asn	Val	Met	Asp	Pro	Ala	Leu	Ile	Gly	Asp	Pro	Phe
		20					25					30			
Ala	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Gly	Pro	Val	Val	Arg	Gly	Arg
		35				40					45				
Phe	Met	Asp	Asp	Ser	Pro	Val	Trp	Phe	Val	Thr	Arg	Phe	Glu	Glu	Val
	50					55				60					
Arg	Glu	Val	Leu	Arg	Asp	Pro	Arg	Phe	Arg	Asn	Asn	Pro	Val	Ser	Ala
65				70				75						80	
Ala	Pro	Gly	Ala	Ala	Pro	Glu	Asp	Thr	Pro	Leu	Ser	Arg	Leu	Met	Asp
			85					90						95	
Met	Met	Gly	Phe	Pro	Glu	His	Leu	Arg	Val	Tyr	Leu	Leu	Gly	Ser	Ile
		100					105						110		
Leu	Asn	Asn	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser
	115					120						125			
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Thr
	130					135					140				
Gln	Ile	Ala	Asp	Glu	Leu	Ala	Arg	Leu	Pro	Glu	His	Ala	Glu	Asp	
145				150				155						160	
Gly	Val	Val	Asp	Leu	Ile	Gln	His	Phe	Ala	Tyr	Pro	Leu	Pro	Ile	Thr
			165					170						175	
Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Pro	Glu	Glu	Asp	Arg	Pro	Gln	Trp
		180					185						190		
Arg	Thr	Trp	Gly	Ala	Asp	Leu	Val	Ser	Leu	Gln	Pro	Asp	Arg	Met	Ser
	195					200						205			
Arg	Ser	Phe	Pro	Ala	Met	Ile	Asp	His	Ile	His	Glu	Leu	Ile	Ala	Ala
	210				215						220				
Arg	Arg	Arg	Ala	Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Thr
225				230						235				240	
His	Asp	Asp	Asp	Gly	Ser	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Met
			245						250					255	
Val	Leu	Thr	Val	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile
			260				265						270		
Gly	Asn	Gly	Thr	Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	Arg	Leu
	275					280						285			
Leu	Lys	Asp	Asp	Pro	Ala	Leu	Leu	Pro	Arg	Ala	Val	His	Glu	Leu	Met
	290					295					300				
Arg	Trp	Cys	Gly	Pro	Val	His	Met	Thr	Gln	Leu	Arg	Tyr	Ala	Ala	Glu

305		310		315		320
Asp Val Glu Leu Ala Gly Val Arg Ile Arg Thr Gly Asp Ala Val Gln						
	325		330		335	
Leu Ile Leu Val Ser Ala Asn Arg Asp Pro Arg His Tyr Thr Asp Pro						
	340		345		350	
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His						
	355		360		365	
Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala						
	370		375		380	
Lys Gln Glu Gly Glu Val Ala Leu Gly Ala Leu Leu Arg His Phe Pro						
385		390		395		400
Glu Leu Ser Leu Ala Val Ala Pro Glu Ala Leu Glu Arg Thr Pro Val						
	405		410		415	
Pro Gly Ser Trp Arg Leu Asn Ala Leu Pro Leu Arg Leu Arg						
	420		425		430	

<210> 9  
 <211> 1299  
 <212> DNA  
 <213> Streptomyces sp.

<400> 9

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gccgcgctcg	ccaggcagga	aggcgaagtg	gcgttcggca	aactgctcgc	gcactacccg	1200
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<210> 10  
 <211> 432  
 <212> PRT  
 <213> Streptomyces sp.

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	20	25	30			
Gly Gly Tyr	Gly Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg					
	35	40	45			
Phe Met Asp Asp Ser Pro Val Trp Phe Val Thr Arg Phe Glu Glu Val						
	50	55	60			
Arg Gln Val Leu Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Ser Pro						
65	70	75	80			
Leu Leu Gly Ser Gln Val Glu Glu Met Pro Met Val Lys Leu Leu Glu						
	85	90	95			
Gln Met Gly Leu Pro Glu His Leu Arg Val Tyr Leu Leu Gly Ser Ile						
	100	105	110			
Leu Asn Ser Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser						
	115	120	125			
Arg Ala Phe Thr Ala Arg Lys Ile Thr Gly Leu Arg Pro Arg Val Glu						
	130	135	140			
Gln Ile Ala Asp Glu Leu Leu Ala Arg Leu Pro Glu His Ala Glu Asp						
145	150	155	160			
Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr						
	165	170	175			
Val Ile Cys Glu Leu Val Gly Ile Pro Glu Ala Asp Arg Pro Gln Trp						
	180	185	190			
Arg Ala Trp Gly Ala Asp Leu Val Ser Leu Glu Pro Asp Lys Leu Ser						
	195	200	205			
Thr Ser Phe Pro Ala Met Ile Asp His Thr His Glu Leu Ile Arg Gln						
	210	215	220			
Arg Arg Gly Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Ala						
225	230	235	240			
His Asp Asp Asp Gly Ser Arg Leu Ser Asp Val Glu Met Val Thr Met						
	245	250	255			
Val Phe Ala Leu Val Phe Ala Gly His Glu Thr Thr Ala His Leu Ile						
	260	265	270			
Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu						
	275	280	285			
Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met						
	290	295	300			
Arg Trp Cys Gly Pro Val His Met Thr Gln Leu Arg Tyr Ala Ser Glu						
305	310	315	320			
Asp Ile Asp Leu Ala Gly Thr Pro Ile Arg Lys Gly Asp Ala Val Gln						
	325	330	335			
Leu Ile Leu Val Ser Ala Asn Phe Asp Pro Arg His Tyr Ser Asp Pro						
	340	345	350			
Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His						
	355	360	365			
Val Gly Phe Gly His Gly Met His Tyr Cys Leu Gly Ala Ala Leu Ala						
	370	375	380			
Arg Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Leu Ala His Tyr Pro						
385	390	395	400			
Asp Val Ala Leu Gly Val Glu Pro Glu Ala Leu Glu Arg Val Pro Met						
	405	410	415			
Pro Gly Ser Trp Arg Leu Asn Ser Leu Pro Leu Arg Leu Ala Lys Arg						
	420	425	430			

<210> 11  
 <211> 1293  
 <212> DNA  
 <213> Streptomyces chattanoogenesis

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 ccgcgcgtca cacagatagc cgacgagctg ctggcccggc tgccggagca cgccgaggac 480  
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 cagagttga tgcgtgggtg cggcccgggt caccatgacc agctgcgcta cgccgcccag 960  
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 gccacgctcg ccaagcagga gggcgaggtc gccctcggcg ccctgtctcag gcacttcccc 1200  
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<210> 12  
 <211> 430  
 <212> PRT  
 <213> Streptomyces chattanoogenesis

<400> 12  
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 35 40 45  
 Phe Met Asp Asp Ser Pro Val Trp Phe Val Thr Arg Phe Glu Glu Val  
 50 55 60  
 Arg Glu Val Leu Arg Asp Pro Arg Phe Arg Asn Asn Pro Val Ser Ala  
 65 70 75 80  
 Ala Pro Gly Ala Ala Pro Glu Asp Thr Pro Leu Ser Arg Leu Met Asp  
 85 90 95  
 Met Met Gly Phe Pro Glu His Leu Arg Val Tyr Leu Leu Gly Ser Ile  
 100 105 110  
 Leu Asn Asn Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser  
 115 120 125  
 Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Thr

130		135		140
Gln Ile Ala Asp Glu Leu	Leu Ala Arg Leu Pro	Glu His Ala Glu Asp		
145		150		155
Gly Val Val Asp Leu Ile	Gln His Phe Ala Tyr	Pro Leu Pro Ile Thr		160
	165		170	
Val Ile Cys Glu Leu Val	Gly Ile Pro Glu Glu	Asp Arg Pro Gln Trp		175
	180		185	
Arg Thr Trp Gly Ala Asp	Leu Val Ser Leu Gln	Pro Asp Arg Met Ser		190
	195		200	
Arg Ser Phe Pro Ala Met	Ile Asp His Ile His	Glu Leu Ile Ala Ala		205
	210		215	
Arg Arg Arg Ala Leu Thr	Asp Asp Leu Leu Ser	Glu Leu Ile Arg Thr		220
	225		230	
His Asp Asp Asp Gly Ser	Arg Leu Ser Asp Val	Glu Met Val Thr Met		235
	245		250	
Val Leu Thr Val Val Leu	Ala Gly His Glu Thr	Thr Ala His Leu Ile		255
	260		265	
Gly Asn Gly Thr Ala Ala	Leu Leu Thr His Pro	Asp Gln Leu Arg Leu		270
	275		280	
Leu Lys Asp Asp Pro Ala	Leu Leu Pro Arg Ala	Val His Glu Leu Met		285
	290		295	
Arg Trp Cys Gly Pro Val	His Met Thr Gln Leu	Arg Tyr Ala Ala Glu		300
	305		310	
Asp Val Glu Leu Ala Gly	Val Arg Ile Arg Thr	Gly Asp Ala Val Gln		315
	325		330	
Leu Ile Leu Val Ser Ala	Asn Arg Asp Pro Arg	His Tyr Thr Asp Pro		335
	340		345	
Asp Arg Leu Asp Leu Thr	Arg His Pro Ala Gly	His Ala Glu Asn His		350
	355		360	
Val Gly Phe Gly His Gly	Ala His Tyr Cys Leu	Gly Ala Thr Leu Ala		365
	370		375	
Lys Gln Glu Gly Glu Val	Ala Leu Gly Ala Leu	Leu Arg His Phe Pro		380
	385		390	
Glu Leu Ser Leu Ala Val	Ala Pro Asp Ala Leu	Glu Arg Thr Pro Val		395
	405		410	
Pro Gly Ser Trp Arg Leu	Asn Ala Leu Pro Leu	Arg Leu Gly		415
	420		425	
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<210> 13  
 <211> 1290  
 <212> DNA  
 <213> Streptomyces sp.

<400> 13

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gcgggaggtg gaagcgggtga	cacaccctcc	aaccgggtga	tggaaatcat	gggcctgccc	300
gagcactacc ggggtgtacct	cgccaacacc	atcctcacca	tggacgcccc	cgaccacacc	360
cggatccggc gattggtctc	ccgggcattc	accgcccgtg	agatcaccca	tctgcgaccc	420
cgggtggagg acatcgcgga	cgatctgctg	aggcgggtgc	ccgagcacgc	cgaggacggc	480
gtcgtcgacc tcatcaagca	ctacgcctat	ccgctgcccc	taacgggtcat	ctgcgaactg	540

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atccgcgaac ggcgcgcggc gctcaccgac gatctgctca gcgaactgat ccgggtccat 720
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<210> 14  
 <211> 429  
 <212> PRT  
 <213> *Streptomyces* sp.

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 35 40 45  
 Phe Ala Asp Asp Thr Pro Val Trp Phe Ile Thr Arg Phe Glu Glu Ala  
 50 55 60  
 Arg Glu Val Leu Arg Asp His Arg Phe Ala Asn Ala Pro Ala Phe Ala  
 65 70 75 80  
 Ala Gly Gly Gly Ser Gly Asp Thr Pro Ser Asn Arg Leu Met Glu Ile  
 85 90 95  
 Met Gly Leu Pro Glu His Tyr Arg Val Tyr Leu Ala Asn Thr Ile Leu  
 100 105 110  
 Thr Met Asp Ala Pro Asp His Thr Arg Ile Arg Arg Leu Val Ser Arg  
 115 120 125  
 Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Glu Asp  
 130 135 140  
 Ile Ala Asp Asp Leu Leu Arg Arg Leu Pro Glu His Ala Glu Asp Gly  
 145 150 155 160  
 Val Val Asp Leu Ile Lys His Tyr Ala Tyr Pro Leu Pro Ile Thr Val  
 165 170 175  
 Ile Cys Glu Leu Val Gly Ile Pro Glu Glu Asp Arg Leu Gln Trp Arg  
 180 185 190  
 Asp Trp Gly Ser Ala Phe Val Ser Leu Gln Pro Asp Arg Leu Ser Lys  
 195 200 205  
 Ala Phe Pro Ala Met Ile Glu His Ile His Ala Leu Ile Arg Glu Arg  
 210 215 220  
 Arg Ala Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Val His  
 225 230 235 240  
 Asp Asp Asp Gly Gly Arg Leu Ser Asp Val Glu Met Val Thr Met Val  
 245 250 255  
 Leu Thr Leu Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile Gly



	260		265		270
Asn Gly Thr	Ala Ala Leu Leu Thr	His Pro Asp Gln	Leu His Leu Leu		
	275	280	285		
Lys Ser Asp	Pro Glu Leu Leu Pro	Arg Ala Val His Glu	Leu Met Arg		
	290	295	300		
Trp Cys Gly	Pro Val Gln Met Thr	Gln Leu Arg Tyr Ala	Thr Glu Asp		
305	310	315	320		
Val Glu Val	Ala Gly Val Gln Val	Lys Gln Gly Glu Ala	Val Leu Ala		
	325	330	335		
Met Leu Val	Ala Ala Asn His Asp	Pro Arg His Phe Ala	Asp Pro Ala		
	340	345	350		
Arg Leu Asp	Leu Thr Arg Gln Pro	Ala Gly Arg Ala Glu	Asn His Val		
	355	360	365		
Gly Phe Gly	His Gly Met His Tyr	Cys Leu Gly Ala Ser	Leu Ala Arg		
370	375	380			
Gln Glu Gly	Glu Val Ala Phe Gly	Asn Leu Leu Ala His	Tyr Pro Asp		
385	390	395	400		
Val Ser Leu	Ala Val Glu Pro Asp	Ala Leu Gln Arg Val	Pro Leu Pro		
	405	410	415		
Gly Asn Trp	Arg Leu Ala Ala Leu	Pro Val Arg Leu Arg			
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<210> 15  
 <211> 1428  
 <212> DNA  
 <213> Streptomyces albobaciens

<400> 15

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caggcgccgc	tcgtccggcg	ccggttcac	gacgactcgc	ccatctggct	ggtgaccgcg	300
ttcgacgtgg	tgcgcgaggt	gatgcgcgac	cagcggttcg	tcaacaacc	gaccctggtg	360
cccggcatcg	gtgcggacca	ggacccgcgc	gccgggctga	tcgagctggt	cgccatcccc	420
gaggacctgg	ccccgtacct	caccgacacc	atcctcacca	gcgaccgcgc	ggaccacacc	480
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 <211> 475  
 <212> PRT  
 <213> Streptomyces albobacillus

<400> 16  
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 35 40 45  
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 50 55 60  
 Pro Glu Leu Leu Asp Asn Pro Tyr Thr Gly Tyr Gly Thr Leu Arg Glu  
 65 70 75 80  
 Gln Ala Pro Leu Val Arg Ala Arg Phe Ile Asp Asp Ser Pro Ile Trp  
 85 90 95  
 Leu Val Thr Arg Phe Asp Val Val Arg Glu Val Met Arg Asp Gln Arg  
 100 105 110  
 Phe Val Asn Asn Pro Thr Leu Val Pro Gly Ile Gly Ala Asp Gln Asp  
 115 120 125  
 Pro Arg Ala Arg Leu Ile Glu Leu Phe Gly Ile Pro Glu Asp Leu Ala  
 130 135 140  
 Pro Tyr Leu Thr Asp Thr Ile Leu Thr Ser Asp Pro Pro Asp His Thr  
 145 150 155 160  
 Arg Leu Arg Arg Leu Val Ser Arg Ala Phe Thr Ala Arg Arg Ile Gln  
 165 170 175  
 Asp Leu Arg Pro Arg Val Glu Arg Ile Thr Asp Glu Leu Leu Ala Arg  
 180 185 190  
 Leu Pro Asp His Ala Glu Asp Gly Val Val Asp Leu Val Glu His Phe  
 195 200 205  
 Ala Tyr Pro Leu Pro Ile Thr Val Ile Cys Glu Leu Val Gly Ile Asp  
 210 215 220  
 Glu Glu Asp Arg Ala Leu Trp Arg Arg Phe Gly Ala Asp Leu Ala Ser  
 225 230 235 240  
 Leu Asn Pro Lys Arg Ile Gly Ala Thr Met Pro Glu Met Ile Ala His  
 245 250 255  
 Ile His Glu Val Ile Asp Glu Arg Arg Ala Asp Leu Arg Asp Asp Leu  
 260 265 270  
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 275 280 285  
 Asp Val Glu Met Val Thr Leu Val Leu Thr Leu Val Leu Ala Gly His  
 290 295 300  
 Glu Thr Thr Ala His Leu Ile Ser Asn Gly Thr Leu Ala Leu Leu Thr  
 305 310 315 320  
 His Pro Asp Gln Arg Arg Leu Ile Asp Glu Asp Pro Ala Leu Leu Pro  
 325 330 335  
 Arg Ala Val His Glu Leu Met Arg Trp Cys Gly Pro Ile Gln Ala Thr  
 340 345 350  
 Gln Leu Arg Tyr Ala Met Glu Asp Thr Glu Val Ala Gly Val Gln Val  
 355 360 365  
 Arg Gln Gly Glu Ala Leu Met Phe Ser Leu Val Ala Ala Asn His Asp

370						375					380					
Pro	Arg	His	Tyr	Thr	Gly	Pro	Glu	Arg	Leu	Asp	Leu	Thr	Arg	Gln	Pro	
385						390				395					400	
Ala	Gly	Arg	Ala	Glu	Asp	His	Val	Gly	Phe	Gly	His	Gly	Met	His	Tyr	
				405					410					415		
Cys	Leu	Gly	Ala	Ser	Leu	Ala	Arg	Gln	Glu	Ala	Glu	Val	Ala	Tyr	Gly	
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Lys	Leu	Leu	Thr	Arg	Tyr	Pro	Asp	Leu	Ala	Leu	Ala	Leu	Thr	Pro	Glu	
			435				440						445			
Gln	Leu	Glu	Asp	Gln	Glu	Arg	Leu	Arg	Gln	Pro	Gly	Thr	Trp	Arg	Leu	
		450				455					460					
Arg	Arg	Leu	Pro	Leu	Arg	Leu	His	Ala	Glu	Ser						
465					470					475						

<210>	17
<211>	1293
<212>	DNA
<213>	Streptomyces platensis

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cagggcccgg	tcgtccgcgg	ccgcttcgtg	gacgactcac	ccgtctggct	ggtgacgcga				180
ttcaggagg	tccgccaaagt	cctgcgcgac	cagcggttcg	tgaacaaccc	ggcgggcgcc				240
tccttgggccc	acgcggccga	ggacaacccg	ctcaccaggc	tgatggacat	gctggggcctc				300
cccgagcacc	tccgcccccta	cctcctcgga	tcgattctca	attacgacgc	ccccgaccac				360
acccggctgc	gccgcctggt	gtcgcggggc	ttcaccgccc	gcaagatcac	cgacctgcgg				420
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ggcgtcgtcg	atctcatccg	gcacttcgcc	taccgcgtgc	cgatcacctg	catctgcgaa				540
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tcgatggagc	cggaccggct	caccgcctcg	ttcccgcoga	tgatcgagca	catccaccgg				660
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gagatggcgt	tgggcgtcgc	accggaagcg	ctggagcgga	cgcacctgcc	gggcaactgg				1260
cggctgaacg	cgctgccggt	gcgggttgggg	tga						1293

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<210> 18
<211> 430
<212> PRT
<213> Streptomyces platensis
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 35 40 45  
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 50 55 60  
 Arg Gln Val Leu Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Ala Pro  
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 Ser Leu Gly His Ala Ala Glu Asp Asn Pro Leu Thr Arg Leu Met Asp  
 85 90 95  
 Met Leu Gly Leu Pro Glu His Leu Arg Pro Tyr Leu Leu Gly Ser Ile  
 100 105 110  
 Leu Asn Tyr Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser  
 115 120 125  
 Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Glu  
 130 135 140  
 Gln Ile Ala Asp Ala Leu Leu Ala Arg Leu Pro Glu His Ala Glu Asp  
 145 150 155 160  
 Gly Val Val Asp Leu Ile Arg His Phe Ala Tyr Pro Leu Pro Ile Thr  
 165 170 175  
 Val Ile Cys Glu Leu Val Gly Ile Pro Glu Ala Asp Arg Pro Gln Trp  
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 Arg Thr Trp Gly Ala Asp Leu Val Ser Met Glu Pro Asp Arg Leu Thr  
 195 200 205  
 Ala Ser Phe Pro Pro Met Ile Glu His Ile His Arg Met Val Arg Glu  
 210 215 220  
 Arg Arg Gly Ala Leu Thr Gly Asp Leu Leu Ser Glu Leu Ile Arg Ala  
 225 230 235 240  
 His Asp Asp Asp Gly Gly Arg Leu Ser Asp Val Glu Met Val Thr Leu  
 245 250 255  
 Ile Leu Thr Leu Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile  
 260 265 270  
 Ser Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu  
 275 280 285  
 Leu Gln Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met  
 290 295 300  
 Arg Trp Cys Gly Pro Val Gln Met Thr Gln Leu Arg Tyr Ala Ala Ala  
 305 310 315 320  
 Asp Val Asp Leu Ala Gly Thr Thr Ile His Arg Gly Asp Ala Val Gln  
 325 330 335  
 Leu Ile Leu Val Ser Ala Asn Phe Asp Pro Arg His Tyr Thr Asp Pro  
 340 345 350  
 Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His  
 355 360 365  
 Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala  
 370 375 380  
 Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Leu Ala His Tyr Pro  
 385 390 395 400  
 Glu Met Ala Leu Gly Val Ala Pro Glu Arg Leu Glu Arg Thr Pro Leu  
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 Pro Gly Asn Trp Arg Leu Asn Ala Leu Pro Val Arg Leu Gly  
 420 425 430

<210> 19  
 <211> 1293  
 <212> DNA  
 <213> *Streptomyces kasugaensis*

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<210> 20  
 <211> 430  
 <212> PRT  
 <213> *Streptomyces kasugaensis*

<400> 20  
 Met Ser Ala Ser Pro Ser Asn Thr Phe Thr Glu His Val Gly Lys His  
 1 5 10 15  
 Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Gly Asp Pro Phe  
 20 25 30  
 Ala Gly Tyr Gly Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg  
 35 40 45  
 Phe Met Asp Asp Ser Pro Val Trp Phe Val Thr Arg Phe Glu Glu Val  
 50 55 60  
 Arg Glu Val Leu Arg Asp Pro Arg Phe Arg Asn Asn Pro Val Ser Ala  
 65 70 75 80  
 Ala Pro Gly Ala Ala Pro Glu Asp Thr Pro Leu Ser Arg Leu Met Asp  
 85 90 95  
 Met Met Gly Phe Pro Glu His Leu Arg Val Tyr Leu Leu Gly Ser Ile  
 100 105 110  
 Leu Asn Asn Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser  
 115 120 125  
 Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Thr  
 130 135 140

Gln Ile Ala Asp Glu Leu Leu Ala Arg Leu Pro Glu His Ala Glu Asp  
 145 150 155 160  
 Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr  
 165 170 175  
 Val Ile Cys Glu Leu Val Gly Ile Pro Glu Glu Asp Arg Pro Gln Trp  
 180 185 190  
 Arg Thr Trp Gly Ala Asp Leu Val Ser Leu Gln Pro Asp Arg Met Ser  
 195 200 205  
 Arg Ser Phe Pro Ala Met Ile Asp His Ile His Glu Leu Ile Ala Ala  
 210 215 220  
 Arg Arg Arg Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr  
 225 230 235 240  
 His Asp Asp Asp Gly Ser Arg Leu Ser Asp Val Glu Met Val Thr Met  
 245 250 255  
 Val Leu Thr Val Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile  
 260 265 270  
 Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu  
 275 280 285  
 Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met  
 290 295 300  
 Arg Trp Cys Gly Pro Val His Met Thr Gln Leu Arg Tyr Ala Ala Glu  
 305 310 315 320  
 Asp Val Glu Leu Ala Gly Val Arg Ile Arg Thr Gly Asp Ala Val Gln  
 325 330 335  
 Leu Ile Leu Val Ser Ala Asn Arg Asp Pro Arg His Tyr Thr Asp Pro  
 340 345 350  
 Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His  
 355 360 365  
 Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala  
 370 375 380  
 Lys Gln Glu Gly Glu Val Ala Leu Gly Ala Leu Leu Arg His Phe Pro  
 385 390 395 400  
 Glu Leu Ser Leu Ala Val Ala Pro Glu Ala Leu Glu Arg Thr Pro Val  
 405 410 415  
 Pro Gly Ser Trp Arg Leu Asn Ala Leu Pro Leu Arg Leu Arg  
 420 425 430

&lt;210&gt; 21

&lt;211&gt; 1428

&lt;212&gt; DNA

&lt;213&gt; Streptomyces rimosus

&lt;400&gt; 21

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atgaccacat cgcccaccga gtcccggggcg gccaccccga ccggctccac cgctccccc 60
tcgaccgctt ccgcccgggc caccaccct tcggccgcca cctcttcgga caccacctat 120
cccgccacca ccgaccgcac cacgctcccc tcctacgtcg gcctccaccc gggcgagccg 180
aacctgatgg aaccggagct gctggacaac ccgtacaccg gctacggcac gctgcgcgag 240
caggccccgc tcgtccgtgc ccggttcacg gacgactcgc ccatctggct ggtgaccgcg 300
ttcgacgtgg tgcgcgaggt gatgcgcgac cagcggttcg tcaacaacc gacctgggtg 360
cccggcatcg gtgcggacaa ggaccgcgcg gcccggtga tcgagctgtt cggcatcccc 420
gaggacctga ccccgtaacct cgccgacacc atcctcacca gcgacccgcc ggaccacacc 480
cggtgcgcgc gcctggtctc ccgtgccttc accgcgcgcc gcatccagga cctgcggccg 540
cgcgtcgagc agatcaccca cgcgctgctg gagcgactgc cggaccatgc cgaggacggc 600

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gtcgtcgacc tcgtcgagca cttcgctac ccgctgcccc tcacgggtcat ctgcgagctg 660
gtcggcatcg acgaggagga cgggacgctg tggcgggcggg tcggcgccga cctcgctca 720
ctgaacccca agcgcatcgg cgccaccatg ccggagatga tcgcgcacat ccacgaggtg 780
atcgacgagc ggcgcgcggc cctgcgggac gacctgetca gcggggtcat ccgggcgcag 840
gacgacgacg gcggccgggt gagcgacgtc gagatgggtca ccctgggtcct gaccctgggtg 900
ctggccgggtc acgagaccac cgcccacctc atcagcaacg gcaccctcgc cctgctcacc 960
caccgccgacc agcggcgggt gatcgacgag gaccgggcac tgctgccgcg cgcgggtccac 1020
gagctgatgc gctggtgcgg gccgatccag gccaccacgc tcgggtacgc catggaggac 1080
accgaggtcg ccggtgtcca ggtccgccag ggcgaggccc tgatgttcag cctcgtcgcg 1140
gccaaccacg acccgcgcca ctacaccggg ccggagcggc tcgacctgac gcggcagccg 1200
gccggccgcg ccgaggacca cgtcggcttc gggcacggga tgcactactg cctgggtgcc 1260
tactcgcgcc ggcaggaggc cgaggtggcc tacgggaagc tgctcaccgc ctaccgggac 1320
ctggagctcg ctctcacacc ggaacagctg gaggaccagg aacgcctgcg gcagcccggc 1380
acctggcgcc tgcggcgggt gccgctgaag ctgcacgcgc ggagctga 1428

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&lt;210&gt; 22

&lt;211&gt; 475

&lt;212&gt; PRT

<213> *Streptomyces rimosus*

&lt;400&gt; 22

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Met Thr Thr Ser Pro Thr Glu Ser Arg Ala Ala Thr Pro Thr Gly Ser
1          5          10          15
Thr Ala Ser Pro Ser Thr Ala Ser Ala Pro Ala Thr Thr Pro Ser Ala
          20          25          30
Ala Thr Ser Ser Asp Thr Thr Tyr Pro Ala Thr Thr Asp Arg Thr Thr
          35          40          45
Leu Pro Ser Tyr Val Gly Leu His Pro Gly Glu Pro Asn Leu Met Glu
          50          55          60
Pro Glu Leu Leu Asp Asn Pro Tyr Thr Gly Tyr Gly Thr Leu Arg Glu
65          70          75          80
Gln Ala Pro Leu Val Arg Ala Arg Phe Ile Asp Asp Ser Pro Ile Trp
          85          90          95
Leu Val Thr Arg Phe Asp Val Val Arg Glu Val Met Arg Asp Gln Arg
          100          105          110
Phe Val Asn Asn Pro Thr Leu Val Pro Gly Ile Gly Ala Asp Lys Asp
          115          120          125
Pro Arg Ala Arg Leu Ile Glu Leu Phe Gly Ile Pro Glu Asp Leu Thr
          130          135          140
Pro Tyr Leu Ala Asp Thr Ile Leu Thr Ser Asp Pro Pro Asp His Thr
145          150          155          160
Arg Leu Arg Arg Leu Val Ser Arg Ala Phe Thr Ala Arg Arg Ile Gln
          165          170          175
Asp Leu Arg Pro Arg Val Glu Gln Ile Thr Asp Ala Leu Leu Glu Arg
          180          185          190
Leu Pro Asp His Ala Glu Asp Gly Val Val Asp Leu Val Glu His Phe
          195          200          205
Ala Tyr Pro Leu Pro Ile Thr Val Ile Cys Glu Leu Val Gly Ile Asp
210          215          220
Glu Glu Asp Arg Thr Leu Trp Arg Arg Phe Gly Ala Asp Leu Ala Ser
225          230          235          240
Leu Asn Pro Lys Arg Ile Gly Ala Thr Met Pro Glu Met Ile Ala His
          245          250          255

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Ile His Glu Val Ile Asp Glu Arg Arg Ala Ala Leu Arg Asp Asp Leu  
 260 265 270  
 Leu Ser Gly Leu Ile Arg Ala Gln Asp Asp Asp Gly Gly Arg Leu Ser  
 275 280 285  
 Asp Val Glu Met Val Thr Leu Val Leu Thr Leu Val Leu Ala Gly His  
 290 295 300  
 Glu Thr Thr Ala His Leu Ile Ser Asn Gly Thr Leu Ala Leu Leu Thr  
 305 310 315 320  
 His Pro Asp Gln Arg Arg Leu Ile Asp Glu Asp Pro Ala Leu Leu Pro  
 325 330 335  
 Arg Ala Val His Glu Leu Met Arg Trp Cys Gly Pro Ile Gln Ala Thr  
 340 345 350  
 Gln Leu Arg Tyr Ala Met Glu Asp Thr Glu Val Ala Gly Val Gln Val  
 355 360 365  
 Arg Gln Gly Glu Ala Leu Met Phe Ser Leu Val Ala Ala Asn His Asp  
 370 375 380  
 Pro Arg His Tyr Thr Gly Pro Glu Arg Leu Asp Leu Thr Arg Gln Pro  
 385 390 395 400  
 Ala Gly Arg Ala Glu Asp His Val Gly Phe Gly His Gly Met His Tyr  
 405 410 415  
 Cys Leu Gly Ala Ser Leu Ala Arg Gln Glu Ala Glu Val Ala Tyr Gly  
 420 425 430  
 Lys Leu Leu Thr Arg Tyr Pro Asp Leu Glu Leu Ala Leu Thr Pro Glu  
 435 440 445  
 Gln Leu Glu Asp Gln Glu Arg Leu Arg Gln Pro Gly Thr Trp Arg Leu  
 450 455 460  
 Arg Arg Leu Pro Leu Lys Leu His Ala Arg Ser  
 465 470 475

&lt;210&gt; 23

&lt;211&gt; 1293

&lt;212&gt; DNA

<213> *Streptomyces tubercidicus*

&lt;400&gt; 23

atgtcggcat tatccaactc cccgctcgcc gcacatgtcg ggaaacaccc tggcgagccg 60  
 aatgtgatgg acccggcgct gatcaccgac ccgttcggcg gctacggcgc actgcgcgag 120  
 caaggcccg tcttacgggg ccggttcgat gacgactcgc ccgtctggct ggtgacgcgc 180  
 ttcaagagg tccgccaagt cctgcgcgat cagcggttcg tgaacaaccc ggccgcaccg 240  
 tccctgggac gctcgatcga cgaaagcccc gcggtcagac ttttgaaat gttgggggtg 300  
 cccgaccatt tccggccgta tctgctcggg togatcctca actacgacgc acccgaccac 360  
 acccggtcc gccgactggt ctcgcgcgcc ttcacggcac gcaagatcac cgacctgcgg 420  
 ccgcgggtcg agcagatcac cgacgacctg ctgacctggc ttcccagca cgccgaggac 480  
 ggtgtggtcg acctcatcca gcacttcgcc taccctcgtc cgatcacctg gatctgcgaa 540  
  
 ctggtcggca tcgccgaagc ggaccgccc caatggcgga agtggggagc cgatctcgtc 600  
 tcgctggagc cggggcggt gatcaccgcg ttcccgcgga tggctcgagca catccatgag 660  
 ctgatccgcg agcggcgcg cgcgctcacc gacgatctgc tcagcgagct gatccgcacc 720  
 catcagcagc acggcggcc gctcagcgac atcgagatgg tcacatgat cctcacgatc 780  
 gtctggccg gccacgagac caccgcccac ctcataggca acggcacggc ggcgtgctc 840  
 acccaccocg accagctgcg cctactcaag gacgatccgg cgctgctgcc gcgcgcgtc 900  
 cagcagctga tgcgctggtg cgggcccgtg cacatgacct agctgcggtt cgcgtccgag 960  
 gacgtcgagg tcgccgggac accgatccac aaggcgacg ccgtacaact catcctggta 1020



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tcggcgaaact tcgacccccg ccactacacc gaccccgacc gtctcgacct gacccgccac 1080
cccgccggcc acgcccagaaa ccatgtgggc ttcggccacg gaatgcacta ctgcctgggt 1140
gccaccctcg ccaaacagga aggcgaagtc gccttctccc gcctcttcac gcactaccg 1200
gaactgtccc tgggcgtcgc ggcggaccag ctggcgcgga cacaggtacc cggcagctgg 1260
cggctggaca ccctgccgct gcgactgggg tga 1293

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&lt;210&gt; 24

&lt;211&gt; 430

&lt;212&gt; PRT

&lt;213&gt; Streptomyces tubercidicus

&lt;400&gt; 24

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Met Ser Ala Leu Ser Asn Ser Pro Leu Ala Ala His Val Gly Lys His
1          5          10          15
Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Thr Asp Pro Phe
          20          25          30
Gly Gly Tyr Gly Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg
          35          40          45
Phe Met Asp Asp Ser Pro Val Trp Leu Val Thr Arg Phe Glu Glu Val
          50          55          60
Arg Gln Val Leu Arg Asp Gln Arg Phe Val Asn Pro Ala Ala Pro
65          70          75          80
Ser Leu Gly Arg Ser Ile Asp Glu Ser Pro Ala Val Arg Leu Leu Glu
          85          90          95
Met Leu Gly Leu Pro Asp His Phe Arg Pro Tyr Leu Leu Gly Ser Ile
          100          105          110
Leu Asn Tyr Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser
          115          120          125
Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Glu
          130          135          140
Gln Ile Thr Asp Asp Leu Leu Thr Arg Leu Pro Glu His Ala Glu Asp
145          150          155          160
Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr
          165          170          175
Val Ile Cys Glu Leu Val Gly Ile Ala Glu Ala Asp Arg Pro Gln Trp
          180          185          190
Arg Lys Trp Gly Ala Asp Leu Val Ser Leu Glu Pro Gly Arg Leu Ser
          195          200          205
Thr Ala Phe Pro Ala Met Val Glu His Ile His Glu Leu Ile Arg Glu
          210          215          220
Arg Arg Gly Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr
225          230          235          240
His Asp Asp Asp Gly Gly Arg Leu Ser Asp Ile Glu Met Val Thr Met
          245          250          255
Ile Leu Thr Ile Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile
          260          265          270
Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu
          275          280          285
Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met
          290          295          300
Arg Trp Cys Gly Pro Val His Met Thr Gln Leu Arg Phe Ala Ser Glu
305          310          315          320
Asp Val Glu Val Ala Gly Thr Pro Ile His Lys Gly Asp Ala Val Gln

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<400> 26  
Met Ser Ala Leu Ser Ser Ser Pro Phe Ala Ala His Val Gly Lys His  
1 5 10 15  
Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Ala Asp Pro Phe

			20					25					30			
Gly	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Gly	Pro	Val	Val	Arg	Gly	Arg	
		35					40					45				
Phe	Met	Asp	Asp	Ser	Pro	Val	Trp	Leu	Val	Thr	Arg	Phe	Glu	Glu	Val	
	50					55					60					
Arg	Gln	Val	Leu	Arg	Asp	Gln	Arg	Phe	Leu	Asn	Asp	Pro	Thr	Ala	Pro	
65					70					75					80	
Ser	Leu	Gly	Arg	Ser	Phe	Asp	Asp	Ser	Pro	Thr	Ala	Arg	Leu	Leu	Glu	
				85				90					95			
Met	Met	Gly	Leu	Pro	Glu	His	Phe	Arg	Pro	Tyr	Leu	Leu	Gly	Ser	Ile	
			100					105					110			
Leu	Asn	Asn	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser	
		115					120					125				
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Glu	
	130					135					140					
Gln	Ile	Ala	Asp	Glu	Leu	Leu	Thr	Arg	Leu	Pro	Glu	Tyr	Ala	Glu	Asp	
145				150						155					160	
Gly	Val	Val	Asp	Leu	Ile	Lys	His	Phe	Ala	Tyr	Pro	Leu	Pro	Ile	Ala	
			165						170					175		
Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Ala	Glu	Ala	Asp	Arg	Pro	Gln	Trp	
		180						185					190			
Arg	Lys	Trp	Gly	Ala	Asp	Leu	Val	Ser	Leu	Gln	Pro	Asp	Arg	Leu	Ser	
		195				200						205				
Thr	Ser	Phe	Pro	Ala	Met	Ile	Glu	His	Ile	His	Glu	Leu	Ile	Arg	Glu	
	210				215						220					
Arg	Arg	Gly	Ala	Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Ala	
225				230						235					240	
His	Asp	Asp	Asp	Gly	Gly	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Met	
			245						250					255		
Ile	Leu	Thr	Val	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile	
			260					265					270			
Gly	Asn	Gly	Thr	Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	Arg	Leu	
		275				280						285				
Leu	Arg	Asp	Asp	Pro	Ala	Leu	Phe	Pro	Arg	Ala	Val	His	Glu	Leu	Leu	
	290				295						300					
Arg	Trp	Cys	Gly	Pro	Val	His	Met	Thr	Gln	Met	Arg	Phe	Ala	Ser	Glu	
305				310						315					320	
Asp	Val	Asp	Ile	Ala	Gly	Thr	Lys	Ile	Arg	Lys	Gly	Asp	Ala	Val	Gln	
			325						330					335		
Leu	Ile	Leu	Val	Ser	Ala	Asn	Phe	Asp	Pro	Arg	His	Tyr	Thr	Asp	Pro	
			340					345					35			

<210> 27

<211> 1293  
 <212> DNA  
 <213> *Streptomyces platensis*

<400> 27  
 atgtcggcat tatccagctc tccgttcgct ggcgatgtcg ggaaacaccc ggggtgagccg 60  
 aatgtgatgg agccggcgct gctcaccgac ccgttcgcgg gctacggcgc gctgcgtgag 120  
 caggccccgg tcgtacgggg ccggttcgtg gacgactcac cgggtctggtt cgtgacgcgc 180  
 ttcgaggagg tccgccaaagt cctgcgcgac cagcgggttcg tgaacaatcc ggccgcgcgc 240  
 cccctggccc catcgccga ggagaacccg ctgaccaggc tgatggacat gctgggcctc 300  
 cccgagcacc tccgcgtcta catgctcggg tcgattctca actacgacgc ccccgaccac 360  
 acccggtcgc gccgtctggt gtcgcgcgcg ttcacggcgc ggaagatcac cgatctgcga 420  
 ccgcgtgtcg agcagatcgc cgacgagctg ctggcccgc tccccgagta cgccgaggac 480  
 ggcgtcgtcg acctcatcca gcatttcgcc taccgctgc cgatcaccgt catctgcgag 540  
 ctggtcggca taccggaagc ggaccgccc cagtggcgga agtggggcgc cgacctcatc 600  
 tcgatggacc cggaccggct cggcgcaacg ttcccgcgga tgatcgagca catccatgag 660  
 atggtccggg agcggcgcg gcgctcacc gatgatctgc tcagcgagct gatccgtacc 720  
 catgacgacg atggcgggcc gctcagcgac gtcgagatgg tcaccatgat cctcacgctc 780  
 gtcctcgccg gtcacgagac caccgcccac ctcatcagca acggcacggc ggcgctgctc 840  
 acccaccgga accagctgcg cctgctcaag gacgaccgg ccctgctccc cggggccgctc 900  
 catgagctga tgcgtggtg cgggcccgtg cagatgacgc agctgcgcta cgcggccgccc 960  
 gacgtcgacc tcgcccgtac gcggatccac aaggcgacg ccgtacaact cctcctgggt 1020  
 gcggcgaaact tcgaccccc ccactacacc gaccccgacc gtctcgatct gacgcgtcac 1080  
 cccgcccggc acgcccagaa ccatgtgggt ttccggccag gtgcgcatta ctgcctgggt 1140  
 gccaccctcg ccaagcagga gggcgaagtc gcgttcggca agctgctcgc gcactaccgc 1200  
 gagatgtccc tgggcacga accggaacgt ctggagcgat tgccgctgcc tggcaactgg 1260  
 cggctgaatt ccctgccgtt gcggctgggg tga 1293

<210> 28  
 <211> 430  
 <212> PRT  
 <213> *Streptomyces platensis*

<400> 28  
 Met Ser Ala Leu Ser Ser Ser Pro Phe Ala Ala His Val Gly Lys His  
 1 5 10 15  
 Pro Gly Glu Pro Asn Val Met Glu Pro Ala Leu Leu Thr Asp Pro Phe  
 20 25 30  
 Ala Gly Tyr Gly Ala Leu Arg Glu Gln Ala Pro Val Val Arg Gly Arg  
 35 40 45  
 Phe Val Asp Asp Ser Pro Val Trp Phe Val Thr Arg Phe Glu Glu Val  
 50 55 60  
 Arg Gln Val Leu Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Ala Pro  
 65 70 75 80  
 Pro Leu Ala Pro Ser Ala Glu Glu Asn Pro Leu Thr Arg Leu Met Asp  
 85 90 95  
 Met Leu Gly Leu Pro Glu His Leu Arg Val Tyr Met Leu Gly Ser Ile  
 100 105 110  
 Leu Asn Tyr Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser  
 115 120 125  
 Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Glu  
 130 135 140  
 Gln Ile Ala Asp Glu Leu Leu Ala Arg Leu Pro Glu Tyr Ala Glu Asp

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<210> 29
<211> 1293
<212> DNA
<213> Streptomyces lydicus
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- 25 -

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catgacgacg  acggcgagccg  gctcagcgac  gtcgagatgg  tcaccatggg  cctcaccgtc  780
gtcctggccg  gccacgagac  caccgcgcac  ctcacgga  acggcacggc  ggccctgctc  840
acccaccccg  accagctgcg  gctgctcaag  gacgaccgg  cgctgctgcc  gcgcgcggtg  900
cacgagttga  tgcgctggtg  cggcccgggtg  cacatgacct  agctgcgcta  cgccgccgag  960
gacgtcgagc  tggcgggcgt  ccggatccgc  aagggggacg  ccgtccagct  catcctggtg  1020
tcggcgaaac  gcgatccgcg  ccactacacc  gaaccgcacc  gtctggacct  gaccgcgcac  1080
cccgccggcc  acgccgagaa  ccattgtggg  ttccggccacg  gggcgacta  ctgtctgggc  1140
gccacgctcg  ccaagcagga  gggcgaggtc  gccctcggcg  ccctgctcag  gcacttcccc  1200
gagctgtcgc  tggcctcgc  gccggacgcc  ctggagcgca  caccggtacc  gggcagctgg  1260
cggtgaatg  cgctgccgct  gcgtctgcgc  tga  1293

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<210> 30
<211> 430
<212> PRT
<213> Streptomyces lydicus

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<400> 30
Met Ser Ala Leu Pro Ser Asn Thr Phe Thr Glu His Val Gly Lys His
1          5          10          15
Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Gly Asp Pro Phe
20          25          30
Ala Gly Tyr Gly Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg
35          40          45
Phe Val Asp Asp Ser Pro Val Trp Phe Val Thr Arg Phe Glu Glu Val
50          55          60
Arg Glu Val Leu Arg Asp Gln Arg Phe Arg Asn Asn Pro Val Ser Ser
65          70          75          80
Ala Pro Asp Ala Asp Pro Glu Asp Thr Pro Leu Ser Arg Leu Met Asp
85          90          95
Met Met Gly Phe Pro Glu His Leu Arg Val Tyr Leu Leu Gly Ser Ile
100          105          110
Leu Asn Asn Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser
115          120          125
Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Ala
130          135          140
Gln Ile Ala Asp Glu Leu Leu Ala Arg Leu Pro Glu His Ala Glu Asp
145          150          155          160
Gly Val Val Asp Leu Ile Gln His Phe Ala Tyr Pro Leu Pro Ile Thr
165          170          175
Val Ile Cys Glu Leu Val Gly Ile Pro Glu Glu Asp Arg Pro Gln Trp
180          185          190
Arg Thr Trp Gly Ala Asp Leu Val Ser Leu Gln Pro Asp Arg Met Ser
195          200          205
Arg Ser Phe Pro Ala Met Ile Asp His Ile His Glu Leu Ile Ala Ala
210          215          220
Arg Arg Arg Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr
225          230          235          240
His Asp Asp Asp Gly Ser Arg Leu Ser Asp Val Glu Met Val Thr Met
245          250          255
Val Leu Thr Val Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile
260          265          270
Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu

```

275                                      280                                      285  
 Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met  
 290                                      295                                      300  
 Arg Trp Cys Gly Pro Val His Met Thr Gln Leu Arg Tyr Ala Ala Glu  
 305                                      310                                      315                                      320  
 Asp Val Glu Leu Ala Gly Val Arg Ile Arg Lys Gly Asp Ala Val Gln  
 325                                      330                                      335  
 Leu Ile Leu Val Ser Ala Asn Arg Asp Pro Arg His Tyr Thr Glu Pro  
 340                                      345                                      350  
 Asp Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His  
 355                                      360                                      365  
 Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Thr Leu Ala  
 370                                      375                                      380  
 Lys Gln Glu Gly Glu Val Ala Leu Gly Ala Leu Leu Arg His Phe Pro  
 385                                      390                                      395                                      400  
 Glu Leu Ser Leu Ala Val Ala Pro Asp Ala Leu Glu Arg Thr Pro Val  
 405                                      410                                      415  
 Pro Gly Ser Trp Arg Leu Asn Ala Leu Pro Leu Arg Leu Arg  
 420                                      425                                      430

<210> 31  
 <211> 1293  
 <212> DNA  
 <213> Streptomyces lydicus

<400> 31  
 atgtcggcat cgaccagctc tcccctcagc gccacgctcg gcaagcacc gggcgaaccc 60  
 catgtgatgg atccggcgct gatcagcgat ccgttcggcg gctacggtgc cctgcgcgag 120  
 cagggaccgg tcgtccgcgg acggttcttc gacgactcgc ccttgtggtt agtgaccgcg 180  
 ttcgaggaag tccgccaggc cctgcgcgac cagcggttcg tgaacaaccc cgccgaccgc 240  
 gcgctcggcg tcgcgcggga ggactccccg cagctgcgcg cgctggcgat gctgggcatc 300  
 cccgagcacc tgcacggcta tctgctcaac tcgatcctca actacgacgc ccccgaccac 360  
 acccggtgc gccgcctggt ctcccgcgcc ttcaccgccc gcaagatcac cgatcttcgg 420  
 ccgcgggtgg cgcagataac cgccgagctg ctggaccgac tcccggagca cgccgaggac 480  
 ggcgtggtcg acctgatcga gcacttcgcc taccgctgc cgatcacggt gatctgcgaa 540  
 cttgtcggca tcgccgcgga ggaccggccc cagtggcggt cctggggcgc cgacctggtc 600  
 tcggtggacc ccgaccgggt cgcccgacc ttcccggcga tgatcgacca catccacgcg 660  
 ctgatcggcc agcggcgggc cgcgctcacc gacgacctgc tcagcgagct gatccggacc 720  
 catgacgacg acggcagccg gctcagcgac gtcgagatgg tcacctggt cctcaccctc 780  
 gtgctggccg gccacgagac caccgcacac ctcatcggca acggcaccgc ggccctgtc 840  
 accaccccg accagctgcg gctgctcaag gacgaccg cgctgctgcc gcgcgccgtc 900  
 cacgagctga tgcgtggtg cgggcccgtg cacgtcacc agctgcggta cgccgcccag 960  
 gacgtcgacc tcgccggcac ccggatccgc aggggcgacg ccgtgcaggc cgtcctggtc 1020  
 tcggcgaacc acgaccgcg ccactacacc gacccgaac gcctggacct gaccggcgag 1080  
 cccgcgggccc gcgccgagaa ccacgtgggc ttcgggcacg gggcgacta ctgcctgggc 1140  
 gccagctcg ccaggcagga gggtaggtc gcctgggccc ccctgttcga ccgctacccc 1200  
 gacctggcgc tggcgggtgc gcccgaggag ctggagcgca ccccggtgcc cggtacctgg 1260  
 cggctgacgt cgctgccggt gcgcctgggc tga 1293

<210> 32  
 <211> 430  
 <212> PRT  
 <213> Streptomyces lydicus

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<400> 32
Met Ser Ala Ser Thr Ser Ser Pro Leu Ser Ala His Val Gly Lys His
1      5      10
Pro Gly Glu Pro His Val Met Asp Pro Ala Leu Ile Ser Asp Pro Phe
20      25      30
Gly Gly Tyr Gly Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg
35      40      45
Phe Phe Asp Asp Ser Pro Leu Trp Leu Val Thr Arg Phe Glu Glu Val
50      55      60
Arg Gln Val Leu Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Asp Pro
65      70      75      80
Ala Leu Gly Val Ala Pro Glu Asp Ser Pro Gln Leu Arg Ala Leu Ala
85      90      95
Met Leu Gly Ile Pro Glu His Leu His Gly Tyr Leu Leu Asn Ser Ile
100     105     110
Leu Asn Tyr Asp Ala Pro Asp His Thr Arg Leu Arg Arg Leu Val Ser
115     120     125
Arg Ala Phe Thr Ala Arg Lys Ile Thr Asp Leu Arg Pro Arg Val Ala
130     135     140
Gln Ile Thr Ala Glu Leu Leu Asp Arg Leu Pro Glu His Ala Glu Asp
145     150     155     160
Gly Val Val Asp Leu Ile Glu His Phe Ala Tyr Pro Leu Pro Ile Thr
165     170     175
Val Ile Cys Glu Leu Val Gly Ile Ala Ala Glu Asp Arg Pro Gln Trp
180     185     190
Arg Ser Trp Gly Ala Asp Leu Val Ser Val Asp Pro Asp Arg Leu Gly
195     200     205
Arg Thr Phe Pro Ala Met Ile Asp His Ile His Ala Leu Ile Gly Gln
210     215     220
Arg Arg Ala Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg Thr
225     230     235     240
His Asp Asp Asp Gly Ser Arg Leu Ser Asp Val Glu Met Val Thr Leu
245     250     255
Val Leu Thr Leu Val Leu Ala Gly His Glu Thr Thr Ala His Leu Ile
260     265     270
Gly Asn Gly Thr Ala Ala Leu Leu Thr His Pro Asp Gln Leu Arg Leu
275     280     285
Leu Lys Asp Asp Pro Ala Leu Leu Pro Arg Ala Val His Glu Leu Met
290     295     300
Arg Trp Cys Gly Pro Val His Val Thr Gln Leu Arg Tyr Ala Ala Glu
305     310     315     320
Asp Val Asp Leu Ala Gly Thr Arg Ile Arg Arg Gly Asp Ala Val Gln
325     330     335
Ala Val Leu Val Ser Ala Asn His Asp Pro Arg His Tyr Thr Asp Pro
340     345     350
Glu Arg Leu Asp Leu Thr Arg Gln Pro Ala Gly Arg Ala Glu Asn His
355     360     365
Val Gly Phe Gly His Gly Ala His Tyr Cys Leu Gly Ala Ser Leu Ala
370     375     380
Arg Gln Glu Gly Glu Val Ala Leu Gly Ala Leu Phe Asp Arg Tyr Pro
385     390     395     400
Asp Leu Ala Leu Ala Val Ala Pro Glu Glu Leu Glu Arg Thr Pro Val
405     410     415

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Pro Gly Thr Trp Arg Leu Thr Ser Leu Pro Val Arg Leu Gly  
 420 425 430

<210> 33  
 <211> 1281  
 <212> DNA  
 <213> Streptomyces tubercidicus

<400> 33  
 atgaactctc cgttcgccgc gcacgtcggg aaacaccccg gcgagccgaa tgtgatggac 60  
 cccgccctga tcaccgaccc gtacacgggc tacggcgcgc tgcgtgagca gggcccggtc 120  
 gtacggggcc gggtcatgga cgactcgccc gtctggctgg tgacgcgggt cgaggagggtc 180  
 cgccagggtc tgcgcgacca gcggttcgtg aacaatcccg cctcgccgtc cctgaactac 240  
 gcgcccagag acaacccgct gacccggctg atggagatgc tgggcctccc cgagcacctc 300  
 cgcgtctacc tgctcggatc gatcctcaac tacgacgccc ccgaccacac ccggctgcgc 360  
 cgtctgggtg cgcgggcggt caccggccgc aagatcaccg acctgcggcc ccgggtcgag 420  
 cagatcgccg acgcgctgct ggcccggctg cccgagcacg ccgaggacgg cgtcgtcgac 480  
 ctcattccagc atttcgcta cccctgcgc atcaccgta tctgcgaact ggtcggcata 540  
 cccgaagcgg accgcccgc gtggcgaaac tggggcgccg acctcatctc gatggatccg 600  
 gaccggctcg gcgcctcgtt cccggcgatg atcgagcaca tccatcagat ggtccgggaa 660  
 cggcgcgagg cgctcaccga cgacctgtc agcgaactga tccgcacca tgacgacgac 720  
 ggcgggcggc tcagcgacgt cgagatggc accatgatcc tcacgctcgt cctcgccggc 780  
 cagagacca ccgcccacct catcagcaac ggcacggcgg cgctgctcac ccaccccgac 840  
 cagctgcgtc tggtaagga cgatccggcc ctctccccc gtgccgtcca cgagctgatg 900  
  
 cgctgggtgc ggccgggtgca catgacccag ctgcgctacg ccaccgccga cgtcgacctc 960  
 gccggcacac cgatccgcca gggcgatgcc gttcaactca tcctgggtatc ggccaacttc 1020  
 gacccccgtc actacaccga cccgaccgc ctcgatctca cccggcacc cgcgggccac 1080  
 gccgagaacc atgtgggttt cggccatgga gcgcactact gcctggggcg cacactcgcc 1140  
 aaacaggaag gtgaagtgc cttcggcaaa ctgctcacgc actaccgga catatcgctg 1200  
 ggcacgccc cggaacacct ggagcggaca ccgctgccgg gcaactggcg gctgaactcg 1260  
 ctgccgggtg ggttggggtg a 1281

<210> 34  
 <211> 426  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 34  
 Met Asn Ser Pro Phe Ala Ala His Val Gly Lys His Pro Gly Glu Pro  
 1 5 10 15  
 Asn Val Met Asp Pro Ala Leu Ile Thr Asp Pro Phe Thr Gly Tyr Gly  
 20 25 30  
 Ala Leu Arg Glu Gln Gly Pro Val Val Arg Gly Arg Phe Met Asp Asp  
 35 40 45  
 Ser Pro Val Trp Leu Val Thr Arg Phe Glu Glu Val Arg Gln Val Leu  
 50 55 60  
 Arg Asp Gln Arg Phe Val Asn Asn Pro Ala Ser Pro Ser Leu Asn Tyr  
 65 70 75 80  
 Ala Pro Glu Asp Asn Pro Leu Thr Arg Leu Met Glu Met Leu Gly Leu  
 85 90 95  
 Pro Glu His Leu Arg Val Tyr Leu Leu Gly Ser Ile Leu Asn Tyr Asp

			100					105					110			
Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser	Arg	Ala	Phe	Thr	
		115					120					125				
Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Glu	Gln	Ile	Ala	Asp	
		130				135					140					
Ala	Leu	Leu	Ala	Arg	Leu	Pro	Glu	His	Ala	Glu	Asp	Gly	Val	Val	Asp	
145					150					155					160	
Leu	Ile	Gln	His	Phe	Ala	Tyr	Pro	Leu	Pro	Ile	Thr	Val	Ile	Cys	Glu	
				165					170					175		
Leu	Val	Gly	Ile	Pro	Glu	Ala	Asp	Arg	Pro	Gln	Trp	Arg	Thr	Trp	Gly	
			180				185						190			
Ala	Asp	Leu	Ile	Ser	Met	Asp	Pro	Asp	Arg	Leu	Gly	Ala	Ser	Phe	Pro	
		195				200					205					
Ala	Met	Ile	Glu	His	Ile	His	Gln	Met	Val	Arg	Glu	Arg	Arg	Glu	Ala	
		210				215					220					
Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Thr	His	Asp	Asp	Asp	
225				230						235					240	
Gly	Gly	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Met	Ile	Leu	Thr	Leu	
				245					250					255		
Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile	Ser	Asn	Gly	Thr	
			260					265					270			
Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	Arg	Leu	Val	Lys	Asp	Asp	
		275				280						285				
Pro	Ala	Leu	Leu	Pro	Arg	Ala	Val	His	Glu	Leu	Met	Arg	Trp	Cys	Gly	
		290				295					300					
Pro	Val	His	Met	Thr	Gln	Leu	Arg	Tyr	Ala	Thr	Ala	Asp	Val	Asp	Leu	
305				310						315					320	
Ala	Gly	Thr	Pro	Ile	Arg	Gln	Gly	Asp	Ala	Val	Gln	Leu	Ile	Leu	Val	
				325				330						335		
Ser	Ala	Asn	Phe	Asp	Pro	Arg	His	Tyr	Thr	Asp	Pro	Asp	Arg	Leu	Asp	
			340				345					350				
Leu	Thr	Arg	His	Pro	Ala	Gly	His	Ala	Glu	Asn	His	Val	Gly	Phe	Gly	
		355				360						365				
His	Gly	Ala	His	Tyr	Cys	Leu	Gly	Ala	Thr	Leu	Ala	Lys	Gln	Glu	Gly	
		370				375					380					
Glu	Val	Ala	Phe	Gly	Lys	Leu	Leu	Thr	His	Tyr	Pro	Asp	Ile	Ser	Leu	
385				390						395					400	
Gly	Ile	Ala	Pro	Glu	His	Leu	Glu	Arg	Thr	Pro	Leu	Pro	Gly	Asn	Trp	
				405					410					415		
Arg	Leu	Asn	Ser	Leu	Pro	Val	Arg	Leu	Gly							

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<210> 35
<211> 195
<212> DNA
<213> Streptomyces tubercidicus
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[illegible]

<210> 36  
 <211> 64  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 36  
 Met Arg Ile Thr Ile Asp Thr Asp Ile Cys Ile Gly Ala Gly Gln Cys  
 1 5 10 15  
 Ala Leu Thr Ala Pro Gly Val Phe Thr Gln Asp Asp Asp Gly Phe Ser  
 20 25 30  
 Ala Leu Leu Pro Gly Arg Glu Asp Gly Ala Gly Asp Pro Leu Val Arg  
 35 40 45  
 Glu Ala Ala Arg Ala Cys Pro Val Gln Ala Ile Thr Val Thr Asp Asp  
 50 55 60

<210> 37  
 <211> 195  
 <212> DNA  
 <213> Streptomyces tubercidicus

<400> 37  
 atgcggatca ccacgcacac cgacatctgc atcggcgccg gccagtgcgc cctgaccgcg 60  
 ccgggagtct tcaccagga cgacgacggt ttcagcgccc tgcgtgcccg ccgcgaggac 120  
 ggcgcgggcg acccgctggt gcgcgaggcc gcccggcct gcccgtgca ggccatttcg 180  
 gtcacggacg actga 195

<210> 38  
 <211> 64  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 38  
 Met Arg Ile Thr Ile Asp Thr Asp Ile Cys Ile Gly Ala Gly Gln Cys  
 1 5 10 15  
 Ala Leu Thr Ala Pro Gly Val Phe Thr Gln Asp Asp Asp Gly Phe Ser  
 20 25 30  
 Ala Leu Leu Pro Gly Arg Glu Asp Gly Ala Gly Asp Pro Leu Val Arg  
 35 40 45  
 Glu Ala Ala Arg Ala Cys Pro Val Gln Ala Ile Ser Val Thr Asp Asp  
 50 55 60

<210> 39  
 <211> 9  
 <212> PRT  
 <213> Artificial Sequence  
 <220>  
 <223> Synthetic peptide.  
 <400> 39

Tyr Pro Tyr Asp Val Pro Asp Tyr Ala  
1 5

<210> 40  
<211> 10  
<212> PRT  
<213> Artificial Sequence  
<220>  
<223> Synthetic peptide.

<400> 40  
Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu  
1 5 10

<210> 41  
<211> 11  
<212> PRT  
<213> Artificial Sequence  
<220>  
<223> Synthetic peptide.

<400> 41  
Gln Pro Glu Leu Ala Pro Glu Asp Pro Glu Asp  
1 5 10

<210> 42  
<211> 11  
<212> PRT  
<213> Artificial Sequence  
<220>  
<223> Synthetic peptide.

<400> 42  
Tyr Thr Asp Ile Glu Met Asn Arg Leu Gly Lys  
1 5 10

<210> 43  
<211> 7  
<212> PRT  
<213> Streptomyces  
<220>  
<221> misc\_feature  
<222>  
<223> Streptomyces consensus sequence

<400> 43  
Ile Ala Gly His Glu Thr Thr  
1 5

<210> 44  
<211> 20  
<212> DNA  
<213> Artificial Sequence  
<220>  
<221> misc\_feature  
<222> (6)..(18)  
<223> Nucleotides 6, 9 and 18 are "s" wherein "s" = g or c.

<400> 44  
atcgcsggsc acgagacsac

20

<210> 45  
<211> 7  
<212> PRT  
<213> Streptomyces  
<220>  
<221> misc\_feature  
<222>  
<223> Streptomyces consensus sequence

<400> 45  
Val Ala Gly His Glu Thr Thr  
1 5

<210> 46  
<211> 20  
<212> DNA  
<213> Artificial Sequence  
<220>  
<221> misc\_feature  
<222> (3)..(18)  
<223> Nucleotides 3, 6, 9, and 18 are "s" wherein "s" = g or c.

<400> 46  
gtsgcsggsc acgagacsac

20

<210> 47  
<211> 7  
<212> PRT  
<213> Streptomyces  
<220>  
<221> misc\_feature  
<222>  
<223> Streptomyces consensus sequence

<400> 47  
Leu Ala Gly His Glu Thr Thr  
1 5

<210> 48  
 <211> 20  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (3)..(18)  
 <223> Nucleotides 3, 6, 9, and 18 are "s" wherein "s" = g or c.

<400> 48  
 ctsgcsgggsc acgagacsac 20

<210> 49  
 <211> 9  
 <212> PRT  
 <213> Streptomyces  
 <220>  
 <221> misc\_feature  
 <222>  
 <223> Streptomyces consensus sequence

<400> 49  
 Leu Leu Leu Ile Ala Gly His Glu Thr  
 1 5

<210> 50  
 <211> 25  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (2)..(17)  
 <223> Nucleotides 2, 5, 8, 14, and 17 are "s" wherein "s" = g or c.

<400> 50  
 tsctsctsat cgcsggscac gagac 25

<210> 51  
 <211> 9  
 <212> PRT  
 <213> Streptomyces  
 <220>  
 <221> misc\_feature  
 <222>  
 <223> Streptomyces consensus sequence

<400> 51  
 His Gln Cys Leu Gly Gln Asn Leu Ala  
 1 5

<210> 52  
 <211> 26  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (12)..(24)  
 <223> Nucleotides 12, 15, and 24 are "s" wherein "s" = g or c.

<400> 52  
 gtggtcacgg asccstgctt ggascg

26

<210> 53  
 <211> 8  
 <212> PRT  
 <213> Streptomyces  
 <220>  
 <221> misc\_feature  
 <222>  
 <223> Streptomyces consensus sequence

<400> 53  
 Phe Gly His Gly Val His Gln Cys  
 1 5

<210> 54  
 <211> 24  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (6)..(15)  
 <223> Nucleotides 6, 12, and 15 are "s" wherein "s" = g or c.

<400> 54  
 aagccsgtgc cscasgtggt cacg

24

<210> 55  
 <211> 8  
 <212> PRT  
 <213> Streptomyces  
 <220>  
 <221> misc\_feature  
 <222>  
 <223> Streptomyces consensus sequence

<400> 55

Phe Gly Phe Gly Val His Gln Cys  
 1 5

<210> 56  
 <211> 24  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (6)..(15)  
 <223> Nucleotides 6, 12, and 15 are "s" wherein "s" = g or c.

<400> 56  
 aaggcsaagc cscasgtggt cacg

24

<210> 57  
 <211> 8  
 <212> PRT  
 <213> Streptomyces  
 <220>  
 <221> misc\_feature  
 <222>  
 <223> Streptomyces consensus sequence

<400> 57  
 Phe Gly His Gly Ile His Gln Cys  
 1 5

<210> 58  
 <211> 24  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (6)..(12)  
 <223> Nucleotides 6 and 12 are "s" wherein "s" = g or c.

<400> 58  
 aagccsgtgc cstaggtggt cacg

24

<210> 59  
 <211> 8  
 <212> PRT  
 <213> Streptomyces  
 <220>  
 <221> misc\_feature  
 <222>  
 <223> Streptomyces consensus sequence



&lt;400&gt; 59

Phe Gly His Gly Val His Phe Cys  
 1 5

&lt;210&gt; 60

&lt;211&gt; 24

&lt;212&gt; DNA

&lt;213&gt; Artificial Sequence

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; (6)..(15)

&lt;223&gt; Nucleotides 6, 12, and 15 are "s" wherein "s" = g or c.

&lt;400&gt; 60

aagccsgtgc cscasgtgaa gacg

24

&lt;210&gt; 61

&lt;211&gt; 24

&lt;212&gt; PRT

&lt;213&gt; Streptomyces tubercidicus

&lt;400&gt; 61

His Pro Gly Glu Pro Asn Val Met Asp Pro Ala Leu Ile Thr Asp Pro  
 1 5 10 15  
 Phe Thr Gly Tyr Gly Ala Leu Arg  
 20

&lt;210&gt; 62

&lt;211&gt; 21

&lt;212&gt; PRT

&lt;213&gt; Streptomyces tubercidicus

&lt;400&gt; 62

Phe Val Asn Asn Pro Ala Ser Pro Ser Leu Asn Tyr Ala Pro Glu Asp  
 1 5 10 15  
 Asn Pro Leu Thr Arg  
 20

&lt;210&gt; 63

&lt;211&gt; 19

&lt;212&gt; PRT

&lt;213&gt; Streptomyces tubercidicus

&lt;400&gt; 63

Leu Leu Thr His Tyr Pro Asp Ile Ser Leu Gly Ile Ala Pro Glu His  
 1 5 10 15  
 Leu Glu Arg

<210> 64  
 <211> 17  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 64  
 Val Tyr Leu Leu Gly Ser Ile Leu Asn Tyr Asp Ala Pro Asp His Thr  
 1 5 10 15  
 Arg

<210> 65  
 <211> 13  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 65  
 Thr Trp Gly Ala Asp Leu Ile Ser Met Asp Pro Asp Arg  
 1 5 10

<210> 66  
 <211> 13  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 66  
 Glu Ala Leu Thr Asp Asp Leu Leu Ser Glu Leu Ile Arg  
 1 5 10

<210> 67  
 <211> 12  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 67  
 Phe Met Asp Asp Ser Pro Val Trp Leu Val Thr Arg  
 1 5 10

<210> 68  
 <211> 12  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 68  
 Leu Met Glu Met Leu Gly Leu Pro Glu His Leu Arg  
 1 5 10

<210> 69

<211> 11  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 69  
Val Glu Gln Ile Ala Asp Ala Leu Leu Ala Arg  
1 5 10

<210> 70  
<211> 11  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 70  
Leu Val Lys Asp Asp Pro Ala Leu Leu Pro Arg  
1 5 10

<210> 71  
<211> 8  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 71  
Asp Asp Pro Ala Leu Leu Pro Arg  
1 5

<210> 72  
<211> 8  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 72  
Thr Pro Leu Pro Gly Asn Trp Arg  
1 5

<210> 73  
<211> 7  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 73  
Leu Asn Ser Leu Pro Val Arg  
1 5

<210> 74  
<211> 7  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 74  
Ile Thr Asp Leu Arg Pro Arg  
1 5

<210> 75  
<211> 7  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 75  
Glu Gln Gly Pro Val Val Arg  
1 5

<210> 76  
<211> 7  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 76  
Ala Val His Glu Leu Met Arg  
1 5

<210> 77  
<211> 5  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 77  
Ala Phe Thr Ala Arg  
1 5

<210> 78  
<211> 5  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 78  
Phe Glu Glu Val Arg  
1 5

<210> 79  
<211> 7  
<212> PRT  
<213> Streptomyces tubercidicus

<400> 79  
Pro Gly Glu Asp Asn Val Met  
1 5

<210> 80  
 <211> 21  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (3)..(18)  
 <223> Nucleotides 3, 6, 12, and 18 are "s" wherein "s" = c or g.  
 <220>  
 <221> misc\_feature  
 <222> (9)..(9)  
 <223> Nucleotide 9 is "r" wherein "r" = a or g.  
 <220>  
 <221> misc\_feature  
 <222> (15)..(15)  
 <223> Nucleotide 15 is "y" wherein "y" = c or t.

<400> 80  
 ccsggsgarc csaaygtsat g 21

<210> 81  
 <211> 7  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 81  
 Ala Leu Ile Thr Asp Pro Phe  
 1 5  
 <210> 82  
 <211> 21  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (3)..(18)  
 <223> Nucleotides 3, 6, 12, and 18 are "s" wherein "s" = c or g.

<400> 82  
 gcsctsatya csgacccstt c 21

<210> 83  
 <211> 8  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 83  
 Phe Met Asp Asp Ser Pro Val Trp  
 1 5

<210> 84  
 <211> 24  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (13)..(13)  
 <223> Nucleotide 13 is "w" wherein "w" = a or t.  
 <220>  
 <221> misc\_feature  
 <222> (14)..(21)  
 <223> Nucleotides 14, 15, 18, and 21 are "s" wherein "s" = c or g.

<400> 84  
 ttcatggacg acwssccsgt stgg

24

<210> 85  
 <211> 8  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 85  
 Leu Asn Tyr Asp Ala Pro Asp His  
 1 5

<210> 86  
 <211> 24  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (3)..(18)  
 <223> Nucleotides 3, 15 and 18 are "s" wherein "s" = c or g.  
 <220>  
 <221> misc\_feature  
 <222> (6)..(9)  
 <223> Nucleotides 6 and 9 are "y" wherein "y" = c or t.

<400> 86  
 ctsaaytayg acgcscsga ccac

24

<210> 87  
 <211> 8  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 87  
 Val Glu Gln Ile Ala Asp Ala Leu  
 1 5

<210> 88  
 <211> 24  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (3)..(24)  
 <223> Nucleotides 3, 15, 21, and 24 are "s" wherein "s" = c or g.  
 <220>  
 <221> misc\_feature  
 <222> (12)..(12)  
 <223> Nucleotide 12 is "y" wherein "y" = c or t.  
 <220>  
 <221> misc\_feature  
 <222> (6)..(6)  
 <223> Nucleotide 6 is "r" wherein "r" = a or g.

<400> 88  
 gtsgarcaga tygcs gacgc scts 24

<210> 89  
 <211> 8  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 89  
 Asp Leu Ile Ser Met Asp Pro Asp  
 1 5

<210> 90  
 <211> 24  
 <212> DNA  
 <213> Artificial Sequence  
 <220>  
 <221> misc\_feature  
 <222> (6)..(21)  
 <223> Nucleotides 6, 11, 12, and 21 are "s" wherein "s" = c or g.  
 <220>  
 <221> misc\_feature  
 <222> (9)..(9)  
 <223> Nucleotide 9 is "r" wherein "r" = a or g.  
 <220>  
 <221> misc\_feature  
 <222> (10)..(10)  
 <223> Nucleotide 10 is "w" wherein "w" = a or t.

<400> 90  
 ctggastarw sstacctggg sctg 24

<210> 91  
 <211> 36

<212> DNA  
 <213> Streptomyces tubercidicus  
  
 <400> 91  
 agattaatta atgtcggaaat taatgaactg tccgtt 36  
  
 <210> 92  
 <211> 32  
 <212> DNA  
 <213> Streptomyces tubercidicus  
  
 <400> 92  
 aaactcacc caaccgcacc ggcagcgagt tc 32  
  
 <210> 93  
 <211> 7  
 <212> PRT  
 <213> Streptomyces tubercidicus  
  
 <400> 93  
 Met Ser Glu Leu Met Asn Ser  
 1 5  
  
 <210> 94  
 <211> 1293  
 <212> DNA  
 <213> Streptomyces tubercidicus  
  
 <400> 94  
 atgtcggcaa tatccagctc cccgttcgcc gcacacgtcg gaaagcatcc cggcgagccg 60  
 aatgtgatgg acccggcgct gatcaccgac ccgttcggcg gctacggcgc actgcgtgag 120  
 caaggccccg tcctaccggg ccggttcgat gacgactcac ccgtctggct cgtgacgcgc 180  
 ttcgaagagg tccgccaagt cctgcgcgat cagcggttcc tgaacaaccc ggccgcgtcg 240  
 tcaccggggc attcgatcga cgagagcccc acggccaggc tgctggacat gatggggatg 300  
 cccgaacatt tccggccgta tctgatgggg tcgatcctca acaacgacgc ccccgaccac 360  
 acccggctgc gccgtctggt gtcacgcgcg ttcacggcac gcaagatcac cgatctgcgg 420  
 ccgcgggtcg agcagctcgc cgacgagctg ctggcccggc ttcccagca cgcgcaggac 480  
 ggtgtggtcg acctgatcaa gcaacttcgcc tatccctcgc cgatcaccgt gatctgcgaa 540  
 ctggtcggca tcccgggaagc ggaccgcccg caatggcgga agtggggcgc cgacctcgtt 600  
 tcgctgcagc cggagcggct cagcacctcg ttcccggcga tgatcgagca catccatgaa 660  
 ctgatccgcg agcggcgcgg cgcgctcacc gacgatctgc tcagcgagct gatccgtacc 720  
 catgacgacg acggcagccg gctcagcgac gtcgagatgg tcaccatggt cctcaccgtc 780  
 gtcctggccg gccacgagac caccgcccac ctgataggca acggcacggc ggcgctgctc 840  
 acccaccctg accagctcgc cctgggtcaag gacgaccggg agctgcttcc gcgtgcgcgtc 900  
 caccgagctgc tgcgctgggt cgggcccggc cagatgaccc agctgcggta cgcctccgag 960  
 gatgtcgaga tcgcccggac gccgatccgt aaggcgacg ccgtacaact catcctggta 1020  
 tcggcgaaact tcgacccccg cactacacc gcccccgaac gctcgcacct gaccgcccac 1080  
 cccgcccggc acgcccagaa ccatgtgggc ttccggccacg gaatgcacta ctgcctgggc 1140  
 gccaccctcg ccaaacagga gggcgaagtc gcgttcggca agctcttcac gcactaccctg 1200  
 gagctgtcgc tggccgtcgc accggacgag ttggagcgaa cgcgggtgcc cggcagctgg 1260



cggttggttgcgctgccgggt gcggttggggg tga

1293

&lt;210&gt; 95

&lt;211&gt; 430

&lt;212&gt; PRT

&lt;213&gt; Streptomyces tubercidicus

&lt;400&gt; 95

Met	Ser	Ala	Ile	Ser	Ser	Ser	Pro	Phe	Ala	Ala	His	Val	Gly	Lys	His
1				5					10					15	
Pro	Gly	Glu	Pro	Asn	Val	Met	Asp	Pro	Ala	Leu	Ile	Thr	Asp	Pro	Phe
			20					25					30		
Gly	Gly	Tyr	Gly	Ala	Leu	Arg	Glu	Gln	Gly	Pro	Val	Leu	Pro	Gly	Arg
		35					40					45			
Phe	Met	Asp	Asp	Ser	Pro	Val	Trp	Leu	Val	Thr	Arg	Phe	Glu	Glu	Val
	50					55					60				
Arg	Gln	Val	Leu	Arg	Asp	Gln	Arg	Phe	Leu	Asn	Asn	Pro	Ala	Ala	Ser
65					70					75					80
Ser	Pro	Gly	His	Ser	Ile	Asp	Glu	Ser	Pro	Thr	Ala	Arg	Leu	Leu	Asp
				85					90					95	
Met	Met	Gly	Met	Pro	Glu	His	Phe	Arg	Pro	Tyr	Leu	Met	Gly	Ser	Ile
			100					105					110		
Leu	Asn	Asn	Asp	Ala	Pro	Asp	His	Thr	Arg	Leu	Arg	Arg	Leu	Val	Ser
		115					120					125			
Arg	Ala	Phe	Thr	Ala	Arg	Lys	Ile	Thr	Asp	Leu	Arg	Pro	Arg	Val	Glu
	130					135					140				
Gln	Leu	Ala	Asp	Glu	Leu	Leu	Ala	Arg	Leu	Pro	Glu	His	Ala	Glu	Asp
145					150					155					160
Gly	Val	Val	Asp	Leu	Ile	Lys	His	Phe	Ala	Tyr	Pro	Leu	Pro	Ile	Thr
				165					170					175	
Val	Ile	Cys	Glu	Leu	Val	Gly	Ile	Pro	Glu	Ala	Asp	Arg	Pro	Gln	Trp
			180					185					190		
Arg	Lys	Trp	Gly	Ala	Asp	Leu	Val	Ser	Leu	Gln	Pro	Glu	Arg	Leu	Ser
		195					200					205			
Thr	Ser	Phe	Pro	Ala	Met	Ile	Glu	His	Ile	His	Glu	Leu	Ile	Arg	Glu
	210					215					220				
Arg	Arg	Gly	Ala	Leu	Thr	Asp	Asp	Leu	Leu	Ser	Glu	Leu	Ile	Arg	Thr
225					230					235					240
His	Asp	Asp	Asp	Gly	Ser	Arg	Leu	Ser	Asp	Val	Glu	Met	Val	Thr	Met
				245					250					255	
Val	Leu	Thr	Val	Val	Leu	Ala	Gly	His	Glu	Thr	Thr	Ala	His	Leu	Ile
			260					265					270		
Gly	Asn	Gly	Thr	Ala	Ala	Leu	Leu	Thr	His	Pro	Asp	Gln	Leu	Arg	Leu
		275					280					285			
Val	Lys	Asp	Asp	Pro	Glu	Leu	Leu	Pro	Arg	Ala	Val	His	Glu	Leu	Leu
	290					295					300				
Arg	Trp	Cys	Gly	Pro	Val	Gln	Met	Thr	Gln	Leu	Arg	Tyr	Ala	Ser	Glu
305					310					315					320
Asp	Val	Glu	Ile	Ala	Gly	Thr	Pro	Ile	Arg	Lys	Gly	Asp	Ala	Val	Gln
				325					330					335	
Leu	Ile	Leu	Val	Ser	Ala	Asn	Phe	Asp	Pro	Arg	His	Tyr	Thr	Ala	Pro
			340					345						350	

Glu Arg Leu Asp Leu Thr Arg His Pro Ala Gly His Ala Glu Asn His  
           355                                  360                                  365  
 Val Gly Phe Gly His Gly Met His Tyr Cys Leu Gly Ala Thr Leu Ala  
           370                                  375                                  380  
 Lys Gln Glu Gly Glu Val Ala Phe Gly Lys Leu Phe Thr His Tyr Pro  
 385                                  390                                  395                                  400  
 Glu Leu Ser Leu Ala Val Ala Pro Asp Glu Leu Glu Arg Thr Pro Val  
                                   405                                  410                                  415  
 Pro Gly Ser Trp Arg Leu Asp Ser Leu Pro Val Arg Leu Gly  
                                   420                                  425                                  430

<210> 96  
 <211> 18  
 <212> DNA  
 <213> Artificial Sequence

<400> 96  
 cgscscscsc tswssaas 18

<210> 97  
 <211> 21  
 <212> DNA  
 <213> Artificial Sequence

<400> 97  
 sassgcstts bcccartgyt c 21

<210> 98  
 <211> 1266  
 <212> DNA  
 <213> Streptomyces tubercidicus

<400> 98  
 gtgggtcgacg cacaccagac gttcgtcatc gtcgggggtg gcctggccgg cgcaaaggcc 60  
 gcggagactc tccgcgcgga ggggttcacc ggccgggtga tcctcatctg tgacgagcgc 120  
 gaccacccgt acgagcgccc cccgctctcc aaggggttcc tgctcggcaa ggaagagcgc 180  
 gacagcgtgt tcgtccatga gccgcctcgg tacgcccagg cacagatcga actgcacctg 240  
 ggccagcccc cgtccgcct cgaccccgag ggcaggaccg tccgcctcgg cgacggcacc 300  
 ctgatcgctt acgacaagct gctgctggcc accggcgccg aaccgcggcg cctggacatc 360  
 cccggcaccg gcctggccgg cgtgcaccac ctgcgccgcc tcgcccacgc cgaacgggtg 420  
 cgcggcgtcc tggcctccct cgcccgcgac aacggccatc tggatgatcg cggagccggc 480  
 tggatcgggc tggaggtcgc cgcccgggcc cgtcctacg gcgccgaggt gaccgtcgtc 540  
 gaggccgccc cgacgccgct gcacggcate ctggggcccc aactcggcgg tctgttcacc 600  
 gatctgcacc gcgagcacgg cgtccgcttc cacttcggcg cccgcttcac cgagatcgtc 660  
 ggagagggcg gcatggtgct cgccgtgcgc accgacgacg gcgaggaaca cccgcccac 720  
 gatgtgctcg ccgcgatcgg cgccgccccg cgcaccgcgc tcgccgaaca ggccgggctg 780  
 gatctcgccg acccgagac cgccggcggg gtggccgtcg acgcggcgct gcgcacctcc 840  
 gaccggtaca tctacgccgc cgggtgacgtc gccgcgccg accaccgct gctggacacc 900

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cggctgcggg tcgaacactg ggccaacgcc ctcaacggcg gcccggccgc cgcccgccgc 960
atgctcggcc aggacatcag ctacgaccgc atcccgctact tcttctccga ccagtacgac 1020
gtcggcatgg agtactccgg ctacgccccg cccggctcgt acgcccagggt cgtctgccgc 1080
ggcgacgtcg ccaagcggga gttcatcgcc ttctggctgg cggcggacgg ccggctgctc 1140
gcgggcatga acgtcaacgt ctgggacgtc gccgagtcca tccagcaact catccgctcc 1200
ggggcgccgt tggagcccgg cgcactggcc gatccgcagg ttccgctggc ggcactgctc 1260
ccgtag                                     1266

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&lt;210&gt; 99

&lt;211&gt; 421

&lt;212&gt; PRT

&lt;213&gt; Streptomyces tubercidicus

&lt;400&gt; 99

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Val Val Asp Ala His Gln Thr Phe Val Ile Val Gly Gly Gly Leu Ala
1      5      10      15
Gly Ala Lys Ala Ala Glu Thr Leu Arg Ala Glu Gly Phe Thr Gly Arg
20      25      30
Val Ile Leu Ile Cys Asp Glu Arg Asp His Pro Tyr Glu Arg Pro Pro
35      40      45
Leu Ser Lys Gly Phe Leu Leu Gly Lys Glu Glu Arg Asp Ser Val Phe
50      55      60
Val His Glu Pro Ala Trp Tyr Ala Gln Ala Gln Ile Glu Leu His Leu
65      70      75      80
Gly Gln Pro Ala Val Arg Leu Asp Pro Glu Gly Arg Thr Val Arg Leu
85      90      95
Gly Asp Gly Thr Leu Ile Ala Tyr Asp Lys Leu Leu Leu Ala Thr Gly
100     105     110
Ala Glu Pro Arg Arg Leu Asp Ile Pro Gly Thr Gly Leu Ala Gly Val
115     120     125
His His Leu Arg Arg Leu Ala His Ala Glu Arg Leu Arg Gly Val Leu
130     135     140
Ala Ser Leu Gly Arg Asp Asn Gly His Leu Val Ile Ala Gly Ala Gly
145     150     155     160
Trp Ile Gly Leu Glu Val Ala Ala Ala Ala Arg Ser Tyr Gly Ala Glu
165     170     175
Val Thr Val Val Glu Ala Ala Pro Thr Pro Leu His Gly Ile Leu Gly
180     185     190
Pro Glu Leu Gly Gly Leu Phe Thr Asp Leu His Arg Glu His Gly Val
195     200     205
Arg Phe His Phe Gly Ala Arg Phe Thr Glu Ile Val Gly Glu Gly Gly
210     215     220
Met Val Leu Ala Val Arg Thr Asp Asp Gly Glu Glu His Pro Ala His
225     230     235     240
Asp Val Leu Ala Ala Ile Gly Ala Ala Pro Arg Thr Ala Leu Ala Glu
245     250     255
Gln Ala Gly Leu Asp Leu Ala Asp Pro Glu Thr Gly Gly Gly Val Ala
260     265     270
Val Asp Ala Ala Leu Arg Thr Ser Asp Pro Tyr Ile Tyr Ala Ala Gly
275     280     285
Asp Val Ala Ala Ala Asp His Pro Leu Leu Asp Thr Arg Leu Arg Val
290     295     300

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Glu His Trp Ala Asn Ala Leu Asn Gly Gly Pro Ala Ala Ala Arg Ala  
 305 310 315 320  
 Met Leu Gly Gln Asp Ile Ser Tyr Asp Arg Ile Pro Tyr Phe Phe Ser  
 325 330 335  
 Asp Gln Tyr Asp Val Gly Met Glu Tyr Ser Gly Tyr Ala Pro Pro Gly  
 340 345 350  
 Ser Tyr Ala Gln Val Val Cys Arg Gly Asp Val Ala Lys Arg Glu Phe  
 355 360 365  
 Ile Ala Phe Trp Leu Ala Ala Asp Gly Arg Leu Leu Ala Gly Met Asn  
 370 375 380  
 Val Asn Val Trp Asp Val Ala Glu Ser Ile Gln Gln Leu Ile Arg Ser  
 385 390 395 400  
 Gly Ala Pro Leu Glu Pro Gly Ala Leu Ala Asp Pro Gln Val Pro Leu  
 405 410 415  
 Ala Ala Leu Leu Pro  
 420

&lt;210&gt; 100

&lt;211&gt; 1314

&lt;212&gt; DNA

&lt;213&gt; Streptomyces tubercidicus

&lt;400&gt; 100

atgcccgtcg cacgccgccg ccttcgacct ccgcaccgga gggcgacct gcctgcccgc 60  
 ccgccggggc gtgcgcaccc accccgtgac cgtccaggac ggcatgatct acgtccatca 120  
 cgccgcggag gagggcaccg ccgcatgaag tcggctcgctg tcatcggggc ctgctggtgc 180  
 ggctgtacg ccgcgcggtc cctgcgttcc caggggttcg acggccgcct ggtgatcgtc 240  
 ggggacgagt gccacggccc ctacgaccgg cccccgtgt ccaaggactt cctcaccggc 300  
 gccaccgacc cgggcccgaact cgccctggcc gacgccgagg agatcgccga actcgacgcc 360  
 gaatggctgc tgggacaccg ggccaccggg ctcgacaccg gcggacgcac ggtgctgctc 420  
 gatggcgggc ggtccctgac caccgacggc gtggtcctcg ccaccggcgc cgccccgcgc 480  
 ctgctccccg gaccggtgcc cgccggggtc cacaccctgc gcaccctcga cgacgccag 540  
 gcgtctcgtg cggatctggc gccggcgccg gtccgggtcg tggatgatcg cggcggcttc 600  
 atcggcgccg aggtcgctc gtctgcgcg gccctaggcc atgacgtcac cgtggtcgag 660  
 gccgcgcgcg tccccctcgt cccccaactc ggccacgcca tggccgagat ctgcgcgcgc 720  
 ctgcatgcgg accacggcgt cacgctgctc accggaaccg gtgtcgcccg gctgcgcagc 780  
 gagggcgacg gccggcgcgt caccggcgtc gagctgaccg acggccgcct gctccccgcc 840  
 gacgtggtcg tcgtcgcat cggggtacgc cccgcaccg cctggctcac ggactccgga 900  
 ctgccgctcg acgacggtgt gctctgcgac gcgggctgtg tcaccccgct gcccgccgctc 960  
 gtggccgctg gcgacgtcgc caggggtggac ggcgcccggt ccgagcactg gaccagcgcc 1020  
 accgaacagg ccgccgtggc ggcgcggaac ctgctggccg gcagcaccgt cgcgaccac 1080  
 cggagcctgc cgtacttctg gtccgaccag tacggcgctc gcattccagt cgcgggccac 1140  
 cggctgccc cccgacacacc gcgcgtcctc gaaggctccc ccgacgaccg cagcttctc 1200  
 gcctgttacg aacggggacg acgcaccacc gcggtgctcg ccctcaaccg gccccgccc 1260  
 ttcatgcggc tccgccgcga actcgcccgc accgcctgt cggccaccac ctga 1314

&lt;210&gt; 101

&lt;211&gt; 437

&lt;212&gt; PRT

&lt;213&gt; Streptomyces tubercidicus

&lt;400&gt; 101

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Met Pro Ala Ala Arg Arg Leu Arg Pro Pro His Arg Ser Gly Asp
1      5      10      15
Leu Pro Ala Arg Pro Pro Gly Arg Ala His Pro Pro Arg Asp Arg Pro
20      25      30
Gly Arg His Asp Leu Arg Pro Ser Arg Arg Gly Gly Gly His Arg Arg
35      40      45
Met Lys Ser Val Ala Val Ile Gly Ala Ser Leu Ala Gly Leu Tyr Ala
50      55      60
Ala Arg Ser Leu Arg Ser Gln Gly Phe Asp Gly Arg Leu Val Ile Val
65      70      75      80
Gly Asp Glu Cys His Gly Pro Tyr Asp Arg Pro Pro Leu Ser Lys Asp
85      90      95
Phe Leu Thr Gly Ala Thr Asp Pro Gly Arg Leu Ala Leu Ala Asp Ala
100     105     110
Glu Glu Ile Ala Glu Leu Asp Ala Glu Trp Leu Leu Gly Thr Arg Ala
115     120     125
Thr Gly Leu Asp Thr Gly Gly Arg Thr Val Leu Leu Asp Gly Gly Arg
130     135     140
Ser Leu Thr Thr Asp Gly Val Val Leu Ala Thr Gly Ala Ala Pro Arg
145     150     155     160
Leu Leu Pro Gly Pro Val Pro Ala Gly Val His Thr Leu Arg Thr Leu
165     170     175
Asp Asp Ala Gln Ala Leu Arg Ala Asp Leu Ala Pro Ala Pro Val Arg
180     185     190
Val Val Val Ile Gly Gly Gly Phe Ile Gly Ala Glu Val Ala Ser Ser
195     200     205
Cys Ala Ala Leu Gly His Asp Val Thr Val Val Glu Ala Ala Pro Leu
210     215     220
Pro Leu Val Pro Gln Leu Gly His Ala Met Ala Glu Ile Cys Ala Ala
225     230     235     240
Leu His Ala Asp His Gly Val Thr Leu Leu Thr Gly Thr Gly Val Ala
245     250     255
Arg Leu Arg Ser Glu Gly Asp Gly Arg Arg Val Thr Gly Val Glu Leu
260     265     270
Thr Asp Gly Arg Leu Leu Pro Ala Asp Val Val Val Val Gly Ile Gly
275     280     285
Val Arg Pro Arg Thr Ala Trp Leu Thr Asp Ser Gly Leu Pro Leu Asp
290     295     300
Asp Gly Val Leu Cys Asp Ala Gly Cys Val Thr Pro Leu Pro Ala Val
305     310     315     320
Val Ala Val Gly Asp Val Ala Arg Val Asp Gly Ala Arg Ala Glu His
325     330     335
Trp Thr Ser Ala Thr Glu Gln Ala Ala Val Ala Ala Arg Asn Leu Leu
340     345     350
Ala Gly Ser Thr Val Ala Thr His Arg Ser Leu Pro Tyr Phe Trp Ser
355     360     365
Asp Gln Tyr Gly Val Arg Ile Gln Phe Ala Gly His Arg Leu Pro Thr
370     375     380
Asp Thr Pro Arg Val Leu Glu Gly Ser Pro Asp Asp Arg Ser Phe Leu
385     390     395     400
Ala Cys Tyr Glu Arg Asp Gly Arg Thr Thr Ala Val Leu Ala Leu Asn
405     410     415

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Arg Pro Arg Pro Phe Met Arg Leu Arg Arg Glu Leu Ala Arg Thr Ala  
                   420                  425                  430  
 Leu Ser Ala Thr Thr  
                   435

<210> 102  
 <211> 1233  
 <212> DNA  
 <213> Streptomyces tubercidicus

<400> 102  
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 gagacactgc gcgcggaggg cttcggcggc cccgtcctgc tgctgggcga cgagcgcgag 120  
 cgteccctacg agcggccgcc gctgtccaag ggctacctct tgggcacctc cgagcgggag 180  
 aaggcgtagc tccatccgcc ccagtggtag gccgagcacg acgtcgatct gcggtggggc 240  
 aacgccgtca ccgccctcga ccgggccggc caccgaggtga ccctcgccga cggcagccgg 300  
 ctgggctacg ccaagctgct gctggccacc gggtccactc cgcgcgggct gccggtgccc 360  
 ggcgccgacc tcgacggggg ccacacgctg cggtagcttg cggacagcga ccgcctcaag 420  
 gacctcttcc ggtccgcgtc ccgtagcttg gtgatcggcg gcggtggat cggcctggag 480  
 accacggccg ccgcgcgtgc ggcgggggtc gaggtgaccg tgctggagtc ggcgcgcgtg 540  
 cccctgctgg ggggtgctgg ccgcgaggtc gccaggtct tcgccgatct gcacaccgag 600  
 caggtgtcgc cgctgcgctg cgacaccag gtcacggaga tcaccggcac gaacggcgcg 660  
 gtcgacgggg tacggctggc cgacggcacc cggatcgcgg ccgacgcggt gatcgctggc 720  
 gtcgggatca cccccaactc cgagacggcc gccgcggccg ggctcaaggc cgacaacggc 780  
 gtgctcgtgg acgagcggct gtgctctctc caccgggaca tctacgcgcg cggcgacgtc 840  
 gccaacgcct accaccccct cctgggcaag cacctccgcg tcgagcactg ggccaacgcc 900  
 ctccaccagc cgaagaccgc ggcccgggcc atgctgggcg gggaggccgg ctacgaccgg 960  
 ctgccgtact tcttcaccga ccagtacgac ctgggcatgg agtacacggg gcatgtggag 1020  
 ccgggcgggt acgaccgcgt ggtgttcgcg ggcgacaccg gtgcccgcga gttcatcgcc 1080  
 ttctggctct ccggcggccg ggtgctggcc gggatgaatg tgaacgtatg ggacgtcacc 1140  
 gaccgatcc gggccctggg ggcgagcggg cgggcccgtg accccgagcg gctcgccgac 1200  
 gcggacgtac cgctggcgga tctggtcccc tga 1233

<210> 103  
 <211> 410  
 <212> PRT  
 <213> Streptomyces tubercidicus

<400> 103  
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 Ala Lys Ala Ala Glu Thr Leu Arg Ala Glu Gly Phe Gly Gly Pro Val  
           20                  25                  30  
 Leu Leu Leu Gly Asp Glu Arg Glu Arg Pro Tyr Glu Arg Pro Pro Leu  
           35                  40                  45  
 Ser Lys Gly Tyr Leu Leu Gly Thr Ser Glu Arg Glu Lys Ala Tyr Val  
           50                  55                  60  
 His Pro Pro Gln Trp Tyr Ala Glu His Asp Val Asp Leu Arg Leu Gly  
   65                  70                  75                  80  
 Asn Ala Val Thr Ala Leu Asp Pro Ala Gly His Glu Val Thr Leu Ala

				85					90					95		
Asp	Gly	Ser	Arg	Leu	Gly	Tyr	Ala	Lys	Leu	Leu	Leu	Ala	Thr	Gly	Ser	
			100					105					110			
Thr	Pro	Arg	Arg	Leu	Pro	Val	Pro	Gly	Ala	Asp	Leu	Asp	Gly	Val	His	
		115					120					125				
Thr	Leu	Arg	Tyr	Leu	Ala	Asp	Ser	Asp	Arg	Leu	Lys	Asp	Leu	Phe	Arg	
	130					135					140					
Ser	Ala	Ser	Arg	Ile	Val	Val	Ile	Gly	Gly	Gly	Trp	Ile	Gly	Leu	Glu	
145				150						155					160	
Thr	Thr	Ala	Ala	Ala	Arg	Ala	Ala	Gly	Val	Glu	Val	Thr	Val	Leu	Glu	
				165					170						175	
Ser	Ala	Pro	Leu	Pro	Leu	Leu	Gly	Val	Leu	Gly	Arg	Glu	Val	Ala	Gln	
			180					185					190			
Val	Phe	Ala	Asp	Leu	His	Thr	Glu	His	Gly	Val	Ala	Leu	Arg	Cys	Asp	
		195					200					205				
Thr	Gln	Val	Thr	Glu	Ile	Thr	Gly	Thr	Asn	Gly	Ala	Val	Asp	Gly	Val	
	210					215					220					
Arg	Leu	Ala	Asp	Gly	Thr	Arg	Ile	Ala	Ala	Asp	Ala	Val	Ile	Val	Gly	
225				230						235					240	
Val	Gly	Ile	Thr	Pro	Asn	Ser	Glu	Thr	Ala	Ala	Ala	Ala	Gly	Leu	Lys	
				245					250						255	
Val	Asp	Asn	Gly	Val	Val	Val	Asp	Glu	Arg	Leu	Cys	Ser	Ser	His	Pro	
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		275					280					285				
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Lys	Thr	Ala	Ala	Arg	Ala	Met	Leu	Gly	Gly	Glu	Ala	Gly	Tyr	Asp	Arg	
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Leu	Pro	Tyr	Phe	Phe	Thr	Asp	Gln	Tyr	Asp	Leu	Gly	Met	Glu	Tyr	Thr	
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Thr	Gly	Ala	Arg	Glu	Phe	Ile	Ala	Phe	Trp	Leu	Ser	Gly	Gly	Arg	Val	
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Leu	Ala	Gly	Met	Asn	Val	Asn	Val	Trp	Asp	Val	Thr	Asp	Pro	Ile	Arg	
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Ala	Leu	Val	Ala	Ser	Gly	Arg	Ala	Val	Asp	Pro	Glu	Arg	Leu	Ala	Asp	
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<211> 1266

<212> DNA

<213> Streptomyces tubercidicus

 $\langle 400 \rangle$  104

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gaccaccCGT	acgagcgccc	cccgctctcc	aaggggttcc	tgctcggcaa	ggaagagcgc	180
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&lt;210&gt; 105

&lt;211&gt; 421

&lt;212&gt; PRT

<213> *Streptomyces tubercidicus*

&lt;400&gt; 105

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 50          55          60
Val His Glu Pro Ala Trp Tyr Ala Gln Ala Gln Ile Glu Leu His Leu
 65          70          75          80
Gly Gln Pro Ala Val Arg Leu Asp Pro Glu Ala Lys Thr Val Arg Leu
 85          90          95
Gly Asp Gly Thr Leu Ile Ala Tyr Asp Lys Leu Leu Leu Ala Thr Gly
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Ala Glu Pro Arg Arg Leu Asp Ile Pro Gly Thr Gly Leu Ala Gly Val
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His His Leu Arg Arg Leu Ala His Ala Glu Arg Leu Arg Gly Val Leu
130          135          140
Ala Ser Leu Gly Arg Asp Asn Gly His Leu Val Ile Ala Gly Ala Gly
145          150          155          160
Trp Ile Gly Leu Glu Val Ala Ala Ala Ala Arg Ser Tyr Gly Ala Glu
165          170          175
Val Thr Val Val Glu Ala Ala Pro Thr Pro Leu His Gly Ile Leu Gly
180          185          190
Pro Glu Leu Gly Gly Leu Phe Thr Glu Leu His Arg Ala His Gly Val
195          200          205
Arg Phe His Phe Gly Ala Arg Phe Thr Glu Ile Val Gly Gln Asp Gly

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210		215		220
Met Val Leu Ala Val Arg Thr Asp Asp Gly Glu Glu His Pro Ala His				
225		230		235
Asp Val Leu Ala Ala Ile Gly Ala Ala Pro Arg Thr Ala Leu Ala Glu				240
	245		250	255
Gln Ala Gly Leu Asp Leu Ala Asp Pro Glu Ala Gly Gly Gly Val Ala				
	260	265	270	
Val Asp Ala Thr Leu Arg Thr Ser Asp Pro Tyr Ile Tyr Ala Ala Gly				
	275	280	285	
Asp Val Ala Ala Ala Asp His Pro Leu Leu Asp Thr Arg Leu Arg Val				
	290	295	300	
Glu His Trp Ala Asn Ala Leu Asn Gly Gly Pro Ala Ala Ala Arg Ala				
305		310	315	320
Met Leu Gly Gln Asp Ile Ser Tyr Asp Arg Val Pro Tyr Phe Phe Ser				
	325	330	335	
Asp Gln Tyr Asp Val Gly Met Glu Tyr Ser Gly Tyr Ala Pro Pro Gly				
	340	345	350	
Ser Tyr Ala Gln Val Val Cys Arg Gly Asp Val Ala Lys Arg Glu Phe				
	355	360	365	
Ile Ala Phe Trp Leu Gly Glu Asp Gly Arg Leu Leu Ala Gly Met Asn				
370	375	380		
Val Asn Val Trp Asp Val Ala Glu Thr Ile Gln Gln Leu Ile Arg Gly				
385	390	395	400	
Gly Val Arg Leu Glu Pro Gly Glu Leu Ala Asp Pro Glu Val Pro Leu				
	405	410	415	
Thr Ser Leu Leu Pro				
	420			

**BUDAPEST TREATY ON THE INTERNATIONAL RECOGNITION OF THE DEPOSIT  
OF MICROORGANISMS FOR THE PURPOSE OF PATENT PROCEDURES**

**INTERNATIONAL FORM**

TO

Syngenta  
3054 Cornwallis Road  
Research Triangle Park, NC 27709

**VIABILITY STATEMENT**

issued pursuant to Rule 10.2 by the  
INTERNATIONAL DEPOSITARY AUTHORITY  
identified at the bottom of this page

NAME AND ADDRESS OF THE PARTY TO WHOM  
THE VIABILITY STATEMENT IS ISSUED

<b>I. DEPOSITOR</b>  Name: Syngenta 3054 Cornwallis Road Address: Research Triangle Park, NC 27709	<b>II. IDENTIFICATION OF THE MICROORGANISM</b>  Depositor's taxonomic designation and accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: <i>Streptomyces lividans</i> NRRL 30478  Date of: May 8, 2001 <input checked="" type="checkbox"/> <sup>2</sup> Original Deposit <input type="checkbox"/> <sup>2</sup> New Deposit <input type="checkbox"/> <sup>2</sup> Repropagation of Original Deposit		
<b>III. (a) VIABILITY STATEMENT</b>  Deposit was found: <input checked="" type="checkbox"/> Viable <input type="checkbox"/> Nonviable on May 10, 2001 (Date) International Depositary Authority's preparation was found viable on May 18, 2001 (Date) <sup>3</sup>			
<b>III. (b) DEPOSITOR'S EQUIVALENCY DECLARATION</b>  Depositor determined the International Depositary Authority's preparation was <input checked="" type="checkbox"/> <sup>2</sup> Equivalent <input type="checkbox"/> <sup>2</sup> Not equivalent to deposit on <u>7/23/01</u> (Date) Signature of Depositor <u>[Signature]</u>			
<b>IV. CONDITIONS UNDER WHICH THE VIABILITY TEST WAS PERFORMED (Depositor/Depository)<sup>4</sup></b>  			
<b>V. INTERNATIONAL DEPOSITARY AUTHORITY</b>  <table border="1"> <tr> <td data-bbox="121 1564 771 1732">           Name: Agricultural Research Culture            Collection (NRRL)            International Depositary Authority             Address: 1815 N. University Street            Peoria, Illinois 61604 U.S.A.         </td> <td data-bbox="771 1564 1485 1732">           Signature(s) of person(s) having the power            to represent the International Depositary            Authority or of authorized official(s):  <u>[Signature]</u>            Date: <u>6-26-01</u> </td> </tr> </table>		Name: Agricultural Research Culture Collection (NRRL) International Depositary Authority  Address: 1815 N. University Street Peoria, Illinois 61604 U.S.A.	Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s): <u>[Signature]</u> Date: <u>6-26-01</u>
Name: Agricultural Research Culture Collection (NRRL) International Depositary Authority  Address: 1815 N. University Street Peoria, Illinois 61604 U.S.A.	Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s): <u>[Signature]</u> Date: <u>6-26-01</u>		

<sup>1</sup> Indicate the date of the original deposit or when a new deposit has been made.

<sup>2</sup> Mark with a cross the applicable box.

<sup>3</sup> In the cases referred to in Rule 10.2(a)(ii) and (iii), refer to the most recent viability test.

<sup>4</sup> Fill in if the information has been requested.

BUDAPEST TREATY ON THE INTERNATIONAL RECOGNITION OF THE DEPOSIT  
OF MICROORGANISMS FOR THE PURPOSE OF PATENT PROCEDURES

INTERNATIONAL FORM

TO

Syngenta  
3054 Cornwallis Road  
Research Triangle Park, NC 27709

VIABILITY STATEMENT

Issued pursuant to Rule 10.2 by the  
INTERNATIONAL DEPOSITARY AUTHORITY  
identified at the bottom of this page

NAME AND ADDRESS OF THE PARTY TO WHOM  
THE VIABILITY STATEMENT IS ISSUED

<p><b>I. DEPOSITOR</b></p> <p>Name: Syngenta 3054 Cornwallis Road Address: Research Triangle Park, NC 27709</p>	<p><b>II. IDENTIFICATION OF THE MICROORGANISM</b></p> <p>Depositor's taxonomic designation and accession number given by the INTERNATIONAL DEPOSITARY AUTHORITY: <i>Pseudomonas putida</i> NRRL B-30479</p> <p>Date of: May 8, 2001  <input checked="" type="checkbox"/> <sup>2</sup> Original Deposit  <input type="checkbox"/> <sup>2</sup> New Deposit  <input type="checkbox"/> <sup>2</sup> Repropagation of Original Deposit</p>		
<p><b>III. (a) VIABILITY STATEMENT</b></p> <p>Deposit was found: <input checked="" type="checkbox"/> Viable <input type="checkbox"/> Nonviable on May 10, 2001 (Date)</p> <p>International Depositary Authority's preparation was found viable on May 16 2001 (Date)<sup>3</sup></p>			
<p><b>III. (b) DEPOSITOR'S EQUIVALENCY DECLARATION</b></p> <p>Depositor determined the International Depositary Authority's preparation was</p> <p><input checked="" type="checkbox"/> <sup>2</sup> Equivalent <input type="checkbox"/> <sup>2</sup> Not equivalent to deposit on <u>7/23/01</u> (Date)</p> <p>Signature of Depositor <u>[Signature]</u></p>			
<p><b>IV. CONDITIONS UNDER WHICH THE VIABILITY TEST WAS PERFORMED (Depositor's/Depository)<sup>4</sup></b></p>			
<p><b>V. INTERNATIONAL DEPOSITARY AUTHORITY</b></p> <table border="1"> <tr> <td data-bbox="136 1541 789 1715"> <p>Name: Agricultural Research Culture Collection (NRRL) International Depositary Authority</p> <p>Address: 1815 N. University Street Peoria, Illinois 61604 U.S.A.</p> </td> <td data-bbox="789 1541 1503 1715"> <p>Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):</p> <p><u>[Signature]</u> Date: <u>6-26-01</u></p> </td> </tr> </table>		<p>Name: Agricultural Research Culture Collection (NRRL) International Depositary Authority</p> <p>Address: 1815 N. University Street Peoria, Illinois 61604 U.S.A.</p>	<p>Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):</p> <p><u>[Signature]</u> Date: <u>6-26-01</u></p>
<p>Name: Agricultural Research Culture Collection (NRRL) International Depositary Authority</p> <p>Address: 1815 N. University Street Peoria, Illinois 61604 U.S.A.</p>	<p>Signature(s) of person(s) having the power to represent the International Depositary Authority or of authorized official(s):</p> <p><u>[Signature]</u> Date: <u>6-26-01</u></p>		

<sup>1</sup> Indicate the date of the original deposit or when a new deposit has been made.

<sup>2</sup> Mark with a cross the applicable box.

<sup>3</sup> In the cases referred to in Rule 10.2(a)(ii) and (iii), refer to the most recent viability test.

<sup>4</sup> Fill in if the information has been requested.